

# Technological University Dublin [ARROW@TU Dublin](https://arrow.tudublin.ie/)

[Articles](https://arrow.tudublin.ie/schfsehart) **School of Food Science and Environmental Health** 

2020

# Adaptive Background Correction of Crystal Image Datasets: **Towards Automated Process Control**

Luke Kiernan Innopharma Technology, Dublin

Ian Jones Innopharma Technology, Dublin

Lauri Kurki VTT Technical Research Center of Finland

See next page for additional authors

Follow this and additional works at: [https://arrow.tudublin.ie/schfsehart](https://arrow.tudublin.ie/schfsehart?utm_source=arrow.tudublin.ie%2Fschfsehart%2F416&utm_medium=PDF&utm_campaign=PDFCoverPages) 

Part of the [Environmental Health and Protection Commons,](https://network.bepress.com/hgg/discipline/172?utm_source=arrow.tudublin.ie%2Fschfsehart%2F416&utm_medium=PDF&utm_campaign=PDFCoverPages) [Investigative Techniques Commons](https://network.bepress.com/hgg/discipline/922?utm_source=arrow.tudublin.ie%2Fschfsehart%2F416&utm_medium=PDF&utm_campaign=PDFCoverPages), and the [Other Analytical, Diagnostic and Therapeutic Techniques and Equipment Commons](https://network.bepress.com/hgg/discipline/994?utm_source=arrow.tudublin.ie%2Fschfsehart%2F416&utm_medium=PDF&utm_campaign=PDFCoverPages)

# Recommended Citation

Kiernan, L., Jones, I., Kurki, L. et al. Adaptive Background Correction of Crystal Image Datasets: Towards Automated Process Control. Sens Imaging 21, 48 (2020). DOI: 10.1007/s11220-020-00310-6

This Article is brought to you for free and open access by the School of Food Science and Environmental Health at ARROW@TU Dublin. It has been accepted for inclusion in Articles by an authorized administrator of ARROW@TU Dublin. For more information, please contact [arrow.admin@tudublin.ie, aisling.coyne@tudublin.ie,](mailto:arrow.admin@tudublin.ie,%20aisling.coyne@tudublin.ie,%20vera.kilshaw@tudublin.ie)  [vera.kilshaw@tudublin.ie](mailto:arrow.admin@tudublin.ie,%20aisling.coyne@tudublin.ie,%20vera.kilshaw@tudublin.ie).

Funder: EC's Seventh Framework; Science Foundation Ireland

# Authors

Luke Kiernan, Ian Jones, Lauri Kurki, Patrick J. Cullen, and Toufic El Arnaout

**ORIGINAL PAPER**



# **Adaptive Background Correction of Crystal Image Datasets[:](http://crossmark.crossref.org/dialog/?doi=10.1007/s11220-020-00310-6&domain=pdf)  Towards Automated Process Control**

Luke Kiernan<sup>1</sup> · Ian Jones<sup>1</sup> · Lauri Kurki<sup>2,3</sup> · Patrick J. Cullen<sup>4</sup> · **Toufc El Arnaout5,[6](http://orcid.org/0000-0003-2570-9668)**

Received: 21 June 2020 / Revised: 3 August 2020 / Accepted: 12 September 2020 / Published online: 3 October 2020 © Springer Science+Business Media, LLC, part of Springer Nature 2020

#### **Abstract**

Improving the data descriptor calculation of crystal's physical properties requires sophisticated imaging techniques and algorithms. It has been possible to construct 2D population balance models benefting from characteristic measurements of both crystal's length and width, compared to the single representative sizes used in 1D models. Our aim is to ameliorate the procedure of determining shape (and not only size) factors, in an automated fashion and directly from the process, for implementation in future models. Here, approaches suitable for real-time applications were employed including engineered imaging sensors and adaptive algorithms. We described the latter in detail for varying 2D image datasets. Their basic concept is similar. Each is applicable to an entire dataset, thus demonstrating efficacy for a variety of particle environments. While the challenge of particle segmentation for higher concentrations was not scrutinized here, this approach reduced processing time, steps and supervision, for the beneft of certain applications requiring process monitoring and automation.

**Electronic supplementary material** The online version of this article ([https://doi.org/10.1007/s1122](https://doi.org/10.1007/s11220-020-00310-6) [0-020-00310-6](https://doi.org/10.1007/s11220-020-00310-6)) contains supplementary material, which is available to authorized users.

 $\boxtimes$  Toufic El Arnaout toufc.arnaout@kappacrystals.com

- <sup>1</sup> Innopharma Technology, Sandyford, Dublin, Ireland
- <sup>2</sup> Timegate Instruments, 90590 Oulu, Finland
- <sup>3</sup> VTT Technical Research Center of Finland, 90570 Oulu, Finland
- <sup>4</sup> School of Chemical and Biomolecular Engineering, The University of Sydney, Darlington, NSW 2008, Australia
- <sup>5</sup> School of Food Science and Environmental Health, TU Dublin City Campus, Technological University Dublin, Dublin, Ireland
- <sup>6</sup> Kappa Crystals Ltd, Dublin, Ireland

#### **Graphic Abstract**



**Keywords** Crystallization imaging · Adaptive background correction · Particle engineering · Analytical technology

# **1 Introduction**

#### **1.1 Crystallization Modelling**

Several spectroscopic, laser and imaging methods now permit access to information in real-time and directly from the reaction (in situ) due to developments in Process Analytical Technology (PAT) [[1,](#page-19-0) [2\]](#page-19-1). This information may be crystal size distributions (CSD) and particle size-shape distribution (PSSD), and many physico-chemical properties, found to infuence population balance equations (PBEs) and models (PBMs) [[3\]](#page-19-2). 2D models supported by size and shape information, mainly possible using imaging, have large advantages over basic 1D models and less assumptions.

One of the main approaches employed to measure 1D information has been based on the chord length, represented by the scanned distance, in a random part across a particle. The method is highly rapid and efficient for a wide range of applications and concentrations. The CSD may be calculated based on necessary transformations from the chord length distribution (CLD). The chord length is a random scan of a particle that may result in diferent values for the same particle, according to its shape, rotation, physical properties, scan direction, as well as the optical properties

of the system [\[4](#page-19-3)] (Table [1\)](#page-5-0). More recently, improved models were produced to support the calculation of 1D CSD through an optimized relationship between CLD, geometry and size [[5\]](#page-19-4), along with considerations of sample properties, laser intensity, refractive indices, crystal velocity, and mathematical optimizations based on the working principles of the laser beam in operation. As for the 2D CSD, additional parameters to the CLD are necessary [\[6](#page-19-5)], and this can be supported by using optical imaging.

Video microscopy permits not only the determination of length and width, but the shape (in 2D), including symmetry, convexity/smoothness, and circularity. Some challenges can be the performance, algorithms for image processing and understanding, sensitivity (e.g., based on resolution, refractive index, particle properties), and particle concentration. Yet, developments in microscopy have been signifcant and for a wide range of applications, due to an amalgamation of lenses, prisms, illumination sources, cameras, and many engineering and electronic approaches. Image and data analysis methods have also evolved [\[15](#page-19-6), [16](#page-19-7)]. Therefore, imaging-based PAT has become more suitable to support monitoring, continuous crystallization and screening platforms [\[17](#page-20-0), [18](#page-20-1)].

#### **1.2 Image Processing: Developments and Challenges**

An image processing workfow typically applies a series of prebuilt equations and calculations on information obtainable and accessible in the image. Several functions and plugins are available with programs like ImageJ, Rgui, Matlab and OpenCV. Certain algorithms may provide limited prediction, such as using an autocomplete function for partially complete features, sometimes with matching with a database, part of a large decision tree of sophisticated steps and loops. However, the 'interpretation' of complex images remains a challenge for machine vision as demonstrated by the inability to solve a Captcha for example, which requires human intervention.

Basic edge detection and watershed operations (e.g., Canny edge detection, Trainable Weka Segmentation (TWS)) also require signifcant computing times, training, use of templates [[19\]](#page-20-2), and/or iterative procedures. Employing a series of assumptions and line assignments (model-based) has been applied for in situ images of α-glycine crystals [\[20](#page-20-3)]. The technique was based on linear features, VIGs (Viewpoint-invariant groups) as properties of an object that are maintained irrespective of the camera's position, and model ftting. A method for particle recognition [[21\]](#page-20-4), in particular overlapped particles that are simple-shaped (e.g., rectangles) was demonstrated and used time-zero background image subtraction, multi-scale edge detection, fltering, and salient corners (intersection of two lines). More recently, the detection of simple-shaped (needles), overlapped crystals was also demonstrated for in situ applications  $[22]$  $[22]$ . A few studies commented on the difficulty with more complex crystal scenes including the presence of several shapes (especially not preknown), unfocused objects, high particle concentration, semi-transparent particles, and strong particle attrition. Overall, it is frequent to identify in the literature studies employing methods in parallel with microscopy, such as the CLD [[14\]](#page-19-8).

<span id="page-5-0"></span>Table 1 A list of challenges encountered with CLD-based PAT methods **Table 1** A list of challenges encountered with CLD-based PAT methods



#### <span id="page-6-1"></span>**1.3 Adaptive Background Correction**

Background correction is a major task for in situ image processing for the purpose of quantitative analysis of size-shape. Most often, users subtract time-zero blank images for correction. However, typical backgrounds from in situ images are not identical throughout an entire dataset, coupled with varying noise distributions, shadows, and inhomogeneities (Figs. [1,](#page-6-0) bottom and [3\)](#page-10-0). Basic median flter to subtract standard backgrounds may eliminate noise and non-homogeneity [[23\]](#page-20-6), but with rejection of particles smaller than 24  $\mu$ m (30×30 pixels<sup>2</sup>). Moreover, several methods of data transformation exist (e.g., linear, convolution, Gaussian, smoothing, Fourier transform). Fortunately, an adaptive approach adopted here permits normalization of such backgrounds [\[24](#page-20-7)] while also smoothing the image. In this paper, the technique employed is compatible with gradients, observable in certain relief contrast methods (Fig. [1](#page-6-0), bottom) used to observe texture particularly for small and thin entities.



<span id="page-6-0"></span>**Fig. 1** Classic challenges of processing images of particles such as crystals. Top: Crystals are optically active causing difficulty compared to opaque particles. Bottom: Challenges with optical engineering setups like prism intensity and gradient. These are two examples of limitations for classic algorithms that follow a sequence of calculations, unable to resolve visually overlapped particles versus agglomerates, watershed/segmentation, out-of-focus particles, gradients of lights, and high concentrations

Visibility conditions in general impact on many measurement technologies [[25\]](#page-20-8). The signal to noise and signal to background ratios (can be due to the imaging approach and/or the camera sensor [[26\]](#page-20-9)), illumination, gradients, and color variations, are important to optimize. Furthermore, in situ imaging challenges (Table [2](#page-8-0)) are a focus of the macro algorithms developed here.

The Rolling Ball (RB) algorithm [\[24](#page-20-7)] is an adaptive method for background correction, independently of time-zero/blank images or of one image to another. With the optimized sliding paraboloid (SP) approach it has been applied in the biomedical, cell biology, geophysical, and materials science sectors  $[31–33]$  $[31–33]$  $[31–33]$ . It is sufficiently fast for real-time analysis and suitable for non-uniform backgrounds (e.g., illumination, intensity, brightness, gradient). The code was successfully implemented initially in the NIH Image Pascal, which has since been superseded by ImageJ. The term "ball" describes the correcting shape that passes ('rolls') over the bottom surface of the image. It has a certain limit in reaching inside the peaks, depending on the sizes of the correction ball and the peaks. Nearly a decade ago, Michael Schmid released a variant of this algorithm, more suitable for certain images in terms of intensities and shapes, known as the "Sliding Paraboloid" of approximation (same curvature at the apex as the ball of a given radius) (Fig. [2\)](#page-10-1). To be more precise, this is sometimes described as "Sliding Parabolae" in four directions  $(x, y, z) \geq 45^{\circ}$ , for practicality and speed reasons. The code was also optimized to correctly process objects in image corners. The parabola (or paraboloid) has a diferent symmetry and shape than a circle (or ball), hence the term 'sliding' rather than 'rolling' when applied. Smoothing and correction are calculated via approximate values also depending on the surrounding local average, using a pre-specifed radius. A "Separate colors" option permits for RGB images to correct not only based on brightness but also on hue and saturation, which are strong visual appearance properties [[34\]](#page-20-12). This possibility was key in the success of certain studies shown later, when color information was necessary from the start (Fig. [2](#page-10-1)), for in situ images containing spatial variations, blurriness, out of focus and in focus objects, shadows, and noise. It was also useful when applying an Enhanced local contrast (CLAHE) or a general 'Enhance contrast' function, via adjustments of their settings such as Blocksize, Histogram pins, Maximum slope, Mask, Histogram equalization, and Saturation.

In this study, we have listed the macros of algorithms in detail and for the respective datasets. Furthermore, the development was also explained step by step, and for several challenging experiments using explanatory fgures. The study shows the main advantage of the adaptive background correction procedure employed here and the potential application opportunities to monitor processes containing particles such as cells, emulsions, bubbles, crystals, and particles in general.

#### **2 Methods**

#### **2.1 Imaging**

The imaging experiment of thiamine hydrochloride shown in Figs. [3](#page-10-0) and [4](#page-11-0) was carried out based on a benchtop optical system [\[27](#page-20-13)] under weak polarization, with a

<span id="page-8-0"></span>







<span id="page-10-1"></span>**Fig. 2** The importance of continuously improving adaptive background correction algorithms. A method comparison is shown based on 3D surface plots (original raw image: bottom left) between the rolling ball (**a**) and sliding paraboloid (**b**) correction functions



<span id="page-10-0"></span>**Fig. 3** A simple procedure for enhancing in-line image processing of particles. Comparison of a classic approach (raw image, top left) with an optimized one (treated image, bottom left). The resulting binary images are shown to the right

camera resolution of  $2592 \times 1944$  pixels<sup>2</sup> and a field of view of 1.25 mm $\times$ 1 mm. Crystals were imaged as semi-opaque in low light bright feld. Other experiments were imaged in bright feld, moderate or strong relief contrast, on a slide (in-line) or in situ, as indicated, based on the sensor probe systems developed [\[27](#page-20-13), [35\]](#page-20-17) with



**Fig. 4** A semi-adaptive approach for in-line particle image datasets. Examples are chosen from a single run whereby a single macro was applied to correct backgrounds, noise, large variations (bubbles, shadows), while preserving the signals of microparticles

<span id="page-11-0"></span>a 10X objective and a camera sensor of  $3376 \times 2704$  pixels<sup>2</sup>. More specifically: (Sect. [3.1](#page-12-0)) thiamine hydrochloride crystals were imaged in a microfuidic slide with a pump system; (Sect. [3.2\)](#page-12-1) lysozyme crystals were imaged statically in a large drop on a slide in bright feld mode; (Sect. [3.3](#page-15-0)) particle size standards, l-glutamic acid crystals, and taurine crystals were imaged in situ with full probe immersion in relief contrast mode; fnally (Sect. [3.4](#page-16-0)) taurine crystals were imaged in a large drop on a slide in relief contrast mode.

#### **2.2 Crystallization Protocols**

Thiamine hydrochloride crystals were used from a stored slurry with solvent, produced by re-crystallization.

Lysozyme was selected for the transparent crystal imaging studies with bright feld mode. Lysozyme powder was frst dissolved at 50 mg/mL in 0.1 M sodium acetate pH 4.5. 1 µL was then added onto a glass slide with 1 µL of precipitant  $(30\%$ w/v MPEG 5000, 1 M sodium chloride and 50 mM sodium acetate pH 4.5). Additional microliters may also be used for diferent protein:precipitant ratios (e.g., 1:2, 1:3, 2:1). Evaporation started as the drop was visualized and crystals began forming while recording an image every 4 s.

To obtain the images shown in Fig. [7,](#page-15-1) particle standards of  $15-150 \mu m$  glass beads (Malvern, Cat. QAS3002) were added to  $H_2O$  at 1% w/v and imaged during stirring. As for l-glutamic acid crystallization, it was carried out by dissolving powder at 3.5% w/v into H<sub>2</sub>O at 65 °C, with a temperature decrease to 47 °C at a rate of 0.14 °C/min. For taurine crystallization, 100 g was added into 700 mL  $H_2O$  at 42 °C, and crystallization from supersaturation occurred over 2 h.

The last example, based on in-line imaging with a strong gradient, was carried out by adding taurine powder to 10 mL of  $H_2O$  at 95 °C until supersaturation was reached. A drop of a few μL was pipetted from the supernatant onto a microscope slide. Crystallogenesis began immediately due to evaporation and temperature decrease.

# **2.3 Software**

Images were automatically captured using certain settings and procedures [[16,](#page-19-7) [27,](#page-20-13) [35](#page-20-17)] during long experiments, or manually saved. For processing, ImageJ Fiji [\[36](#page-21-0)] was used for batch processing and applying the series of functions described under "Macros" below and in full detail in the Online Appendix section. For the background removal in ImageJ through the rolling ball/sliding paraboloid function, the "Separate color" function is a visible option in older versions (e.g., v. 2014 Jun 02 or v. 2014 Nov 25) possibly compared to the more recent versions some of which were heavily transitioned to Java (e.g., v. 2017 May 30 or v. 2015 Dec 22). After binary conversion and export of the particle size-shape data, Microsoft Excel was used for binning and graph generation (Fig. [6\)](#page-14-0).

# **2.4 Macros**

The image background processing and particle analysis functions applied in each experiment are shown in the Online Appendix part, Section (a). These macros (A to D) describe the procedure for each case based on the optical setting used with the crystal images they were applied on: (A) thiamine hydrochloride (Figs. [3](#page-10-0), [4](#page-11-0)),  $(B)$ lysozyme (Fig. [5](#page-13-0)), (C) polydisperse particles, l-glutamic acid, and taurine (Fig. [7\)](#page-15-1), and  $(D)$  taurine (Fig. [9\)](#page-17-0).

# **3 Results and Discussion**

# <span id="page-12-0"></span>**3.1 In‑Line Bright Field Imaging of Semi‑opaque Particles**

Thiamine hydrochloride crystals were observed at moderate concentrations in a lowlight setting. The particles are semi-opaque in this opto-illumination setup (Fig. [1\)](#page-6-0), with a strong image noise (Fig. A.1). The general strategy succeeded by following the sequence: an adaptive background subtraction (rolling ball, light background, color separation), a CLAHE contrast algorithm, smoothing, Gaussian blur smoothing, contrast enhancement, 8 bit conversion, specifed thresholding (according to the brightness/contrast levels, and pixel intensities), and general binary/outlier removal/ flling holes tasks. This was found to be more adequate for particle extraction and analysis compared to applying a threshold directly (Fig. [3](#page-10-0) and Fig. A.2), to overcome the image to image variations in noise, shadows, intensities, and the presence of microcrystals or large obstructions (e.g., bubbles) (Fig. [4](#page-11-0)).

# <span id="page-12-1"></span>**3.2 In‑Line Bright Field Imaging of Transparent Crystals**

Crystals are optically active due to their inherent properties and the illumination and optical imaging setup. They may appear transparent with their inner parts displaying similar intensities and color to those of the background (Fig. [5](#page-13-0)b) during



<span id="page-13-0"></span>**Fig. 5** Double background correction of static images of transparent crystals. Lysozyme crystals were grown by vapor difusion. **a** The background at time zero (blank image), **a′**: the image in 'a' following correction with the rolling ball flter, **b** the crystal image (raw), **b′** the image in '**b**' after correction with the rolling ball flter, **c** thresholded and binary image of the **a′** and **b′** "Diference" calculation, **d** the nonoptimized fnal result if the second background correction operation (with the time-zero background) was not performed

growth, as observed with benzoic acid  $[27]$  $[27]$ . For binary signal recovery in situations like these, a double background correction approach was found to be useful for transparent lysozyme crystals (Fig. [5c](#page-13-0)) by employing the adaptive rolling ball correction, but also a time-zero (blank image) background subtraction. This was compared against the situation whereby this double subtraction approach was not incorporated (Fig. [5](#page-13-0)). Therefore, the general procedure was to carry out a rolling ball correction of the image of interest (image 1), then, using the time-zero blank image (also rolling ball pre-corrected) (image 0), both images (0 and 1) were subtracted from each other using a 'diference create' calculation. The resulting image is then converted into 32-bit format, thresholded with specifc parameters



<span id="page-14-0"></span>**Fig. 6** Size-shape tracking of lysozyme crystals by image analysis. Average per image for all particles is shown over 800 + images. Top: average size (feret)  $(1 \text{ A.U.}=0.38 \text{ µm})$  and total counts per image. Bottom: average circularity and solidity (shape descriptors) per image as a factor (min=0 max = 1)

and transformed into binary, before applying the usual steps of outlier correction, flling holes, smoothing, and basic watershed.

This processing approach was useful for tracking size-shape changes over 800 images (Fig. [6\)](#page-14-0). During the initial stages (frst 150 images), crystals grew rapidly while their positions were slowly changing (movement) before reasonably settling in the fxed optical feld of view. Here, the particle analysis procedure excludes particles touching the image borders. Larger crystals were changing slightly in positions during imaging and subsequently became wholly captured within the image borders. Microcrystals decreased in counts, therefore infuencing on the overall size average (jump of  $3-4 \mu$ m) and counts (drop of  $14\%$ ) (Fig. [6](#page-14-0), top). Furthermore, due to the resolution requirement and optical parameters in this experiment, operating at a narrow focal plane range of<20 μm causes certain particles to be out-of-focus. and thus strong vibrations or changes of the focal plane within the static drop may alter the processing outcome for the image recorded at that time (e.g., small peaks in images 214 and 345). Overall, rapid changes in the observed physical characteristics were most signifcant during the frst 100 images (Fig. [6,](#page-14-0) bottom), with circularity decreasing, and shapes becoming more defned. The average size also increased until frame #215, with the total count increasing gradually after frame #460 with some infuence on the size average. Therefore, this image processing has advantages to track the size at a micrometer



**Fig. 7** Imaging particles in situ using a video microscopy probe equipped with a relief contrast imaging mode. The gradient is caused by the prism properties which are benefcial in certain applications to visualize a stronger contrast/3D-like appearance particularly for imaging small or thin features. **a** Particle size standards (15–150 μm), **b** <sup>l</sup>-Glutamic acid crystals, **c** taurine crystals. Both raw (top) and processed (bottom, binary) images are presented (rotated here 90°)

<span id="page-15-1"></span>resolution, and not only shape. Nevertheless, general challenges of the imaging and processing approach employed in this paper are listed in Table [2](#page-8-0).

#### <span id="page-15-0"></span>**3.3 In Situ Relief Contrast Imaging with a Gradient**

A stronger detection of particle characteristics is desired in advanced applications aiming at texture studies or for the enhancement of weakly visible features. This may be possible via relief contrast methods including the classic DIC or more recent ones such as PlasDIC (Polarization optical diferential interference contrast, or Plastic DIC) [\[37](#page-21-1)]. In the latter, components in the optical assembly may be reduced compared to those in DIC, while providing improved imaging fexibility and a certain compatibility with anisotropic materials [\[38](#page-21-2)]. However, illumination recovery is not as strong as in the bright feld mode. In low noise images the user may be able to detect the natural diference between (or within) the object, and its environment in terms of the refractive index. In DIC/PlasDIC, the optical settings, including those of the main prism (and its lateral translation), may result in a gradient in the captured image (Figs. [2](#page-10-1) and [7\)](#page-15-1). In our setup, this was signifcant when a strong contrast was required to enhance the observation of thin/transparent particles (Fig. [9\)](#page-17-0) (via illumination and/or prism adjustments). Equally the gradient efect was especially signifcant during in situ imaging mode (Fig. [7](#page-15-1)). Color information also appeared to be afected and contained signals that are important to recover successfully.

To correct the background in this case, the sliding paraboloid (based on the rolling ball) method was employed. In particular, it has permitted another "color separation" option (refer to Sect. [1.3](#page-6-1) for defnition). For this application and assuming that the background is 'light' (bright mode) this approach was sufficient to process the images. The macro of the in situ image processing frst and foremost started with a general, classic contrast enhancement based on a specifed saturation level, with histogram equalization [[39\]](#page-21-3). Combined with the relief contrast imaging approach, this proves the importance of appropriate contrast levels. Next, the background was subtracted using the sliding paraboloid method (as in Fig. [2b](#page-10-1)), in light mode, with the color separation option. The image was then converted to binary, after which general operations were applied such as outlier removal, dilation and flling holes. This procedure permitted to obtain a balance between signal recovery and noise reduction. Yet, it was more favorable with the colored, contrast enhanced images (although these take longer to process than greyscale images), than with images converted to 32 bit greyscale. Therefore, raw RGB images were necessary, to maintain the information associated with object completeness contained in color channels.

The images obtained in relief contrast mode contained a gradient (Figs. [7](#page-15-1) and [9](#page-17-0)). In one side the taint was dark, which made the noise streaks more prominent, particularly at increasing crystal concentrations. This led to difficulties in extracting complete information. Nevertheless, examples shown here demonstrated that it is possible to extract signals into binary (Fig. [7](#page-15-1)), while the shape information is maintained. This supports the shape tracking in many applications, such as downstream processing optimization. However, the concentration challenge not only impacts on imaging (Table [2\)](#page-8-0), but often on equipment, leading in some cases to the formation of a crust on the body of the probe. Yet, this happened more at the surface of the solution than inside the imaging gap, and during strong decrease from high temperatures (Fig.  $8$ ).

#### <span id="page-16-0"></span>**3.4 In‑Line Relief Contrast Imaging at a Strong Prism Setting**

Static drops containing transparent crystals in a single plane were used. In this experiment, it is possible to increase the relief contrast intensity via adjustment of the prism's lateral translation on top of the sample. The corresponding images exhibit a strongly visible light and contrast gradient efect. In some instances, the visualization with a strong contrast may permit the early detection of shape changes and very thin and small particles.

Taurine crystals were imaged and a macro of a double step 'light+dark' signal recovery was applied for binary extraction of signals for particle analysis purposes (Online Appendix, Algorithm D). A graphical representation of these steps is shown in Fig. [9.](#page-17-0) The raw image (left) was duplicated and then each copy was processed separately (as 'light' or 'dark'). Binary information was recovered (in red, middle) for each light or dark part and both outputs were then merged into one fnal image (right).



**Fig. 8** Visible incrustation on an immersible probe during crystallization. The experiment included gradual decrease of temperature during which there was continuous stirring in an open container and a normally occurring evaporation. A 'crust' due to the dissolved chemical formed on the stainless steel body

<span id="page-17-1"></span>

<span id="page-17-0"></span>**Fig. 9** Extracting transparent crystal signals from images with a strong gradient and light variation. Original image (left) is duplicated and each copy is treated separately with an adaptive background correction algorithm based on light (bright) or dark information, and then thresholded (middle). The two resulting images are then merged by adding them together (right). The cyan and red colors (right) are to show the two groups of signals recovered in the previous step (Color fgure online)

In this experiment, each copy was converted to 32 bit prior to full processing. This procedure was successful for these images which contained low noise, thinner sample, and from a static drop. One copy had the background subtracted with the rolling ball, light mode, while the second copy had the correction with the rolling ball, dark mode. Thresholding for each copy was then carried out with specifc threshold values (as indicated in the algorithm), optimized based on the pixel intensity thresholds, and which work for the entire dataset. Both processed copies

were then converted to binary, merged together through the "Add" operation, and saved. Particle analysis and statistics were then carried out following general operations like smoothing, flling holes, removing outliers, dilation, etc. As previously explained, in most setups, including this one, background subtraction of time-zero image was not applied, or not beneficial. It is not uncommon to rely on the blank background [[21,](#page-20-4) [40](#page-21-4)], but it is not ideal in automated applications as the image backgrounds during the reaction difer from the time-zero background (particularly with in situ imaging and in the presence of an increasing particle concentration), due to light intensity changes, random particle positions, inhomogeneity and refractions, obstructions of light, etc.

#### **4 Conclusion**

An image processing workfow designated for a specifc application does not randomly apply to another one. Such a workfow may also not be applicable throughout the same, entire dataset for all images. However, the background correction approach employed here was successful and time-efficient for real-time implementation (e.g., 10–30 s per image, with the possibility for multithreading images in parallel). It was possible to integrate and customize it in several procedures for different datasets. Example datasets contained varying levels of noise or distributions of pixels on diferent background types. These backgrounds also displayed varying gradients and intensities of brightness and colors, and with several particle transparency levels. Therefore, challenges such as noise and shadows, while maintaining the signals of microparticles were overcome. Examples presented were for (a) dark images, (b) images of transparent crystals, (c) in situ images, and (d) images of thin/ transparent crystals in the presence of a strong contrast gradient.

Size-shape data descriptors were generated and their evolution between images was tracked. An additional advantage over 1D/spectroscopic/laser methods is that the reliability can be further verifed by manually checking the raw images by direct observation. These descriptors do not only concern width and length (aspect ratio), as over 65% of APIs may have a median aspect ratio of 0.6–0.8 [[41\]](#page-21-5), but also further geometrical characteristics which are discussed including circularity and solidity (in 2D). These data descriptors will be important for the advanced modelling, change tracking, and polymorph research. Finally the work is likely to support research related to co-crystallization and/or intensifed downstream processing to improve crystal characteristics with impact on efficacy, quality, safety, dissolution and bioavailability of a product [\[42](#page-21-6)[–44](#page-21-7)].

The steps presented in this article may be also employed in the future with object classifcations, decision nodes, and clustering techniques, and implemented into a large neural network [[45–](#page-21-8)[47\]](#page-21-9), to minimize supervision and improve prediction. The major challenge to overcome remains particle segmentation. This requires advanced image understanding, recognition and continuous training. Yet, it may be case-specifc when lacking the qualitative and predictive human's brain capacities necessary to solve a Captcha as a previously mentioned example.

**Acknowledgements** The research leading to these results has received funding from the European Community's Seventh Framework Program (FP7-SME-2013) under the CRYSTAL-VIS project, Grant Agreement Number 605814, and from Science Foundation Ireland (SFI) through a Technology Innovation Development Award (TIDA). The authors would like to thank the PAT group (TU Dublin, Ireland), VTT (Oulu, Finland), and Topchem (Ireland).

### **References**

- <span id="page-19-0"></span>1. Gouveia, F. F., Rahbek, J. P., Mortensen, A. R., Pedersen, M. T., Felizardo, P. M., Bro, R., et al. (2017). Using PAT to accelerate the transition to continuous API manufacturing. *Analytical and Bioanalytical Chemistry, 409*(3), 821–832. [https://doi.org/10.1007/s00216-016-9834-z.](https://doi.org/10.1007/s00216-016-9834-z)
- <span id="page-19-1"></span>2. Reid, L. G., Ward, W. H., Palm, A. S., & Muteki, K. (2012). Process analytical technology (PAT) in pharmaceutical development. *American Pharmaceutical Review, 15*.
- <span id="page-19-2"></span>3. Chen, S., Liu, T., Xu, D., Huo, Y., & Yang, Y. (2019). Image based measurement of population growth rate for l-glutamic acid crystallization. In *Chinese control conference (CCC)* (pp. 7933– 7938). [https://doi.org/10.23919/chicc.2019.8866441.](https://doi.org/10.23919/chicc.2019.8866441)
- <span id="page-19-3"></span>4. Ruf, A., Worlitschek, J., & Mazzotti, M. (2000). Modeling and experimental analysis of PSD measurements through FBRM. *Particle & Particle Systems Characterization, 17*(4), 167–179. [https://doi.](https://doi.org/10.1002/1521-4117(200012)17:4%3c167:AID-PPSC167%3e3.0.CO;2-T) [org/10.1002/1521-4117\(200012\)17:4%3c167:AID-PPSC167%3e3.0.CO;2-T.](https://doi.org/10.1002/1521-4117(200012)17:4%3c167:AID-PPSC167%3e3.0.CO;2-T)
- <span id="page-19-4"></span>5. Kail, N., Briesen, H., & Marquardt, W. (2008). Analysis of FBRM measurements by means of a 3D optical model. *Powder Technology, 185*(3), 211–222. [https://doi.org/10.1016/j.powtec.2007.10.015.](https://doi.org/10.1016/j.powtec.2007.10.015)
- <span id="page-19-5"></span>6. Jiang, M., Zhu, X., Molaro, M. C., Rasche, M. L., Zhang, H., Chadwick, K., et al. (2014). Modifcation of crystal shape through deep temperature cycling. *Industrial and Engineering Chemistry Research, 53*(13), 5325–5336. [https://doi.org/10.1021/ie400859d.](https://doi.org/10.1021/ie400859d)
- <span id="page-19-9"></span>7. Adlington, N. K., Black, S. N., & Adshead, D. L. (2013). How to use the lasentec FBRM probe on manufacturing scale. *Organic Process Research & Development, 17*(3), 557–567. [https://doi.](https://doi.org/10.1021/op300326b) [org/10.1021/op300326b](https://doi.org/10.1021/op300326b).
- <span id="page-19-10"></span>8. Hefels, C., Willemse, A., & Scarlett, B. (1996). Possibilities of near backward light scattering for characterizing dense particle systems. *Powder Technology, 86*(1), 127–135. [https://doi.](https://doi.org/10.1016/0032-5910(95)03047-6) [org/10.1016/0032-5910\(95\)03047-6.](https://doi.org/10.1016/0032-5910(95)03047-6)
- <span id="page-19-11"></span>9. Whelan, J., Murphy, E., Pearson, A., Jefers, P., Kieran, P., McDonnell, S., et al. (2012). Use of focussed beam refectance measurement (FBRM) for monitoring changes in biomass concentration. *Bioprocess and Biosystems Engineering, 35*(6), 963–975. [https://doi.org/10.1007/s0044](https://doi.org/10.1007/s00449-012-0681-9) [9-012-0681-9](https://doi.org/10.1007/s00449-012-0681-9).
- <span id="page-19-12"></span>10. Abu Bakar, M. R., Nagy, Z. K., & Rielly, C. D. (2010). Investigation of the efect of temperature cycling on surface features of sulfathiazole crystals during seeded batch cooling crystallization. *Crystal Growth & Design, 10*(9), 3892–3900. [https://doi.org/10.1021/cg1002379.](https://doi.org/10.1021/cg1002379)
- <span id="page-19-13"></span>11. Kail, N., Briesen, H., & Marquardt, W. (2007). Advanced geometrical modeling of focused beam refectance measurements (FBRM). *Particle & Particle Systems Characterization, 24*(3), 184–192. <https://doi.org/10.1002/ppsc.200601036>.
- <span id="page-19-14"></span>12. Tadayyon, A., & Rohani, S. (1998). Determination of particle size distribution by Par-Tec® 100: Modeling and experimental results. *Particle & Particle Systems Characterization, 15*(3), 127–135.
- <span id="page-19-15"></span>13. Yu, Z. Q., Chow, P. S., & Tan, R. B. H. (2008). Interpretation of focused beam refectance measurement (FBRM) data via simulated crystallization. *Organic Process Research & Development, 12*(4), 646–654.<https://doi.org/10.1021/op800063n>.
- <span id="page-19-8"></span>14. Pandit, A., Katkar, V., Ranade, V., & Bhambure, R. (2019). Real-time monitoring of biopharmaceutical crystallization: Chord length distribution to crystal size distribution for lysozyme, rHu insulin, and vitamin B12. *Industrial and Engineering Chemistry Research, 58*(18), 7607–7619. [https://doi.](https://doi.org/10.1021/acs.iecr.8b04613) [org/10.1021/acs.iecr.8b04613.](https://doi.org/10.1021/acs.iecr.8b04613)
- <span id="page-19-6"></span>15. Zhang, B., Abbas, A., & Romagnoli, J. A. (2011). Multi-resolution fuzzy clustering approach for image-based particle characterization for particle systems. *Chemometrics and Intelligent Laboratory Systems, 107*(1), 155–164.<https://doi.org/10.1016/j.chemolab.2011.03.001>.
- <span id="page-19-7"></span>16. El Arnaout, T., & Cullen, P. J. (2017). Non-invasive 3D and 360° optical imaging of micro-particles. *Scientifc Reports, 7*(1), 6384. <https://doi.org/10.1038/s41598-017-06830-8>.
- <span id="page-20-0"></span>17. El Arnaout, T., & Cullen, P. J. (2020). In situ image processing and data binning strategy for particle engineering applications. *Chemical Engineering and Technology*. [https://doi.org/10.1002/](https://doi.org/10.1002/ceat.201900311) [ceat.201900311.](https://doi.org/10.1002/ceat.201900311)
- <span id="page-20-1"></span>18. Gao, Z., Rohani, S., Gong, J., & Wang, J. (2017). Recent developments in the crystallization process: toward the pharmaceutical industry. *Engineering, 3*(3), 343–353. [https://doi.org/10.1016/J.](https://doi.org/10.1016/J.ENG.2017.03.022) [ENG.2017.03.022](https://doi.org/10.1016/J.ENG.2017.03.022).
- <span id="page-20-2"></span>19. Wei, H., Yang, C., & Yu, Q. (2017). Contour segment grouping for object detection. *Journal of Visual Communication and Image Representation, 48,* 292–309. [https://doi.org/10.1016/j.jvcir](https://doi.org/10.1016/j.jvcir.2017.07.003) [.2017.07.003](https://doi.org/10.1016/j.jvcir.2017.07.003).
- <span id="page-20-3"></span>20. Larsen, P. A., Rawlings, J. B., & Ferrier, N. J. (2007). Model-based object recognition to measure crystal size and shape distributions from in situ video images. *Chemical Engineering Science, 62*(5), 1430–1441. [https://doi.org/10.1016/j.ces.2006.11.018.](https://doi.org/10.1016/j.ces.2006.11.018)
- <span id="page-20-4"></span>21. Ahmad, O., Suleiman, D. J., Gherras, N., Presles, B., Févotte, G., & Pinoli, J.-C. (2012). Recognizing overlapped particles during a crystallization process from in situ video images for measuring their size distributions. *Journal of Electronic Imaging, 21*(2), 0211115. [https://doi.org/10.1117/1.](https://doi.org/10.1117/1.jei.21.2.021115) [jei.21.2.021115](https://doi.org/10.1117/1.jei.21.2.021115).
- <span id="page-20-5"></span>22. Zou, K., Liu, T., Huo, Y., Zhang, F.-K., & Ni, X. (2017). Image analysis for in situ detection of agglomeration for needle-like crystals.
- <span id="page-20-6"></span>23. Agimelen, O. S., Jawor-Baczynska, A., McGinty, J., Dziewierz, J., Tachtatzis, C., Cleary, A., et al. (2016). Integration of in situ imaging and chord length distribution measurements for estimation of particle size and shape. *Chemical Engineering Science, 144,* 87–100. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.ces.2016.01.007) [ces.2016.01.007](https://doi.org/10.1016/j.ces.2016.01.007).
- <span id="page-20-7"></span>24. Sternberg, S. R. (1983). Biomedical image processing. *Computer, 16*(1), 22–34. [https://doi.](https://doi.org/10.1109/MC.1983.1654163) [org/10.1109/MC.1983.1654163](https://doi.org/10.1109/MC.1983.1654163).
- <span id="page-20-8"></span>25. Cho, J. W., Choi, Y. S., & Jeong, K. M. (2019). Performance of the eye-safe LRS and color CCD camera under aerosol environments. *Sensing and Imaging, 20*(1), 10. [https://doi.org/10.1007/s1122](https://doi.org/10.1007/s11220-019-0232-4) [0-019-0232-4](https://doi.org/10.1007/s11220-019-0232-4).
- <span id="page-20-9"></span>26. Zapata-Pérez, J., Doménech-Asensi, G., Ruiz-Merino, R., Martínez-Álvarez, J. J., Fernández-Berni, J., & Carmona-Galán, R. (2020). Fixed pattern noise analysis for feature descriptors in CMOS APS images. *Sensing and Imaging, 21*(1), 14.<https://doi.org/10.1007/s11220-020-0278-3>.
- <span id="page-20-13"></span>27. El Arnaout, T., Kurki, L., Vaarala, T., Ojala, K., Cullen, P. J., & Sullivan, C. (2016). Crystallization monitoring using simultaneous bright feld and PlasDIC imaging. *Chemical Engineering Journal, 300,* 64–74.<https://doi.org/10.1016/j.cej.2016.04.126>.
- <span id="page-20-14"></span>28. Huo, Y., Liu, T., Yang, Y., Ma, C. Y., Wang, X. Z., & Ni, X. (2020). In situ measurement of 3D crystal size distribution by double-view image analysis with case study on L-glutamic acid crystallization. *Industrial and Engineering Chemistry Research, 59*(10), 4646–4658. [https://doi.org/10.1021/](https://doi.org/10.1021/acs.iecr.9b05828) [acs.iecr.9b05828.](https://doi.org/10.1021/acs.iecr.9b05828)
- <span id="page-20-15"></span>29. Cardona, J., Ferreira, C., McGinty, J., Hamilton, A., Agimelen, O. S., Cleary, A., et al. (2018). Image analysis framework with focus evaluation for in situ characterisation of particle size and shape attributes. *Chemical Engineering Science, 191,* 208–231. [https://doi.org/10.1016/j.ces.2018.06.067.](https://doi.org/10.1016/j.ces.2018.06.067)
- <span id="page-20-16"></span>30. Lu, Z., Zhang, L., Jiang, Y., Zhang, C., Zhang, G., & Liu, M. (2019). Crystal morphology monitoring based on in situ image analysis of L-glutamic acid crystallization. In: *Advances in computer science research*.
- <span id="page-20-10"></span>31. Rashed, M., & Rashed, E. A. (2017). Double-sided sliding-paraboloid (DSSP): A new tool for preprocessing GPR data. *Computers & Geosciences, 102,* 12–21. [https://doi.org/10.1016/j.cageo](https://doi.org/10.1016/j.cageo.2017.02.005) [.2017.02.005](https://doi.org/10.1016/j.cageo.2017.02.005).
- 32. Dimov, I. K., Lu, R., Lee, E. P., Seita, J., Sahoo, D., Park, S.-M., et al. (2014). Discriminating cellular heterogeneity using microwell-based RNA cytometry [Article]. *Nature Communications, 5,* 3451. [https://doi.org/10.1038/ncomms4451.](https://doi.org/10.1038/ncomms4451)
- <span id="page-20-11"></span>33. Ketteler, R., Freeman, J., Stevenson, N., Ferraro, F., Bata, N., Cutler, D. F., et al. (2017). Imagebased siRNA screen to identify kinases regulating Weibel–Palade body size control using electroporation [Data Descriptor]. *Scientifc Data, 4,* 170022.<https://doi.org/10.1038/sdata.2017.22>.
- <span id="page-20-12"></span>34. Poddar, S., Pedersen, M., & Karar, V. (2018). Color image modifcation with and without hue preservation. *Sensing and Imaging, 19*(1), 35. [https://doi.org/10.1007/s11220-018-0219-6.](https://doi.org/10.1007/s11220-018-0219-6)
- <span id="page-20-17"></span>35. El Arnaout, T., Cullen, P. J., & Sullivan, C. (2016). A novel backlight fber optical probe and image algorithms for real time size-shape analysis during crystallization. *Chemical Engineering Science, 149,* 42–50. [https://doi.org/10.1016/j.ces.2016.04.025.](https://doi.org/10.1016/j.ces.2016.04.025)
- <span id="page-21-0"></span>36. Schindelin, J., Arganda-Carreras, I., Frise, E., Kaynig, V., Longair, M., Pietzsch, T., et al. (2012). Fiji: An open-source platform for biological-image analysis. *Nature Methods, 9*(7), 676–682. [https://](https://doi.org/10.1038/nmeth.2019) [doi.org/10.1038/nmeth.2019](https://doi.org/10.1038/nmeth.2019).
- <span id="page-21-1"></span>37. Danz, R., Vogelgsang, A., Käthner, R., Zeiss, C., & Plant, G. (2004). PlasDIC—A useful modifcation of the diferential interference contrast according to Smith/Nomarski in transmitted light arrangement. *Photonik, 1,* 42–45.
- <span id="page-21-2"></span>38. Cameron, R. P., Vogl, U., & Trautmann, N. (2020). Interference-contrast optical activity: a new technique for probing the chirality of anisotropic samples and more. *Royal Society Open Science, 7*(5), 192201. [https://doi.org/10.1098/rsos.192201.](https://doi.org/10.1098/rsos.192201)
- <span id="page-21-3"></span>39. Zhu, H., Chan, F. H. Y., & Lam, F. K. (1999). Image contrast enhancement by constrained local histogram equalization. *Computer Vision and Image Understanding, 73*(2), 281–290. [https://doi.](https://doi.org/10.1006/cviu.1998.0723) [org/10.1006/cviu.1998.0723.](https://doi.org/10.1006/cviu.1998.0723)
- <span id="page-21-4"></span>40. Sarkar, D., Doan, X.-T., Ying, Z., & Srinivasan, R. (2009). In situ particle size estimation for crystallization processes by multivariate image analysis. *Chemical Engineering Science, 64*(1), 9–19. [https://doi.org/10.1016/j.ces.2008.09.007.](https://doi.org/10.1016/j.ces.2008.09.007)
- <span id="page-21-5"></span>41. Yu, W., Liao, L., Bharadwaj, R., & Hancock, B. C. (2017). What is the "typical" particle shape of active pharmaceutical ingredients? *Powder Technology*. [https://doi.org/10.1016/j.powte](https://doi.org/10.1016/j.powtec.2017.02.043) [c.2017.02.043.](https://doi.org/10.1016/j.powtec.2017.02.043)
- <span id="page-21-6"></span>42. FDA. (2007). *ANDAs: Pharmaceutical solid polymorphism—Chemistry, manufacturing, and controls information (FDA, USA)*. White Oak: Food and Drug Administration.
- 43. Datta, S., & Grant, D. J. W. (2004). Crystal structures of drugs: advances in determination, prediction and engineering. *Nature Reviews Drug Discovery, 3*(1), 42–57. [https://doi.org/10.1038/nrd12](https://doi.org/10.1038/nrd1280) [80](https://doi.org/10.1038/nrd1280).
- <span id="page-21-7"></span>44. Upadhyay, P. P., Pudasaini, N., Mishra, M. K., Ramamurty, U., & Rantanen, J. (2018). Early assessment of bulk powder processability as a part of solid form screening. *Chemical Engineering Research and Design*. <https://doi.org/10.1016/j.cherd.2018.05.020>.
- <span id="page-21-8"></span>45. Wang, K., Zhuo, L., Li, J., Jia, T., & Zhang, J. (2020). Learning an enhancement convolutional neural network for multi-degraded images. *Sensing and Imaging, 21*(1), 25. [https://doi.org/10.1007/](https://doi.org/10.1007/s11220-020-00289-0) [s11220-020-00289-0](https://doi.org/10.1007/s11220-020-00289-0).
- 46. Wu, Y., Lin, M., & Rohani, S. (2020). Particle characterization with on-line imaging and neural network image analysis. *Chemical Engineering Research and Design*. [https://doi.org/10.1016/j.cherd](https://doi.org/10.1016/j.cherd.2020.03.004) [.2020.03.004](https://doi.org/10.1016/j.cherd.2020.03.004).
- <span id="page-21-9"></span>47. Gao, Z. (2019). Non-classical nucleation phenomena study and following process monitoring and optimization in solution crystallization process. Thesis. Western University. [https://ir.lib.uwo.ca/](https://ir.lib.uwo.ca/etd/6130/) [etd/6130/](https://ir.lib.uwo.ca/etd/6130/).

**Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.