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# Horseradish and Soybean Peroxidases: Comparable Tools for Alternative Niches?

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Horseradish and soybean peroxidases (HRP and SBP, respectively) are useful biotechnological tools. HRP is often termed the classical plant heme peroxidase, and although it has been studied for decades our understanding has deepened since its cloning and subsequent expression, which has enabled numerous mutational and protein engineering studies. SBP, however, has been neglected until recently; despite offering a real alternative to HRP that actually outperforms it in terms of stability. SBP is now used in numerous biotechnological applications, including biosensors. Review of both is timely. This article summarises and discusses the main insights into the structure and mechanism of HRP, with special emphasis on HRP mutagenesis, and outlines its use in a variety of applications. It also reviews current knowledge and applications to date of SBP, particularly biosensors. The final paragraphs speculate on the future of plant heme-based peroxidases, with probable trends outlined and explored.

**Key Words:** Horseradish Peroxidase, Soybean Peroxidase, Review, Biosensors, Biocatalysis, Mutagenesis.

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## Abbreviations:

**ABTS**, 2,2'-Azino-di-(3-ethyl-benzthiazoline-6-sulphonic acid); **Co(bpy)**, Tris-2,2'-dipyridylcobalt(III) perchlorate trihydrate; **CV**, Cyclic voltammogram. **DAB**, 3,3'-diaminobenzidine tetrahydrochloride. **DMPC**, Dimyristoylphosphatidylcholine epoxidised olefins. **DTT**, dithiothreitol; **ELISA**, Enzyme-Linked Immunosorbent Assay. **GOx**, Glucose oxidase; **GP**, Graphite Powder; **H<sub>2</sub>O<sub>2</sub>**, Hydrogen peroxide; **HQ**, hydroquinone; **IPTG**, iso-propyl-β-thiogalactoside; **MET**, mediated electron transfer; **mm**, millimetre. **PEGDGE**, Poly(ethylene glycol) (400) diglycidylether. **PeIB**, pectate lyase B fragment. **PDB**, Protein Data Bank; **POs-EA**, Os(bpy)<sub>2</sub>Cl<sup>+2+</sup> poly(4-vinylpyridine) quaternised with 2-bromoethylamine. **Pt**, Platinum. **PVA**, Polyvinyl alcohol. **PVP**, Polyvinyl pyridine. **PVP-Os<sup>III</sup>**, polyvinyl pyridine-osmium complex. **POCT**, point-of-care testing; **Px**, peroxidase. **Px<sub>ox</sub>**, oxidised peroxidase. **rHRP**, recombinant HRP; **TTCA**, poly-5,2':5',2"-terthiophene-3'-carboxylic acid.

41 **Introduction.**

42 Peroxidase enzymes span the bioscience and biotechnology spectra, ranging from  
43 bioremediation [1] and biocatalysis [2] through diagnostics [3] and biosensors [4] to  
44 recombinant protein expression [5], transgenics [6], bioinformatics [7], protein  
45 engineering [8] and even to therapeutics [9]. This article contrasts two key heme-  
46 containing plant peroxidases, horseradish (HRP) and soybean (SBP), with special  
47 emphasis on mutagenesis studies and biosensor applications. HRP is a ‘traditional’  
48 enzyme, whereas SBP emerged in the 1990s. A comparison of these key  
49 biotechnological tools is timely.

50

51 **Brief biochemistry of horseradish and soybean peroxidase:**

52 All heme peroxidases (E.C. 1.11.1.7) have a ferriprotoporphyrin IX prosthetic group  
53 located at the active site [10]. Both HRP and SBP are classified as Class III Classical  
54 Secretory plant peroxidases [7 & 11] and as such share common features (Table 1).  
55 Their catalytic mechanism involves a two-electron oxidation of the heme moiety to an  
56 intermediate known as Compound I. Successive one-electron reductions return the  
57 enzyme to its resting state via a second intermediate, Compound II [6].

58 Determining the *in vivo* function of peroxidases is complex owing to the numbers of  
59 isoenzymes in[E1] the family [12]. Interestingly, despite the several *in vitro* uses of  
60 HRP, its actual *in vivo* role has never been elucidated. Several suggestions have been  
61 proposed based on the known roles of other plant peroxidases. Peroxidases are usually  
62 found in the cell wall, vacuoles, transport organelles and the rough endoplasmic  
63 reticulum, and have noted roles in lignification, wound healing and auxin catabolism  
64 [4]. SBP has been isolated from that plant’s seed coat and its presence prevents  
65 premature germination[E2] [13]. Plant peroxidases can use lignin and other plant  
66 compounds as reducing substrates. Indeed, SBP has been noted to polymerise  
67 coniferyl alcohol, indicating that it can efficiently catalyse reactions involving lignin  
68 precursors [14]. Therefore, it is possible that peroxidases are involved in the  
69 lignification or suberisation processes of plants [15].

70 **Recombinant Peroxidase Expression.**

71 Recombinant hemoprotein expression has been plagued by inclusion body formation,  
72 most notably in recombinant HRP expression. Several general methods have been  
73 suggested to reduce the formation of inclusion bodies, including reducing cultivation  
74 temperature and altering inducer composition and concentration [16]. Other, more  
75 peroxidase-specific, methodologies have been cited, including use of specific *E. coli*  
76 strains [17], inclusion of chaperones [18] and use of leader sequences [19]. Another  
77 major obstacle in the recombinant expression of hemoproteins is the limited  
78 availability of heme and iron within a bacterial cell. Bacterial cells each contain  $10^5$  to  
79  $10^6$  iron ions, which are essential for many metabolic pathways [20]. Culture  
80 supplementation with the heme precursor  $\delta$ -aminolevulinic acid has been suggested  
81 [21]; but Goodwin and co-workers have recently developed an elegant co-expression  
82 system incorporating a membrane heme receptor, allowing the use of exogenous heme  
83 as an iron source [22]. Jung and co-workers [5] noted an increased ratio of holoprotein  
84 to apoprotein with less-intense induction conditions, suggesting that slow recombinant  
85 hemoprotein production appears to allow easier incorporation of the available heme  
86 into the apoprotein [5]. Developments in HRP expression in both prokaryotic and  
87 eukaryotic systems are outlined in the supplementary online material.

88

89 **HRP: Cloning and Expression.**

90 The gene coding for the HRP protein was first synthesised by Smith and co-workers  
91 [23] based on the protein sequence published by Welinder [24]. This 940 basepairs  
92 synthetic gene was designed using commonly used codons in *E. coli* to minimise  
93 protein truncation owing to codon bias [25]. Recombinant HRP was over-expressed  
94 by induction but this led to the formation of misfolded apoprotein and the requirement  
95 to disrupt these aggregates, refold the protein correctly and add the heme centre.  
96 Disruption involved addition of EDTA to chelate ions, lysozyme and DNase to  
97 reduce viscosity of the bacterial cell lysate, urea to solubilise the protein, and  
98 dithiothreitol (DTT) to break disulphide bonds. Refolding required slow exchange of  
99 disrupting reagents with folding facilitators such as calcium (for structural integrity),  
100 oxidized glutathione (to reform disulphide bridges) and hemin (to provide the  
101 prosthetic heme group) [23].

102

103 **SBP: Cloning and Expression.**

104 The gene encoding SBP was first derived from a soybean plant cDNA library  
105 screened with a peroxidase-specific probe [26]. The open reading frame for the SBP  
106 protein was cloned into the pET-34b (+) expression vector; however, induction of  
107 rSBP led to inclusion body formation and *E. coli* cell death. Active SBP was achieved  
108 by a refolding strategy similar to that of HRP. Unlike rHRP refolding, however,  
109 inclusion of oxidised glutathione in the refolding medium decreased active SBP  
110 recovery, possibly due to the formation of mispaired disulphides [26]. Henriksen and  
111 co-workers [15] also developed a recombinant SBP for crystallisation studies based  
112 on previous cDNA work, in which they successfully refolded SBP from inclusion  
113 bodies using a cocktail that included both oxidized and reduced glutathione [27].  
114 There have been several other recent examples of recombinantly expressed plant  
115 peroxidases including Hushpulia and co-workers' [28] work on tobacco anionic  
116 peroxidase.

117 **Mutagenesis of HRP.**

118 Mutagenic studies on HRP began following successful cloning of a synthetic HRP  
119 gene [23]. Before the elucidation of the crystal structure [29], most mutations focused  
120 on ascertaining the key residues in the active site. Before 1997, researchers based their  
121 assumptions on crystal structures of closely related peroxidases, such as cytochrome *c*  
122 peroxidase, which suggested positions 38 through 42 as key catalytic residues.  
123 Mutations of Arg 38, Phe 41 and His 42 led to dramatic decreases in peroxidase  
124 catalytic activity. However, some Phe 41 mutants revealed an augmented thioether  
125 sulfoxidation activity owing to increased access channel area [30].

126 Asn 70 was also noted as an important residue in HRP catalysis: although it lies some  
127 distance from the heme iron atom, it is hydrogen-bonded to the side chain of the distal  
128 His 42. [31] Mutations in this region showed a decrease in HRP activity and a re-  
129 orientation of active site residues. Mutation of Phe 221 altered the heme iron of the  
130 resting enzyme to a quantum mixed-spin state [32]. Substitutions of Trp 117 revealed  
131 this residue's role in protein folding and electron transfer [33]. Mutations within the  
132 active site entrance revealed the key role of Phe 142 in binding aromatic molecules  
133 [34], whilst mutations within the proximal region (the area below the heme plane)  
134 disclosed the parts played by Phe 179 in aromatic molecule binding [35] and by His  
135 170 in heme group anchorage [36]. Table 2 lists the various site directed HRP  
136 mutants. Recent examples of site directed mutants of HRP have been fewer [37], but  
137 Colas and de Montellano [38] identified the key role of carboxylate side chain amino  
138 acids in HRP protein-heme interactions.

139 To date, there have been few reports of HRP random mutagenesis. Arnold and co-  
140 workers directionally evolved HRP with the aim of increasing activity and stability.  
141 Development of a stabilised recombinant HRP is of great importance to increase and  
142 consolidate the range of peroxidase applications. Three rounds of random mutagenesis  
143 improved expression in yeast, yielding a nine-position HRP mutant displaying an 85-  
144 fold increase in activity over the parental molecule. One round of random mutagenesis  
145 was also carried out to improve stability, resulting in three mutants more stable than  
146 the parent in relation to temperature and H<sub>2</sub>O<sub>2</sub> tolerance (supplementary online  
147 material) [8, 19, 39, & 40]. Recent publications suggest that targeted, "semi-rational",  
148 evolution of enzymes might yield superior mutants in less time [41 and references  
149 within]. Mendive and co-workers developed a rapid screening methodology for

150 random mutants displaying increased peroxidase activity, using DAB as substrate.  
151 Whereas Arnold and co-workers expressed HRP in *E. coli*, *S. cerevisiae*. and *P.*  
152 *pastoris*, Mendive and co-workers utilised a baculovirus expression system [42].  
153 No mutagenesis studies on recombinant SBP have been reported to date, and the  
154 authors believe that this requires urgent attention. For example, mutagenesis could  
155 reveal which residues in SBP endow it with its enhanced stability *vis a vis* HRP [15].  
156 Also, similar to HRP, SBP could be subjected to focussed directed evolution to  
157 increase the number of substrates accepted.

158

### 159 **HRP in Biosensors**

160 One of the most common uses of HRP is in biosensors. A biosensor is “*an analytical*  
161 *device that brings together an immobilised biological sensing material [often HRP]*  
162 *and a transducer to produce an electronic signal that is proportional to the*  
163 *concentration of the target chemical substance”* [43]. Although reports of SBP-based  
164 biosensors are emerging, HRP biosensor research dominates and has continued to  
165 develop through many forms, from the traditional voltammetric- and amperometric-  
166 based methods of detection, to nano-sized devices. Real time quantification of  
167 hydrogen peroxide continues to be one of the main reasons for sensor development  
168 [44], although other diverse applications include the detection of glucose [45], ethanol  
169 [46] and tumour markers *in vivo* [47]. Enzyme-based biosensors require rapid and  
170 uniform transfer of electrons generated at the enzyme active site to the transducer.  
171 The distance between the active site and the transducer can hinder electron transfer;  
172 often, posttranslational modifications such as glycosylation increase this distance.  
173 Recombinant HRP, devoid of glycans, offers a shorter path for electron transfer and  
174 numerous reports of rHRP-based sensors have appeared [48 and references  
175 within]. While a detailed review of HRP-based sensors is beyond the scope of this  
176 article, we outline some of the emerging trends in HRP-based biosensor development.  
177 Biosensors, including immunosensors [49] and electroensors [50], incorporating  
178 organic solvents have developed as an expanding area of peroxidase research,  
179 primarily due to insolubility of many analytes in aqueous solutions. Recently, Konash  
180 and Magner [51] developed a HRP-immobilised, mediated H<sub>2</sub>O<sub>2</sub> sensor, which  
181 demonstrated good catalytic activity in 2-butanone and ethyl acetate. Organic solvent  
182 compatible bi-enzyme peroxidase sensors have also been cited in the literature [52].

183 Size reduction remains a pivotal area in sensor research. The use of nanoparticles  
184 offers increased surface area for enzyme immobilisation, whilst simultaneously  
185 reducing apparatus size [53]. Currently, HRP-based nano-sensors are at the forefront  
186 of biosensor research [54 and Table 3].

187

### 188 **SBP in biosensors.**

189 Although HRP is the classical heme peroxidase, there is increasing interest in SBP.  
190 SBP has advantages over HRP in terms of catalytic activity and stability [55]; these  
191 can be exploited in biosensors. Also, unlike HRP, SBP is active in the pH range 2-6,  
192 offering a greater range of potential biosensing applications [56]. The first SBP  
193 biosensor was reported in 1995 by Vreeke and co-workers [57] as a thermostable  
194 wired enzyme electrode using an osmium-based mediator, which aids electron transfer  
195 from the active site to the electrode, modified by an epoxide. Kenausis and co-  
196 workers [58] also used a poly(4-vinylpyridine) polymer, complexing the pyridine  
197 nitrogens to the osmium-based mediator, quaternised with 2-bromoethylamine. Until  
198 the use of SBP by Heller and Vreeke [59], no peroxidase-based sensors could be used  
199 at 37°C for an extended period (~100 hours). Monitoring of glucose “*in vivo*” for  
200 diabetes mellitus, and of lactate for confirmation of hypoxia and ischemia, are vital in  
201 patient management; use of thermostable SBP immobilised into a mediator enables  
202 this [59].

203 The typical electrochemical reactions of a H<sub>2</sub>O<sub>2</sub> sensing, peroxidase based, osmium-  
204 mediated electrode system are outlined in Box 1 [60]. In addition to conveying  
205 electrons, generated at the active site, to the electrode surface, the pyridin-N-ethylene  
206 groups of the osmium-containing mediator also increase the hydration and provide  
207 primary amines for cross-linking [58].

208 Table 3 summarizes reports to date of SBP-based biosensors, most of which use a  
209 mediator. H<sub>2</sub>O<sub>2</sub> can be electrochemically detected by its electrooxidation on a Pt (or  
210 other inert Pt group metal) electrode [59]. Utilisation of an enzyme/mediator system  
211 produces a multi-step mediated electron-transfer (MET) process, in which each step  
212 transports the electron a small distance [61]. However, use of a mediator with a redox  
213 enzyme can create its own problems: the mediator-enzyme film can, depending upon  
214 its thickness, obstruct substrate diffusion [62]. Chemical modification of redox  
215 enzymes with an electron relay moiety can increase multi-step MET by decreasing the



216 electron transfer distance. This leads to improved electrical communication between  
217 the enzyme's redox centre and its external environment [61].

218

### 219 **Peroxidase based Micro- and Nano-Systems**

220 An emerging field in peroxidase research is the use of micro- and nano- sized  
221 structures in diagnostic and biosensing fields. Enzyme microreactors, for example,  
222 permit chemical and biochemical reactions to be carried out on a microscale. HRP has  
223 previously been used as a model microreactor system to monitor HRP catalytic  
224 activity and as a diagnostic tool; however, the development of this field will be  
225 determined by the ability to immobilise peroxidases onto suitable support structures  
226 [63]. Miniaturisation of enzymatic processes is also evident in the diagnostics sector,  
227 in “Lab on a Chip” and “Point of Care Testing”(POCT) research. The classical  
228 application of HRP in POCT is Clinistix<sup>TM</sup>; however, recently Cho and co-workers  
229 [64] have applied HRP to a portable sequential cross flow immunoanalytical device.  
230 Cross flow immunoassays are capable of introducing the antigen-antibody complex  
231 to the flow cell whilst sequentially extracting the catalytic signal, thus simplifying the  
232 complex traditional ELISA procedure. This device demonstrates many advantages  
233 over ELISA-based analytical methodologies including a rapid, sensitive and  
234 inexpensive in-situ diagnosis for the presence of Hepatitis B surface antigen in a  
235 sample. Miniaturisation of peroxidase-based devices also features in other research  
236 fields, including the use of micro-crystals for oxidoreductase-based catalysis in  
237 organic solvent [65], nano-immobilisation techniques for peroxidase based  
238 wastewater treatment [66], and a more widespread use of nano-structures for  
239 peroxidase based sensors [67 & 68]. Recently Yan and co-workers [69] described a  
240 microcantilever based biosensor, modified with HRP, for H<sub>2</sub>O<sub>2</sub> detection. In this  
241 system the enzyme-functionalised microcantilevers deflected irreversibly in response  
242 to H<sub>2</sub>O<sub>2</sub> concentrations in the nanomolar range. The deflection was caused by  
243 conformational change within the HRP molecule as it underwent oxidation by the  
244 H<sub>2</sub>O<sub>2</sub>; the irreversibility was due to the absence of a second, reducing substrate  
245 required for reversion of HRP to the resting state. This technique may also provide a  
246 sensitive tool for investigating protein structural change. HRP has also been utilised as  
247 the functional component of self-assembled three-dimensional (3-D) nano-structures.  
248 Rauf and co-workers [70] utilised self-assembly layer-by-layer technology to

249 construct controlled 3-D catalytically active nano-structures. This method of  
250 peroxidase immobilisation allows for increased catalytic activity per unit area, and  
251 will aid in the miniaturisation of biosensors, biochips and immobilised biocatalysts.  
252 With increasing sophistication of support structures on the micro- and nano-scale,  
253 miniaturisation of peroxidase-based devices will continue to develop in the future,  
254 particularly in the fields of POCT and biosensing.

255

### 256 **Peroxidase based Biocatalysis.**

257 A major shortcoming of all heme-dependent peroxidases is their low operational  
258 stability, owing to oxidative degeneration of the heme group [71]. Operational  
259 stability of SBP can be increased by generating  $H_2O_2$  *in situ* from glucose and  $O_2$ .  
260 When co-immobilized with glucose oxidase in a polyurethane foam, SBP could act as  
261 a peroxygenase to convert thioanisole to its sulphoxide (i.e. by inserting an oxygen  
262 atom). Here, SBP uses the  $H_2O_2$  generated *in situ* by glucose oxidase; it formed no  
263 sulphoxide with free, exogenous  $H_2O_2$  [71]. Such an arrangement avoids excessive  
264 initial  $H_2O_2$  concentrations and, hence, formation of compound III (a reversible dead-  
265 end complex formed from compound II in the presence of an excess  $H_2O_2$ , which  
266 slowly reverts to native enzyme; [10]) and/or irreversibly inhibited SBP. HRP has  
267 been subjected to intense experimentation, including a large body of work focussing  
268 on site directed mutants (see above and Table 2). Now that the key catalytic residues  
269 are known, researchers have begun to use site directed mutagenesis to alter the  
270 function of the HRP molecule [48], e.g. by construction of an improved luminol  
271 binding site [72]. Directed evolution stabilised HRP against thermal denaturation [39]  
272 and has endowed it with increased  $H_2O_2$  tolerance and increased catalytic activity  
273 [40]. Further targeted directed evolution, focussing on the substrate access channel  
274 and binding pocket, could allow HRP to accept an increased variety of substrates [73],  
275 and promote further diversification of HRP applications in organic synthesis [41].  
276 Peroxidase catalysis in organic solvents, both aqueous and anhydrous, offers a huge  
277 advantage to organic chemists, as difficult asymmetric oxidation and reduction  
278 reactions can take place rapidly and with high specificity [74]. The major problems of  
279 substrate solubility and unwanted side reactions promoted by water are also overcome  
280 during organic solvent based synthesis. Additionally, in some anhydrous solvents  
281 peroxidase (HRP and SBP) activity was actually increased [75], with additional

282 methods, such as salt activation [76] and excipient aided lyophilisation [77] also  
283 resulting in increased peroxidase activity. However, in some low water solvents,  
284 peroxidases can lose their conformational structure [78]; although recent advances in  
285 peroxidase encapsulation in amphiphilic matrices [79], the use of reverse micelles  
286 [80] and oil emulsions [81] allow for peroxidase activity in an extended range of  
287 anhydrous solvents. Reactions carried out in these solvents include hydroxylations, N-  
288 demethylations and sulphoxidations [2]. An interesting recent environmental  
289 application of SBP polymerisation in organic solvents is the production of  
290 polycardanol in as a potential anti-biofouling agent [82], whilst recently it has been  
291 noted that HRP requires a mediator to catalyse the same substrate [83]. The interested  
292 reader is directed to a recent review of this area [74].

293

#### 294 **Peroxidase based Bioremediation.**

295 Highly expressed, stabilized, recombinant HRP [39, 40] could be very useful for  
296 wastewater cleanup, provided the recombinant enzyme can be produced cost  
297 effectively and in sufficient quantities. Phenol cleanup by HRP has been widely  
298 reviewed [1] but several drawbacks limit its widespread application, including  
299 intolerance of high concentrations of the primary substrate H<sub>2</sub>O<sub>2</sub> [84], low enzymatic  
300 reusability, and financial costs. Plant heme peroxidase expression in *E.coli* can be  
301 frustrating, but advances in peroxidase expression, without formation of inclusion  
302 bodies, may pave the way for increased production of recombinant peroxidases (See  
303 supplementary online material). SBP has proven itself a worthy alternative  
304 peroxidase: it displays superior stability and activity characteristics to the classical  
305 HRP. However, research into this enzyme lags far behind HRP. SBP can effectively  
306 cleanup phenolic wastewater, yet recent publications cite HRP as being a superior,  
307 albeit less stable, catalyst than SBP for phenol cleanup [85]. Development of an  
308 enhanced catalytic SBP mutant would provide a powerful tool for wastewater  
309 treatment. Bódalo and co-workers [85] noted that the choice of peroxidase for  
310 wastewater treatment also depends on effluent characteristics, operational  
311 requirements and costs. SBP has been shown to outperform HRP in oxidative dye  
312 removal [86]. SBP, possibly owing to its larger substrate access channel, and, hence,  
313 greater exposure of the catalytically important delta heme edge, can accept more  
314 substrates than HRP [87].

315

316 **HRP: an Unlikely Therapeutic**

317 An exciting application of HRP is as a novel cancer treatment via gene-directed  
318 enzyme/prodrug therapy. It has been noted that the non-toxic HRP substrate, indole-3-  
319 acetic acid (IAA), forms a radical that is toxic to cancer cells upon HRP catalysis. The  
320 exact mechanism of toxicity remains unclear: it is believed to involve lipid  
321 peroxidation induced by the free radical formation [88]; however, in human  
322 melanoma cells, death receptor-mediated and mitochondrial apoptotic pathways are  
323 known to be involved also [89]. Leaving aside the actual reason(s) for toxicity,  
324 inactive IAA can be introduced to the body; and then becomes activated by HRP at  
325 the region of interest. Localisation of the HRP molecule is achieved via its  
326 conjugation to an antibody specific to an extracellular tumour antigen. This approach  
327 has become the focus of much research and numerous clinical trials, due to several  
328 attractive features: these include the robust nature of the activating enzyme and the  
329 low toxicity of the prodrug [90]. HRP has been shown to activate other pro-drugs  
330 including ellipticine [91] and halogenated IAA derivatives [92]. The interested reader is  
331 directed to the excellent recent review of Dachs et al. [93].

332 **Conclusion and Future Directions:**

333 As outlined, peroxidases are widely studied and very important enzymes, with many  
334 applications in the life sciences and beyond. They remain pivotal to advancing  
335 biotechnology, and as such, we present two clearly distinct, yet similar members of  
336 this classical family. Continued research into the “traditional” HRP, has been  
337 accompanied by the slow, but steady progression of SBP. Crude SBP, isolated from  
338 waste soybean hulls, offers a cheap bulk peroxidase catalyst for applications such as  
339 wastewater treatment and organic synthesis, whilst the more costly peroxidase  
340 alternatives (plant HRP and recombinant HRP and SBP) will prove themselves in  
341 higher value niches, such as diagnostics and therapeutics. With improved  
342 understanding of the catalytic and stability characteristics, the detection of new  
343 substrates and the increasing use of implantable devices in the medical field, SBP will  
344 rapidly develop its own high value market niche. As noted for HRP, use of  
345 recombinant SBP would also benefit the biosensor field by permitting more rapid  
346 electron transfer, due to the lack of protein glycosylation. Improvement of these two  
347 peroxidases, by rational mutation and “focussed” directed evolution, will widen their  
348 applications and expand their roles as key biotechnological tools in the future.

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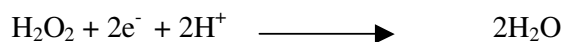
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**Box[E4] 1:** Typical biosensor based on a mediated peroxidase (Px) reaction scheme. Upon the addition of  $\text{H}_2\text{O}_2$ , Px catalyses the reaction forming water; in the process, Px goes through its catalytic cycle. This causes the mediator to go from its resting state of  $\text{Os}^{2+}$  to  $\text{Os}^{3+}$ . The osmium species is seen as a one-electron donor, used as the mediator to assist in electron transfer from the active site of Px to the electrode surface.  $\text{Px}_{\text{ox}}$  corresponds to the catalytic intermediate Compound I, formed by a two-electron oxidation. The individual one-electron reduction steps that take place on the enzyme itself (formation of catalytic intermediate Compound II and reversion to resting enzyme, Px) have been omitted from equation 2 for the sake of clarity. Adapted from *Analytica Chimica Acta*. **418**, Li W. et al. (2000). Fabrication of multilayer films containing horseradish peroxidase and polycation-bearing Os complex by means of electrostatic layer-by-layer adsorption and its applications as a hydrogen peroxide sensor. 225-232. [60] Copyright 2000, with permission from Elsevier.

**Table 1:** Comparison of the biochemical and structural properties of HRP and SBP. Key references are noted.

	<b>Horseradish Peroxidase</b>	<b>Soybean Peroxidase</b>
Species Name	<i>Armoracia rusticana</i> [23]	<i>Glycine max</i> [14]
Number of Amino Acids	306 [23]	306 [94]
Enzyme Classification	1.11.1.7 [29]	1.11.1.7 [15]
PDB Accession Number	1ATJ	1FHF
Molecular Weight	44,100 Da [6]	40,660 Da [15]
Carbohydrate	7,580 Da [6]	7,400 Da [15]
Heme Group	550 Da [6]	550 Da [15]
Calcium Ions	80 Da [6]	80 Da [15]
Glycosylation Sites	Asn: 13, 57, 158, 186, 198, 214, 255, 268, 316 [95]	Asn: 185, 197, 211, 216 [96]
pI	9.0 [24]	4.1 [14]
pH Activity Range	4-8 [97]	2-10 [15]
Secondary Structure	13 $\alpha$ -helices 3 $\beta$ -sheets [29]	13 $\alpha$ -helices 2 $\beta$ -sheets [15]
Disulphide Bridges	11-91, 44-49, 97-301, 177-209 [29]	11-91, 44-49, 97-299, 176-208 [15]
<i>In vivo</i> localisation	Roots, cell wall, vacuoles [6]	Hourglass cells, seed coat [14]

**Table 2:** Summary of HRP site directed mutants[E5]

Mutations		Conclusions
<b>Active Site.</b> [98, 99, & 100]		
R38A	F41T	Arg38 and His42 are key residues in enzyme catalysis.
R38E	F41V	Arg38 and His42 mutations decrease Compound I formation.
R38G	F41W	Mutants affect reactivity towards reducing substrates. NB- Morimoto ref.
R38H	H42A	Arg 38 and His42 are acid base partners. Arg 38 stabilises His 42.
R38K	H42E	Arg38 and His42 operate in concert to distally bind BHA.
R38L	H42L	Arg38 and His42 are dioxygen-heterolytic cleavers
R38S	H42Q	Arg38 plays a role in H <sub>2</sub> O <sub>2</sub> binding and cleavage.
F41A	H42R	Hydrophobicity of active site region is critical in enzymatic catalysis.
F41H	H42V	Space creating active site mutants alter substrate specificity.
F41L		Phe41 acts as hydrophobic barrier between Arg38 and His42.
<b>Active Site Entrance.</b> [101]		
S35K	F143E	Phe 142 plays a critical role in aromatic substrate binding.
F142A	F176E	Charged residues are important at the active site entrance.
F143A		Luminol binds to active site via electrostatic interactions in binding area.
<b>Proximal Region.</b> [102 & 36]		
F179A	H170A	Phe 179 is crucial for aromatic substrate binding.
F179H	F172T	His 170 tethers heme moiety in position. Prevents distal His coordination.
F179S		His 170 maintains heme moiety in penta-coordinated state.
<b>Asparagine 70.</b> [103]		
N70V	N70D	Asn70 hydrogen bonds to His42, mutations alter distal heme orientation. Mutant protein displays increased redox potential.
<b>Tryptophan 117.</b> [33 & 104]		
W117F		Mutants displayed increased acid stability. Trp 117 is important in internal electron transfer and protein unfolding.
<b>Threonine 171.</b> [37]		
T171S		Proximal structural alteration, affects proximal pocket hydrogen bonding.
<b>Phenylalanine 221.</b> [105 & 32]		
F221M	F221W	Mutants display decreased stability in alkaline conditions. Trp introduction destabilises protein, due to unfavourable surroundings.

Footnote:

Mutants are grouped into active site, active site entrance, proximal heme region, asparagine 70, tryptophan 117 and Phe 221 mutations. Mutants were expressed in a variety of hosts including *E.coli*, *Trichoplusia ni*, and *Spodoptera frugiperda* cell lines. Key references only are noted for each collection of mutants; further references may be found within these.

Electrode type	Size of Electrode	Enzyme	Method of immobilisation	Mediator	Analyte measured	Method of measurement	Reference
<b>Soybean Peroxidase</b>							
Glassy carbon	4mm	SBP	Adsorption	PVA/PVP	H <sub>2</sub> O <sub>2</sub>	Amperometry	[106]
Glassy carbon	4mm	SBP	Entrapment	Sol-gel	H <sub>2</sub> O <sub>2</sub>	Amperometry	[56]
Glassy carbon	3mm	SBP	Entrapment	Pos-EA, PEGDGE	H <sub>2</sub> O <sub>2</sub>	Amperometry	[59]
Pyrolytic graphite (Rotating disk)	0.2mm	SBP	Entrapment	DMPC	H <sub>2</sub> O <sub>2</sub>	Amperometry /CV	[107]
Glassy carbon	3mm	GOx/SBP	Entrapment	Pos-EA	Glucose	Amperometry	[58]

<b>Horseradish Peroxidase</b>							
Gold	-	HRP	Adsorption	DNA	H <sub>2</sub> O <sub>2</sub>	Amperometry	[108]
Glassy Carbon	3mm	HRP	Adsorption	HQ	H <sub>2</sub> O <sub>2</sub>	Amperometry	[53]
Carbon Ceramic	6mm	HRP	Covalent	Nano Au	H <sub>2</sub> O <sub>2</sub>	Amperometry	[109]
Carbon Paste	7mm	HRP	Entrapment	GP	Biogenic Amine	Amperometry	[110]
Titanium	0.8mm	HRP	Adsorption	Thionine	H <sub>2</sub> O <sub>2</sub>	Amperometry	[111]
Platinum	1mm	HRP	Entrapment	Co(bpy)	H <sub>2</sub> O <sub>2</sub>	Amperometry	[112]
Carbon Fibre	3mm	Cho/HRP	Adsorption	Os- PVP	Choline	Amperometry	[113]
Platinum	7mm	Cho/HRP	Adsorption	TTCA	Choline	Amperometry	114

<b>Recombinant HRP</b>							
Gold	0.15mm	rHRP	Adsorption	Direct	H <sub>2</sub> O <sub>2</sub>	Amperometry	[115]
Graphite Rotating disk	3mm	rHRP	Adsorption	Direct	H <sub>2</sub> O <sub>2</sub>	Amperometry	[116]
Gold	0.3mm	CytC / rHRP	Adsorption	Direct	Superoxide Anion radical	Amperometry	[117]
Gold	0.15mm	L-LO / rHRP	Adsorption	Direct	L-lysine	Amperometry	[118]

**Table 3:** Some SBP, HRP and rHRP biosensors, and their properties, from the literature.