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Quality and Nutritional Status of Fresh-cut Tomato as Affected by Spraying of Delactosed Whey Permeate Compared to Industrial Washing Treatment.

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Full Title

**Quality and Nutritional status of Fresh-cut Tomato as affected by Spraying of
Delactosed Whey Permeate compared to Industrial Washing Treatment**

Running Head

Shelf-life Extension of Tomatoes by DWP

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27 **Abstract**

28 The aim of this study was to examine the applicability of delactosed whey permeate
29 (DWP) treatment on preserving the quality and antioxidant attributes of fresh-cut
30 tomato. Tomatoes were treated with 3 % DWP by dipping, spraying and a
31 combination of both, stored at 4 °C for 10 days and compared with the industrial
32 standard, chlorine. The combination of dipping and spraying of DWP showed the best
33 results for all the markers tested. The combined treatment of dipping and spraying of
34 DWP significantly lowered total counts (~ 1.0 log cfu/g), yeast and moulds (~ 1.2 log
35 cfu/g), inhibited the loss of firmness (25 %) and reduced POD activity (15 %) of the
36 tomato slices after 10 days compared to the chlorine treatment. Moreover, DWP
37 treated tomatoes maintained significantly ($p < 0.05$) higher levels of vitamin C, total
38 phenols (TP) and antioxidant activity (DPPH) than the chlorine treated samples
39 during storage. Sensory scores confirmed that DWP treated tomatoes retained better
40 aroma and texture. Also, the appearance and overall acceptability were higher than
41 chlorine treated tomatoes. Thus DWP treatment has potential to extend the shelf-life
42 of fresh-cut tomatoes.

43

44 **Key words:** Whey permeate; Fresh-cut; Tomato; Shelf-life; Quality, Antioxidants.

45

46 **1. Introduction**

47 Tomato (*Lycopersicon esculentum* Mill.) is a versatile vegetable that is consumed
48 fresh as well as in the form of processed products (Toor & Savage, 2005). It is
49 considered as an important source of dietary antioxidants as it is rich in vitamins,
50 carotenoids and phenolic compounds (Lenucci et al., 2006). Results from
51 epidemiological studies have shown that high consumption of tomato fruit is
52 consistently correlated with a reduced risk of chronic diseases such as cardiovascular
53 disease and certain types of cancer (Sgherri et al., 2008). The increase in the
54 consumers' awareness of the health benefits of fruits and vegetables and the emerging
55 need for convenience due to a fast-paced lifestyle is leading to an increasing demand
56 of fresh-cut fruits and vegetables (Odriozola-Serrano et al., 2008). Retention of the
57 quality and shelf-life of these products during storage is now the interest of the
58 industry and consumers (Camargo et al., 2010). Fresh-cut fruit and vegetables should
59 offer consumers highly nutritious, convenient and healthy food while still maintaining
60 the desired freshness. However, as a result of peeling, cutting and preparation of
61 ready-to-eat fruits and vegetables, a large number of physiological phenomena such as
62 biochemical changes and microbiological spoilage take place and may result in
63 degradation of colour, texture, and flavour. The marketing of fresh-cut vegetables is
64 limited by their short shelf-life due to quick decline in post-processing quality.
65 Chlorinated water (50–200 ppm) is widely used to wash fruits and vegetables as well
66 as fresh-cut produce in order to preserve their quality (Ahmed et al., 2011a, b).
67 However, the possible formation of carcinogenic chlorinated compounds in water
68 (chloramines and trihalomethanes) has called into question the use of chlorine for this
69 purpose (Alegria et al., 2010). In recent years interest is growing in the use of natural
70 products for the preservation of fresh-cut produce. Research and commercial

71 applications have shown that natural components could replace traditional washing
72 agents (Rojas-Graü et al., 2009). The development of chlorine-free fruit and vegetable
73 products enriched with natural bio-products could contribute greatly to a new and
74 growing market, where the consumers' concerns about their health are met.

75 Whey permeate is a by-product of the production of whey protein concentrates from
76 cheese whey. The main composition of whey permeate are water, lactose, low
77 molecular peptides and minerals. Whey and whey ultrafiltrated permeate have been
78 proposed to be used as a natural antioxidant in foods (Contreras et al., 2011). Whey
79 proteins and peptides (Lactoferrin, α -lactalbumin and β -lactoglobulin, casein macro
80 peptide, α_1 - and α_2 - caseins) exhibit a growing number of biological effects including
81 anti-hypertensive, anti-cancer, hypocholesterolemic, opiodergic, and anti-microbial
82 activities (Román et al., 2011; Yalcin, 2006). They are also a rich source of the amino
83 acids - lysine, leucine, threonine, tryptophan and cysteine. Whey could be a promising
84 natural bio-active alternative to chlorine. Martin-Diana et al. (2006) used acidic whey
85 permeate for washing fresh-cut lettuce and carrots during storage. Whey protein has
86 been found to reduce the enzymatic browning of 'Golden Delicious' apples (Perez-
87 Gago et al., 2006). Coronado et al. (2002) successfully used rosemary extract and
88 whey powder for the oxidative stability of wiener sausages during 10 months frozen
89 storage. Whey is used as a fermentation feedstock for the production of lactic acid,
90 acetic acid, propionic acid, ethanol, and single cell protein, etc (Panesar et al., 2007).

91 However, these applications still do not utilise all the whey produced and new uses
92 for this by-product are needed as the high chemical oxygen demand (COD) (50 kg
93 O₂/ton permeate) of whey makes its disposal a cost-effective and significant pollution
94 problem.

95 Therefore the aim of this study was to explore the effect of different application
96 methods of delactosed whey permeate treatment on the quality and antioxidant
97 components of fresh-cut tomatoes during storage as an alternative to chlorine.

98 **2. Materials and Methods**

99 *2.1. Sampling*

100 Irish vine ripened tomatoes (*Lycopersicon esculentum* L. Mill.) cv. Moneymaker were
101 purchased from a local supermarket (Dunnes Stores). According to the grower, the
102 tomato plants were grown commercially in a greenhouse with a 14 h light period from
103 February until November. The aerial environment of the greenhouse and crop
104 irrigation and nutrition were precisely controlled. The temperature of the greenhouse
105 was 16-21 °C which is optimum for lycopene synthesis in tomato fruits. The tomatoes
106 were then brought to the food processing lab and stored at 4 °C before processing.
107 The experiments were carried out between April to September 2010.

108 *2.2. Preparation of Treatment Solution*

109 Liquid delactosed whey permeate was kindly supplied by Glanbia Ltd. Ingredients,
110 Ireland. Delactosed whey permeate (DWP) was obtained after removal of lactose
111 crystals from whey permeate. The pH for DWP solution was 5.0.

112 *2.3. Processing*

113 Whole tomatoes were rinsed in water prior to washing in order to avoid soil
114 contamination. Washing treatment was performed by applying 3 % DWP solution in
115 three different ways such as dipping, spraying and a combination of both. In case of
116 dipping treatment, whole tomato was dipped in DWP solution (200 g tomatoes/L) for
117 1 min (with agitation). In case of spraying treatment, the tomatoes were sliced 6 mm
118 in thickness with a commercial slicing machine (Maxwell chase MCT-25, Baltimore

119 Innovations, UK) and 3 % DWP treatment solution were sprayed over the sliced
120 tomato. In case of the combination of dipping and spraying, tomatoes were treated
121 with 3 % DWP in both ways. For control samples, tomatoes were dipped into 120
122 ppm chlorine solution (pH 8.0) for 1 min (Delaquis et al., 2004). The tomato slices
123 were then air-dried for 30 mins in RT. Processed tomatoes were then pooled, mixed
124 and ~100 grams placed in a polypropylene tray (180 mm length×130 mm width×25
125 mm depth) from Sharp Interpack Ltd., UK containing one layer of absorbent paper on
126 the bottom (Fresh-R-Pax absorbent pads, Maxwell Chase Technologies, Atlanta). The
127 principal ingredient in fresh-R-Pax absorbent pads is food grade sodium
128 carboxymethyl cellulose (CMC), a common ingredient in ice-cream, sauces, low-fat
129 foods, etc. The trays were then packed in bags (200 × 320 mm) of 35 µm oriented
130 polypropylene film (OPP) with permeability at 23 °C and 90 % RH of 3.3×10^{-12}
131 mol/s/m²/Pa for O₂ (Amcor Flexibles Europe-Brighthouse, United Kingdom). The
132 packages were then heat-sealed under atmospheric conditions and stored at 4 °C for
133 10 days (Ahmed et al., 2011a).

134 2.4. *Markers Analysis of Fresh-cut Tomato*

135 Different quality (headspace gas composition, colour, pH and firmness), sensorial and
136 nutritional markers (ascorbic acid, lycopene, total phenols, antioxidant activity as
137 measured by DPPH), enzymatic activity (POD and PME) and microbial enumerations
138 (total aerobic counts and yeast and moulds) of fresh-cut tomato were monitored
139 throughout the 10 days of storage at 4 °C.

140 2.4.1. *Quality Markers*

141 2.4.1.1. *Headspace Gas Composition*

142 Changes in O₂ and CO₂ concentration of the headspaces of the fresh-cut tomatoes
143 packages were monitored during the shelf-life of fresh-cut tomatoes. A Gaspacer
144 analyser (Systech Instruments, UK) was used to monitor O₂ and CO₂ levels. Gas
145 extractions were performed with a hypodermic needle, inserted through an adhesive
146 septum previously fixed to the bags, at a flow rate of 150 ml/min for 10 sec. Three
147 bags per treatment were monitored for each experiment and all bags for other analyses
148 were checked before analysis (Ahmed et al., 2011b).

149 2.4.1.2. *Colour*

150 For colour analysis each piece of tomato in the storage pack was analysed
151 individually to minimise the variability of the product. Colour was quantified using a
152 Colour Quest XE colorimeter (HunterLab, Northants, UK). A tomato slice was placed
153 directly on the colorimeter sensor (3.5 cm of diameter) and measured. 20 – 30
154 measurements were taken per treatment and day. The L* parameter (lightness index
155 scale) range from 0 (black) to 100 (white). The a* parameter measures the degree of
156 red (+a*) or green (-a*) colour and the b* parameter measures the degree of yellow
157 (+b*) or blue (-b*) colour. The CIE L* a* b* parameters were converted to Hue
158 ($\arctan b^*/a^*$) and Chroma $(a^{*2}+b^{*2})^{1/2}$.

159 2.4.1.3. *pH*

160 For pH value 10 g of tomato tissue were blended for 2 min. The pH was measured at
161 room temperature using an Orion research pH-meter, UK.

162 2.4.1.4. *Firmness*

163 Four measurements were made on each slice, two in the outer pericarp and two in the
164 radial pericarp, applying the force in the axial direction. The force necessary to cause
165 a deformation of 3 mm with a speed of 0.02 mm/s was recorded using a an Instron

166 texture analyser (Instron 4302 Universal Testing Machine, Canton MA, USA), with a
167 3.5 mm diameter flat faced cylindrical probe. Only the central slice in the stack was
168 used in the analyses. The firmness measurement was performed immediately after
169 removing the slice from the storage chamber (at storage temperature). Data were
170 analysed with the Instron series IX software for Windows.

171 2.4.2. *Enzymatic Activity*

172 2.4.2.1. *Browning-related Enzyme - Peroxidase (POD)*

173 POD enzyme was assayed in homogenates that were prepared as follows: 10 g of
174 tomato puree was placed in a 100 ml beaker in a 1:2 (w:v) ratio with 0.5 M phosphate
175 buffer, pH 6.5, containing 50 g/l polyvinylpyrrolidone. Then homogenisation was
176 carried out twice with an Ultra-Turrax T-25 tissue homogeniser at 4 °C and 20,500
177 rpm, for 1 min each time with a break of 3 min between homogenisations to avoid
178 excess heating of the sample. The homogenate was centrifuged at 12,720 g for 30 min
179 at 4 °C. It was then filtered through Whatman no. 4 filter paper. The resulting crude
180 extract was used without further purification. All the extracts were kept at 4 °C in the
181 dark and used immediately (within 1 hr). POD activity was assayed
182 spectrophotometrically by a modified method based on Martin-Diana et al. (2006).
183 The reaction mixture contained 0.2 ml of extract and 2.7 ml of 0.05 M phosphate
184 buffer, pH 6.5, containing 1.85 ml of hydrogen peroxide (1.5 %, v/v) as oxidant and
185 3.7 ml of p-phenyldiamine as hydrogen donor. The oxidation of p-phenyldiamine
186 was monitored at 485 nm and 25 °C. A unit of enzyme activity was defined as an
187 increase of 0.1 absorbance units per minute.

188 2.4.2.2. *Texture-related Enzyme - Pectin Methyl Esterase (PME)*

189 PME activity was measured using the method described by Yoo et al. (2009). Ten
190 grams of tomato puree was diluted in an extraction solution (0.2 M sodium phosphate
191 buffer, pH 7.5 containing 1 M sodium chloride and 10 mM dithiothreitol) and
192 homogenised at 4 °C for 2 min at 5,500 rpm. The macerate was incubated at 4 °C for
193 30 min with agitation and centrifuged at 12,720 g for 30 min at 4 °C. It was then
194 filtered through Whatman no. 4 filter paper. One ml of this extract was mixed with 40
195 ml of substrate solution (1 % pectin). The solution was adjusted to pH 7.0 with 1.0 M
196 NaOH, and the pH of the solution was readjusted to pH 7.5 with 0.05 M NaOH. After
197 the pH reached 7.5; 0.2 ml of 0.05 N NaOH was added. The time required to return to
198 pH 7.5 was recorded. Activity was quantified as carboxyl groups formed by the
199 hydrolysis of methyl esters of pectin and was measured titrimetrically using a pH
200 electrode to monitor the production of H⁺. The enzymatic activity was expressed as:

$$201 \quad PME = \frac{0.2[mL]NaOH * 0.05[Mol \cdot L^{-1}]NaOH \cdot X[mL] \cdot 10^3[mMol \cdot Mol^{-1}] \cdot 10^3[L \cdot mL^{-1}]}{Y[mL] \cdot Z[g] \cdot time[min]} \quad (1)$$

202 Where, X = total volume (ml) extracted, Y = volume used (1 ml) in the assay, Z =
203 sample used in the assay (10 g). Three macerates per treatment and day were
204 prepared. Triplicates of the enzymatic activity were analysed.

205 2.4.3. Sensory Analysis

206 A panel of 12 judges aged 20 - 35 years (postgraduate students of the School of Food
207 Science and Environmental Health, DIT) was trained in discriminate evaluation of
208 fresh-cut tomato. Panellists were required to score changes in fresh appearance,
209 texture, aroma and general acceptability. Before starting the sensory experiments,
210 panellists were familiarised with the product and scoring methods. This consisted of
211 demonstration exercises involving examination of fresh-cut tomatoes at different
212 levels of deterioration and agreeing appropriate scores. After becoming familiar with

213 the test facilities and scoring regime, they were invited to score samples. This
214 procedure was repeated several times until a level of consistency in scoring was
215 obtained. Fresh appearance, texture, aroma and general acceptability of tomato
216 samples were scored on a scale of 1 to 9, where a score of one indicated a product of
217 very poor quality, etc. (Ferreira et al., 2008). The evaluation was carried out in the
218 sensory evaluation laboratory. Products were placed in plastic cups with lid, on a
219 white surface and judges were isolated from each-other in a booth in an odour-free
220 environment. Samples were presented in randomised order to minimise possible
221 sequence influence. The results of the sensory analysis were reported as means of
222 three separate trials. Data were analysed using Compusense® Five software (Release
223 4.4, Ontario, Canada).

224 2.4.4. *Nutritional Markers*

225 2.4.4.1. *Ascorbic Acid*

226 The ascorbic acid content in fresh-cut tomatoes was analysed by high performance
227 liquid chromatography (HPLC) following the method described by Lee & Castle
228 (2001) with a slight modification. A 25-ml of 6 % meta-phosphoric acid (pH 3.0) was
229 added to 2.5 g of tomato samples. The sample was homogenised for 1 min at 24,000
230 rpm using an Ultra-Turrax T-25 Tissue homogeniser. Then the sample was shaken
231 with a Gyrotory Shaker G-2 (USA) for 2 hrs at 150 rpm and centrifuged for 15 min at
232 3,000 g at 4 °C (Sanio MSE Mistral 3000ii, UK). Following centrifugation, 10 ml of
233 the supernatant was filtered through PTFE syringe filters (pore size 0.45 µm,
234 Phenomenex, UK) and stored at - 20 °C in foil covered plastic test tubes for further
235 analysis by HPLC. The analysis of ascorbic acid content was performed with Waters
236 600 Satellite HPLC, with a reverse phase analytical polymeric C₁₈ column (150 × 4.6
237 mm, 5 µm) (Waters, Ireland) with a UV-tunable absorbance detector (Waters 486) at

238 230 nm. Ten μ l of the sample was injected. An isocratic mobile phase of 25 mM
239 monobasic potassium phosphate (pH 3.0) with a flow rate of 1.0 ml/min was used.
240 Five concentrations of ascorbic acid standard in 6 % meta-phosphoric acid in the
241 range 10 - 50 μ g/ml were injected.

242 2.4.4.2. *Lycopene*

243 Ten grams of tomato samples were weighed and transferred into a 100 ml beaker
244 (wrapped with aluminium foil). A 50-ml volume of hexane-acetone-ethanol solution
245 (2:1:1 v/v/v) containing 2.5 % BHT was added to solubilise the lycopene (Shi & Le
246 Maguer, 2000). Following this the samples were homogenised with an Ultra-Turrax
247 T-25 tissue homogeniser for 1 min at 20,500 rpm. The samples were then shaken with
248 a Gyrotory Shaker G-2 (USA) for 2 hrs at 150 rpm followed by 10 ml of distilled
249 water was added and stirred for additional 10 min. The polar and non-polar layers
250 were separated, and the upper hexane layer was collected and filtered through a 0.45
251 μ m PVDF membrane filter. It was transferred to a new 15 ml aluminium wrapped test
252 tubes and kept at - 80 °C for analysis. The analysis of lycopene was performed with
253 Waters 600 Satellite HPLC, with a reverse phase analytical polymeric C₁₈ column
254 (150 \times 4.6mm, 5 μ m) (Waters, Ireland) with a UV tunable absorbance detector
255 (Waters 486) for spectrometric peak. The lycopene peaks were identified at 475 nm.
256 An isocratic mobile phase of methyl t-butyl ether/methanol/ethyl acetate (40:50:10,
257 v/v) with a flow rate of 1 ml/min was used. The column temperature and mobile phase
258 was maintained at 25 °C. Analyses were performed under dim light to prevent sample
259 degradation by photo-oxidation. Three concentrations of lycopene standard in the
260 range 0.01 - 0.03 mg/ml were injected.

261 2.4.4.3. *Total Phenols*

262 For extraction of total phenol content 1.25 g of tomato sample was weighed and 25 ml
263 of methanol was added. Following this the sample was homogenised in a 50 ml tube
264 with an Ultra-Turrax T-25 tissue homogeniser for 1 min at 24,000 rpm. The samples
265 were then thoroughly mixed with a vortex mixer (V400 Multitude Vortexer, Alpha
266 laboratories) for 2 hrs at 150 rpm. Then it was centrifuged for 15 min at 3,000 g using
267 a Sanyo MSE Mistral 3000i, UK. Following centrifugation, 10 ml samples of the
268 supernatant were filtered through PTFE syringe filters (pore size 0.45 µm,
269 Phenomenex, UK). Finally the extracts were stored at – 20 °C in foil covered plastic
270 test tubes for further analysis. Total phenol content of tomatoes was determined using
271 the Folin-Ciocalteu method (Singleton et al., 1999). In a 1.5 ml Eppendorf tube, 100
272 µl of appropriately diluted methanolic extract, 100 µl of MeOH and 100 µl of FC
273 reagent were added and vortexed. After exactly 1 min, 700 µl of sodium carbonate
274 (20 %) was added, and the mixture was vortexed and allowed to stand at room
275 temperature in the dark for 20 min. Then the tubes were centrifuged at 12,720 g for 3
276 min. The absorbance of the supernatant was read at 735 nm in 1 ml plastic cuvettes.
277 The blank was MeOH. Each sample of the three batches was measured in triplicate.
278 Results were expressed as mg/L Gallic acid equivalents (GAE).

279 *2.4.4.4. Antioxidant Activity Test - 2, 2-Diphenyl-1-picrylhydrazyl Radical*
280 *Scavenging Capacity Assay (DPPH)*

281 DPPH scavenging activity assay was performed as per the method described by
282 Sanchez-Moreno (2002) with a slight modification. For extraction, 1.25 g of tomato
283 sample was weighed and 25 ml of methanol was added to it. Following this the
284 sample was homogenised in a 50 ml tube with an Ultra-Turrax T-25 tissue
285 homogeniser for 1 min at 24,000 rpm. The sample was then thoroughly mixed with a
286 vortex mixer (V400 Multitude Vortexer, Alpha laboratories) for 2 hrs at 150 rpm.

287 Then the sample was centrifuged for 15 min at 3,000 rpm using a Sanyo MSE Mistral
288 3,000i, UK. Following centrifugation, 10 ml samples of the supernatant were filtered
289 through PTFE syringe filters (pore size 0.45 µm, Phenomenex, UK). Finally the
290 extracts were stored at - 20 °C in foil covered plastic test tubes for further analysis. In
291 a 1.5-ml Eppendorf tube 500 µl of appropriately diluted methanolic extract and 500 µl
292 DPPH Reagent were added and vortexed. After that they were kept for 30 min in
293 dark. The absorbance of the supernatant was read at 515 nm in 1 ml plastic cuvettes.
294 Each sample of the three batches was measured in triplicate.

295 *2.4.5. Microbial Analyses*

296 Microbiology analyses were carried out on the samples before and after the treatment
297 at regular intervals through the storage period. 25 g of tomatoes were blended in 225
298 ml of peptone water (Scharlau Chemie, S.A., Barcelona, Spain) with a Stomacher
299 circulator homogeniser (VWR, Dublin, Ireland). Enumeration and differentiation of
300 total aerobic counts were quantified at 30 °C in plate count agar (Scharlau Chemie,
301 S.A., Barcelona, Spain) over 72 hrs. Yeast and moulds were quantified at 25 °C in
302 potato dextrose agar (Scharlau Chemie, S.A., Barcelona, Spain) over 72 hrs. The
303 results were expressed as log colony forming units per gram (cfu/g).

304 *2.4.6. Statistical Analysis*

305 Data were analysed by multivariate analysis of variance (MANOVA) using
306 Statgraphics software (version: Centurium XV; Statistical Graphics Co., Rockville,
307 USA) for different washing treatments. Analysis of variance one-way (ANOVA) was
308 used to analyse each treatment over storage. In the case of significant differences the
309 LSD range test ($p < 0.05$) was used.

310 **3. Results and Discussion**

311 3.1. *Quality Markers*

312 3.1.1. *Headspace Gas Composition*

313 Fig. 1 illustrates the changes of headspace gas (O₂ and CO₂) composition inside the
314 fresh-cut tomato packages during the 10 days of storage. Both the oxygen and carbon
315 dioxide gas compositions significantly ($p < 0.05$) changed over storage. The oxygen
316 gas decreased and the carbon dioxide gas increased throughout storage, as expected.
317 Oxygen decreased from atmospheric levels (21 % - packaging conditions) to values
318 around 15 % by day 10. A sharp increase in carbon dioxide was observed during
319 storage, from 1.0 % to reaching values around 4.5 % at day 10. The final
320 concentrations for both gases in the fresh-cut tomato packages stored at 4 °C were
321 between the tolerance limits for this commodity which are neither lower than 10 % for
322 oxygen nor higher than 15 % for carbon dioxide (Villanueva et al., 2005). Similar gas
323 concentration levels at day 10 were also described by Artés et al. (1999) using passive
324 MAP where the fresh-cut tomatoes retained good quality. Gil et al. (2002) also found
325 this increase in CO₂ after cutting of fresh tomatoes. There was no significant ($p < 0.05$)
326 difference among the control and the DWP treatments for headspace gas composition
327 as the changing pattern of the gases was the same over time.

328 3.1.2. *Colour*

329 Colour is one of the most important factors in the perception of the quality of fresh-
330 cut fruits and vegetables. In the present study, fresh-cut tomatoes showed a significant
331 decrease ($p < 0.05$) in luminosity during storage (Table 1). Lana et al. (2006) also
332 found a similar trend. There were significant ($p < 0.05$) differences in L* values
333 between samples treated with DWP by spraying and samples treated with chlorine.
334 The other two DWP treatments did not show significant difference with the chlorine

335 treatment. The parameters a^* and b^* were not significantly ($p>0.05$) affected by the
336 DWP treatment. The parameter a^* increased significantly ($p<0.05$) during storage.
337 The a^* value is an important parameter for red colour development and the degree of
338 ripening in tomato. Lana et al. (2006) also found the increase of a^* values during
339 storage. However, the parameter b^* decreased during storage for all treatments. Hue
340 and Chroma also decreased during storage and the decrease was more prominent in
341 samples treated with chlorine than samples treated with DWP, though not significant.
342 Hue is negatively correlated with the maturity of tomato. Hue decreases as tomato
343 matures during storage.

344 3.1.3. pH

345 Significantly ($p<0.05$) higher pH was observed for the control samples (chlorine) than
346 samples treated with DWP (Fig. 2A). At day 10 the lowest pH appeared in samples
347 washed with DWP by a combination of dipping and spraying method, followed by
348 spraying only. Tomatoes treated with DWP by dipping had the highest pH among the
349 three DWP treatments, though the difference with the tomatoes treated with DWP by
350 spraying was not significant ($p>0.05$). The pH increased significantly ($p<0.05$) over
351 storage in all the samples. This is in agreement with Artés et al. (1999). The increase
352 in pH during storage might be associated with bacterial growth (Rico et al., 2007).

353 3.1.4. Texture

354 Texture (firmness) decreased significantly ($p<0.05$) during storage for all treatments
355 (Fig. 2B). These instrumental result correlated with the sensory panel's texture scores
356 (Fig. 3). All three DWP treated samples maintained significantly ($p<0.05$) better
357 texture than chlorine treated samples. Significant differences ($p<0.05$) among DWP
358 treatments were also observed. Samples treated with DWP by a combination of

359 dipping and spraying method maintained significantly higher texture throughout the
360 storage than samples treated with DWP dipping and DWP spraying. There was no
361 significant ($p>0.05$) difference between the samples treated with DWP dipping and
362 DWP spraying. The presence of calcium in whey permeates might help to maintain
363 the firmness of tomato during storage (Evans et al., 2010). Different calcium salts
364 have been used for firmness improvement of fresh fruits and vegetables. Calcium
365 carbonate and calcium citrate are the main calcium salts added to foods in order to
366 enhance the nutritional value. Calcium chloride has been widely used as preservative
367 and firming agent in the fruits and vegetables industry for whole and fresh-cut
368 commodities (Chardonnet et al., 2003).

369 3.2. *Enzymatic Activity*

370 3.2.1. *Browning-related Enzyme- Peroxidase (POD)*

371 The data for POD activity of fresh-cut tomato showed that samples treated with DWP
372 had significantly ($p<0.05$) lower activity compared to chlorine treated samples (Fig.
373 2C). Significant differences ($p<0.05$) in POD activity were observed among the DWP
374 treatments. Samples treated with DWP by a combination of dipping and spraying
375 method and DWP spraying showed significantly ($p<0.05$) lower POD activity than
376 samples treated with DWP dipping throughout the storage. The decreased activity of
377 POD in DWP treated samples might be associated with the potential antioxidant
378 activity of the whey permeate used (Peña-Ramos & Xiong, 2003). A significant
379 ($p<0.05$) increase in the activity in all the samples was observed during storage
380 regardless of the treatments, although showing little fluctuations over storage. The
381 initial increase at day 3 might be due to mechanical stress during minimal processing
382 (Cantos et al., 2001). The depletion of antioxidants at day 10 might be attributed to
383 the sharp increase of POD activity in whey treated samples in the period of day 7 to

384 10. Edible composite coatings prepared from whey protein concentrate and beeswax
385 with ascorbic acid or 0.5 % cystine reduced the enzymatic browning of ‘Golden
386 Delicious’ apples (Perez-Gago et al., 2006).

387 3.2.2. *Texture-related Enzyme- Pectin Methyl Esterase (PME)*

388 Significant differences ($p < 0.05$) were observed in PME activity among the samples
389 treated with DWP and chlorine (Fig. 2D). Samples treated with chlorine showed
390 significantly ($p < 0.05$) lower PME activity than DWP treated samples. There was no
391 significant difference ($p > 0.05$) among the DWP treated samples. PME activity
392 increased significantly ($p < 0.05$) during storage for all treatments. Other authors have
393 attributed that the variability of intrinsic factors and the pre- and postharvest factors
394 can affect enzyme activity, vitamin content, etc of the samples (Yoo et al., 2009).
395 Similar behaviour has been observed for certain browning related enzymes (Perez-
396 Gago et al., 2006).

397 3.3. *Sensory Analysis*

398 Significant differences ($p < 0.05$) for fresh appearance, aroma, texture and general
399 acceptability scores were observed between samples treated with DWP and chlorine
400 (Fig. 3). DWP treated fresh-cut tomatoes scored significantly higher ($p < 0.05$) than
401 chlorine treated samples. Among the samples treated with DWP, the combination of
402 dipping and spraying scored the highest, followed by the spraying only. Lowest scores
403 of fresh appearance, aroma, texture and general acceptability were observed in
404 samples treated with DWP by dipping only among the three DWP treated samples.
405 This was in agreement with most of the physico-chemical markers of fresh-cut
406 tomatoes studied in the current research. All the attributes evaluated (such as, texture,
407 aroma, first impression, general acceptability) decreased significantly ($p < 0.05$) during

408 storage which is associated with a loss of quality. However, the values at the end of
409 the storage (10 days) were still above the acceptability threshold of 5 for all the
410 attributes scored (Ferreira et al., 2008). The non-hypoxic O₂ and CO₂ concentration in
411 the packages might have helped to maintain acceptable levels of colour and aroma
412 (Aguayo et al., 2006). Nykänen et al. (1998) analysed the effect of nisin-whey
413 permeate washing solutions on total counts and sensory characteristics in rainbow
414 trout. They found that nisin-whey treatment caused no negative effect on sensory
415 attributes.

416 3.4. *Nutritional Markers*

417 3.4.1. *Ascorbic Acid*

418 Ascorbic acid decreased significantly ($p < 0.05$) during 10 days of storage for all
419 treatments (from 19 mg/100 g FW to 15 mg/100 g FW). This result is in agreement
420 with Toor & Savage (2005). Significant differences ($p < 0.05$) were observed in
421 ascorbic acid content among the samples treated with DWP and chlorine (Fig. 4A).
422 Samples treated with chlorine retained the lowest ascorbic acid over storage. Samples
423 treated with DWP by a combination of dipping and spraying had significantly
424 ($p < 0.05$) higher ascorbic acid content than the other two DWP treated samples after
425 10 days of storage. However, there was no significant difference ($p > 0.05$) between the
426 samples treated with DWP by dipping and by spraying. Ascorbic acid contributes 28–
427 38 % to the antioxidant activity, while the remaining activity is mainly due to
428 phenolics in tomatoes (Toor & Savage, 2005). Phenolic substances have been
429 reported to have a protective effect on the ascorbic acid.

430 3.4.2. *Lycopene*

431 The average amount of lycopene in the tomato samples was 5.7 mg/100 g FW. The
432 treatments did not show a significant effect ($p < 0.05$) on the lycopene concentration of
433 the samples during storage (Fig. 4B). Samples treated with chlorine retained the
434 lowest lycopene over storage. The highest lycopene content was observed in samples
435 treated with DWP by a combination of dipping and spraying, though the difference
436 was not significant ($p > 0.05$). However, storage time had significant effect ($p < 0.05$) on
437 the tomato samples. The lycopene content increased during storage. This is because
438 fruits biosynthesise carotenoids during ripening throughout storage time (Odrizola-
439 Serrano et al., 2008). On the other hand, Shi & Le Maguer (2000) observed that
440 carotenoids are susceptible to oxidation in the presence of light, oxygen and low pH.
441 Therefore, the increase in the lycopene concentration during 10 days of storage might
442 be due to the biosynthesis of lycopene induced by ripening and the low oxidation of
443 this carotenoid as a result of low availability of O_2 in the package headspace.

444 3.4.3. Total Phenols

445 The total phenol content of tomato samples differed significantly over storage time
446 (Fig. 4C). Chlorine treated samples showed significantly ($p < 0.05$) lower phenolic
447 content than DWP treated samples. All three DWP treated tomato samples maintained
448 similar level of phenolic content during storage. The total phenol content decreased
449 significantly ($p < 0.05$) during storage irrespective of treatments. This was in
450 accordance with other studies (Toor & Savage, 2005; Gil et al., 2002). The decrease
451 was slow until day 3. After this, all samples demonstrated a rapid decrease in the total
452 phenolic content. Chlorine treated samples decreased the most to a value of around
453 19.8 mg GAE/100 g FW from 21.0 mg GAE/100 g FW over 10 days of storage.
454 Phenolics are the major antioxidant compounds in plant extracts. Toor & Savage

455 (2005) reported that phenolic compounds might contribute 60 to 70% antioxidant
456 activity of tomato extracts.

457 3.4.4. Antioxidant Activity Test - 2, 2-Diphenyl-1-picrylhydrazyl Radical 458 Scavenging Capacity Assay (DPPH)

459 The antioxidant capacity as measured by DPPH radical scavenging activity differed
460 significantly ($p < 0.05$) between treatments (Fig. 4D). All three whey permeate treated
461 samples showed significantly ($p < 0.05$) higher DPPH reduction than chlorine treated
462 samples. The higher antioxidant activity of whey permeates treated samples could be
463 associated with the intrinsic antioxidant activity of whey permeates (Ahmed et al.,
464 2011a). Whey permeates might have also helped to retain the antioxidant activity of
465 tomato slices. These results could be related to the total phenolic content of the
466 samples since the samples containing higher phenolic content exhibited stronger
467 DPPH reduction and vice versa. There was no significant difference ($p > 0.05$) between
468 the samples treated with DWP by a combination of dipping and spraying and spraying
469 only. Samples treated with DWP by dipping only had the lowest DPPH reduction
470 among the three DWP treatments. On the other hand, the antioxidant capacity of
471 fresh-cut tomatoes depleted with storage time irrespective of the treatments. The
472 antioxidant activity was reduced on an average $\sim 4\%$ over 10 days of storage at $4\text{ }^{\circ}\text{C}$.

473 3.5. Microbial Enumerations

474 3.5.1. Total Aerobic Counts

475 Fresh-cut tomatoes stored at $4\text{ }^{\circ}\text{C}$ had initial loads of total aerobic counts
476 approximately $\sim 4.5\text{ log cfu/g}$. The DWP treated samples had significant difference
477 ($p < 0.05$) of bacterial growth to those treated with chlorine (Fig. 5A). Samples treated
478 with DWP by a combination of dipping and spraying showed the highest reduction (\sim

479 1.0 log cfu/g) of total counts than samples treated with chlorine after 10 days of
480 storage. Samples treated with DWP by dipping only and spraying only also showed
481 significantly ($p<0.05$) better reduction in total aerobic counts (~ 0.6 log cfu/g) than
482 samples treated with chlorine after 10 days of storage. In fresh-cut tomatoes, total
483 counts increased during storage for all the washing treatments. This increase was
484 more obvious between days 7 and 10. The values of DWP treated samples at the end
485 of the storage were lower than the recommended 10^8 cfu/g for consumer consumption
486 of fresh-cut vegetables (Alegria et al., 2010).

487 3.5.2. *Yeast and Moulds*

488 The initial loads of yeast and moulds of fresh-cut tomatoes stored at 4 °C was
489 approximately 4 log cfu/g. This result was in agreement with the finding of Prakash et
490 al. (2002) for diced tomato. Chlorine treated samples showed the highest yeast and
491 moulds growth (~ 7.0 log cfu/g, day 10). DWP treated tomatoes showed significantly
492 ($p<0.05$) lower counts than chlorine treated samples (Fig. 5B). Samples treated with
493 DWP by a combination of dipping and spraying showed the highest reduction (~ 1.2
494 log cfu/g) of yeast and moulds counts than samples treated with chlorine after 10 days
495 of storage. DWP treated samples by spraying also showed significantly ($p<0.05$)
496 better reduction in yeast and moulds counts (~ 0.8 log cfu/g) than samples treated
497 with chlorine after 10 days of storage. Yeast and moulds increased significantly
498 ($p<0.05$) for all the washing treatments with storage time.

499 The presence of antimicrobial peptides in the whey permeate might contribute to its
500 antimicrobial capacity (Clare & Swaisgood, 2003). Antimicrobial peptides have been
501 identified from whey protein hydrolysates. The most studied are the lactoferrins, α_{S1} -
502 casein and α_{S2} -casein (McCann et al., 2006). These antimicrobial peptides act against
503 different gram-positive and gram-negative bacteria (*Escherichia*, *Helicobacter*,

504 *Listeria*, *Salmonella* and *Staphylococcus*), yeasts and filamentous fungi (Rizzello et
505 al., 2008; Fitzgerald & Murray, 2006). The amphipathic nature of these peptides
506 presumably underlies their biological activities which enables them to associate with
507 lipid membranes and disrupt normal membrane functions of bacteria. The mechanism
508 of action has been investigated for whey antimicrobial peptides by Saint-Sauveur et
509 al. (2008).

510 **4. Conclusion**

511 The results showed that the use of delactosed whey permeate (DWP) is a viable
512 alternative to chlorine in controlling the microbiota associated with the quality
513 deterioration of fresh-cut tomatoes, since the growth of total aerobic counts and yeasts
514 and moulds were substantially inhibited by its application. Moreover, DWP treated
515 samples retained the antioxidant compounds better during storage than the chlorine
516 treated samples. The three application methods of DWP differed significantly for
517 extending the shelf-life of fresh-cut tomatoes. The combination of dipping and
518 spraying of DWP showed the best results for all the markers tested. Further research
519 on antimicrobial and antioxidant properties of DWP is recommended.

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648

649 **Figure Captions**

650 **Fig. 1.** Effect of treatments on headspace gas composition O₂ (A) and CO₂ (B) in
651 fresh-cut tomato packages over 10 days storage at 4 °C. Points designated on any
652 curve by different letters are significantly different (p<0.05). Lower case letters are
653 used for comparisons during storage and upper case letters for treatment comparisons.
654 Three independent trials were carried out in triplicate.

655 **Fig. 2.** Effect of DWP and chlorine treatments on pH (A), texture (B), POD (C) and
656 PME (D) in fresh-cut tomato packages over 10 days storage at 4 °C. Points designated
657 on any curve by different letters are significantly different (p<0.05). Lower case
658 letters are used for comparisons during storage and upper case letters for treatment
659 comparisons. Three independent trials were carried out in triplicate.

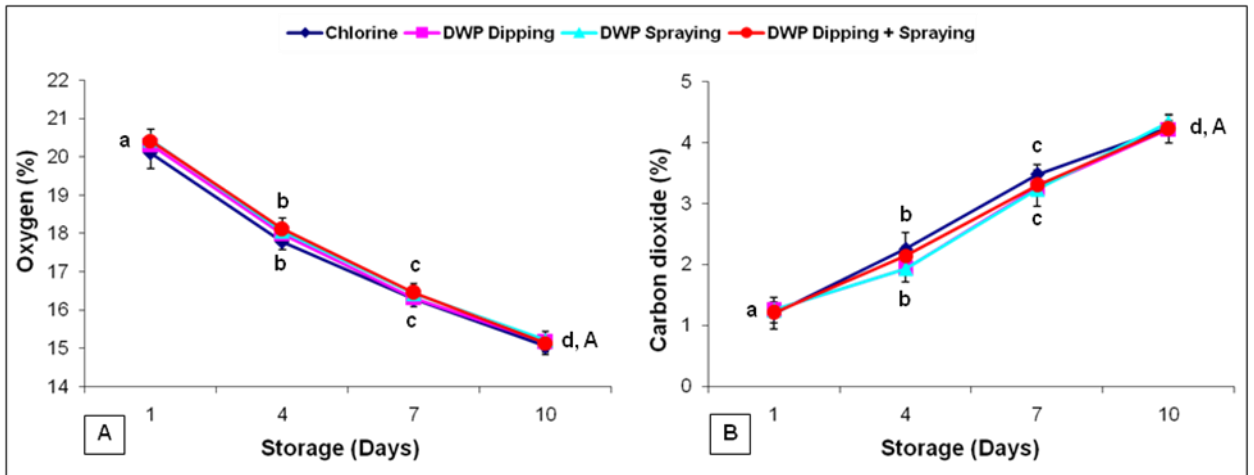
660 **Fig. 3.** Sensory evaluation of fresh-cut tomatoes packaged and stored for 10 days at 4
661 °C and treated with DWP and chlorine.

662 **Fig. 4.** Ascorbic acid (A), lycopene (B), total phenols (C) and antioxidant activity -
663 DPPH (D) in fresh-cut tomatoes treated with DWP and chlorine during the 10 days of
664 storage at 4 °C. Points designated on any curve by different letters are significantly
665 different (p<0.05). Lowercase letters are used for comparisons during storage and
666 uppercase letters for treatment comparisons. Three independent trials were carried out
667 in triplicate.

668 **Fig. 5.** Effect of washing treatments on total aerobic counts (A) and yeast and moulds
669 (B) during 10 days storage of fresh-cut tomato at 4 °C. Points designated on any curve
670 by different letters are significantly different (p<0.05). Lowercase letters are used for
671 comparisons during storage and uppercase letters for treatment comparisons. Three
672 independent trials were carried out in triplicate.

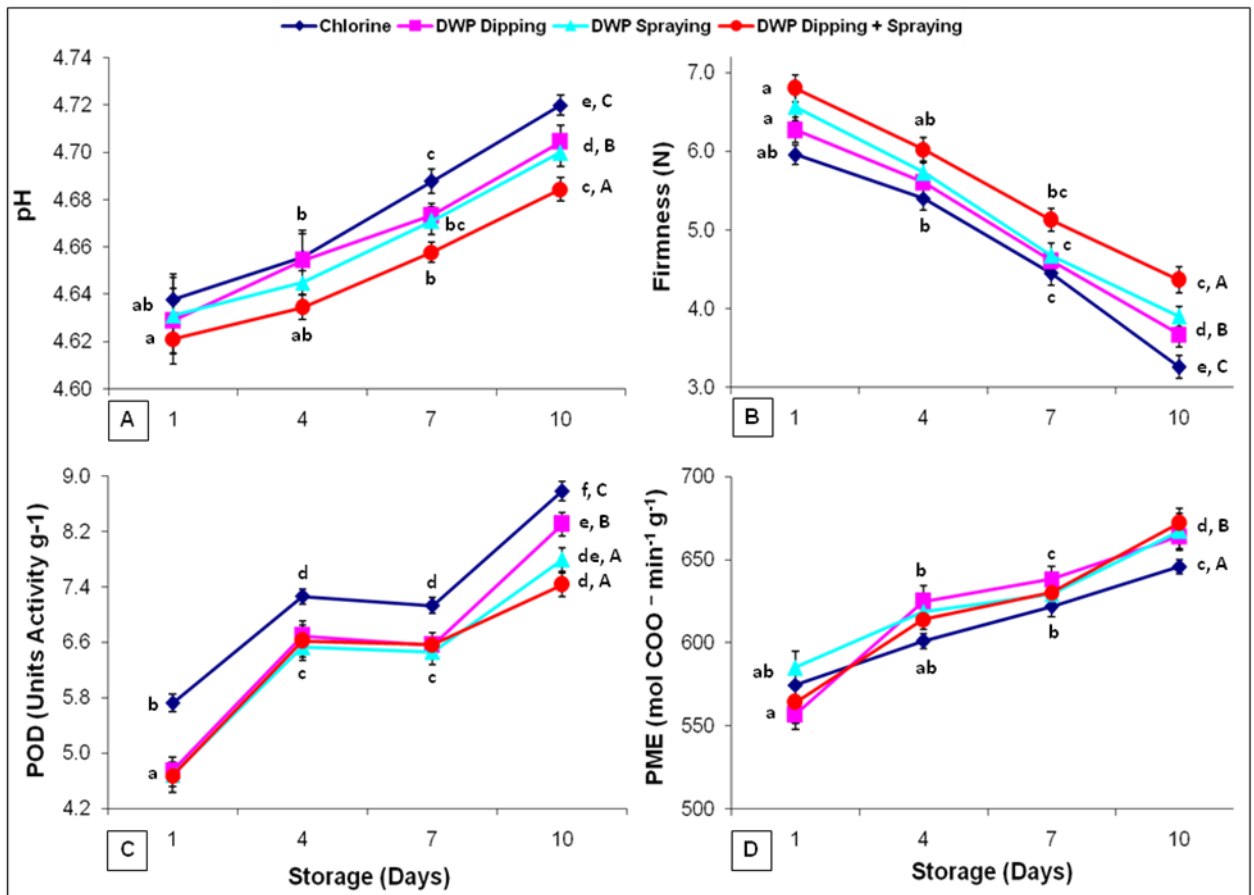
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Fig. 1



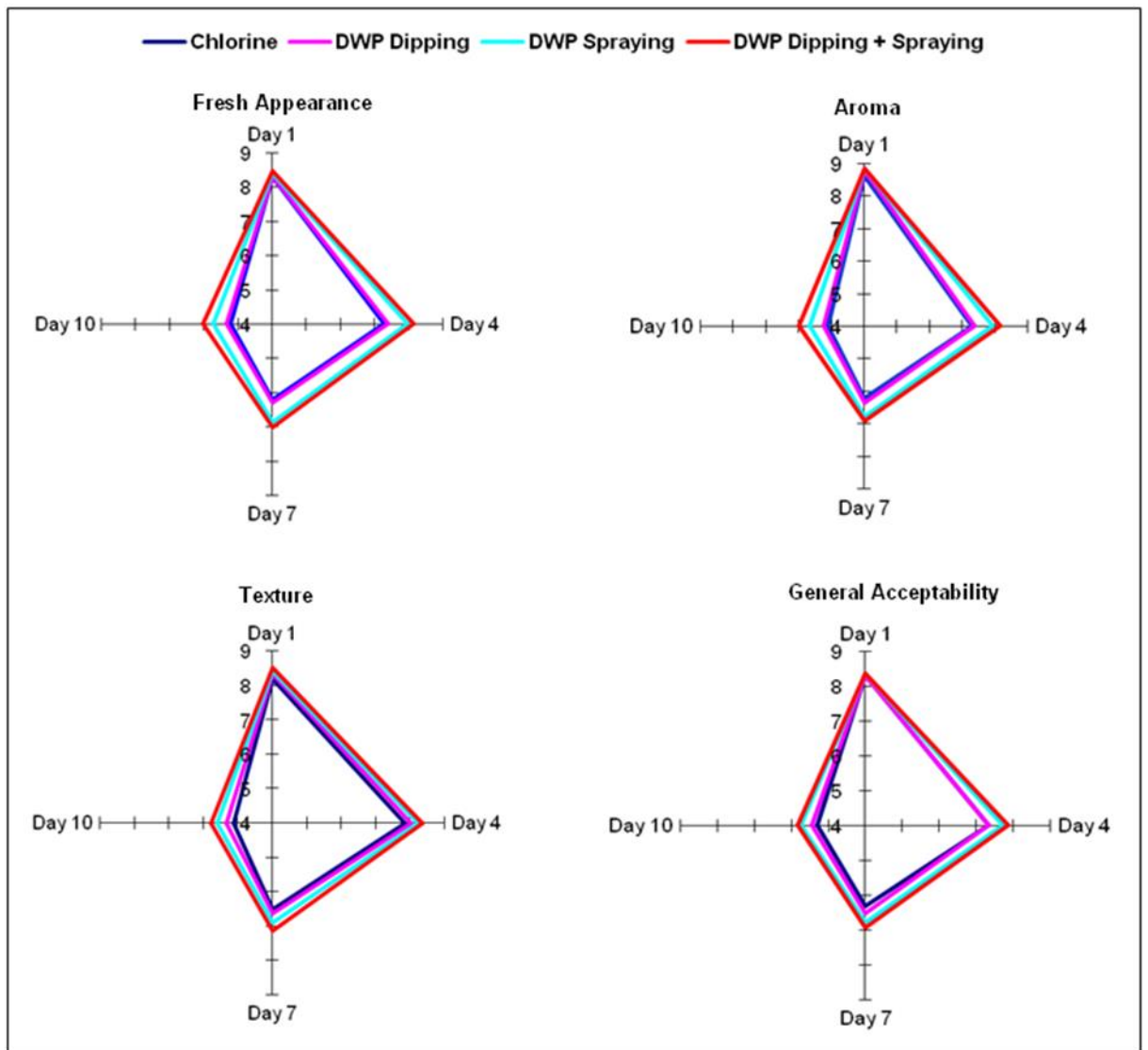
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Fig. 2



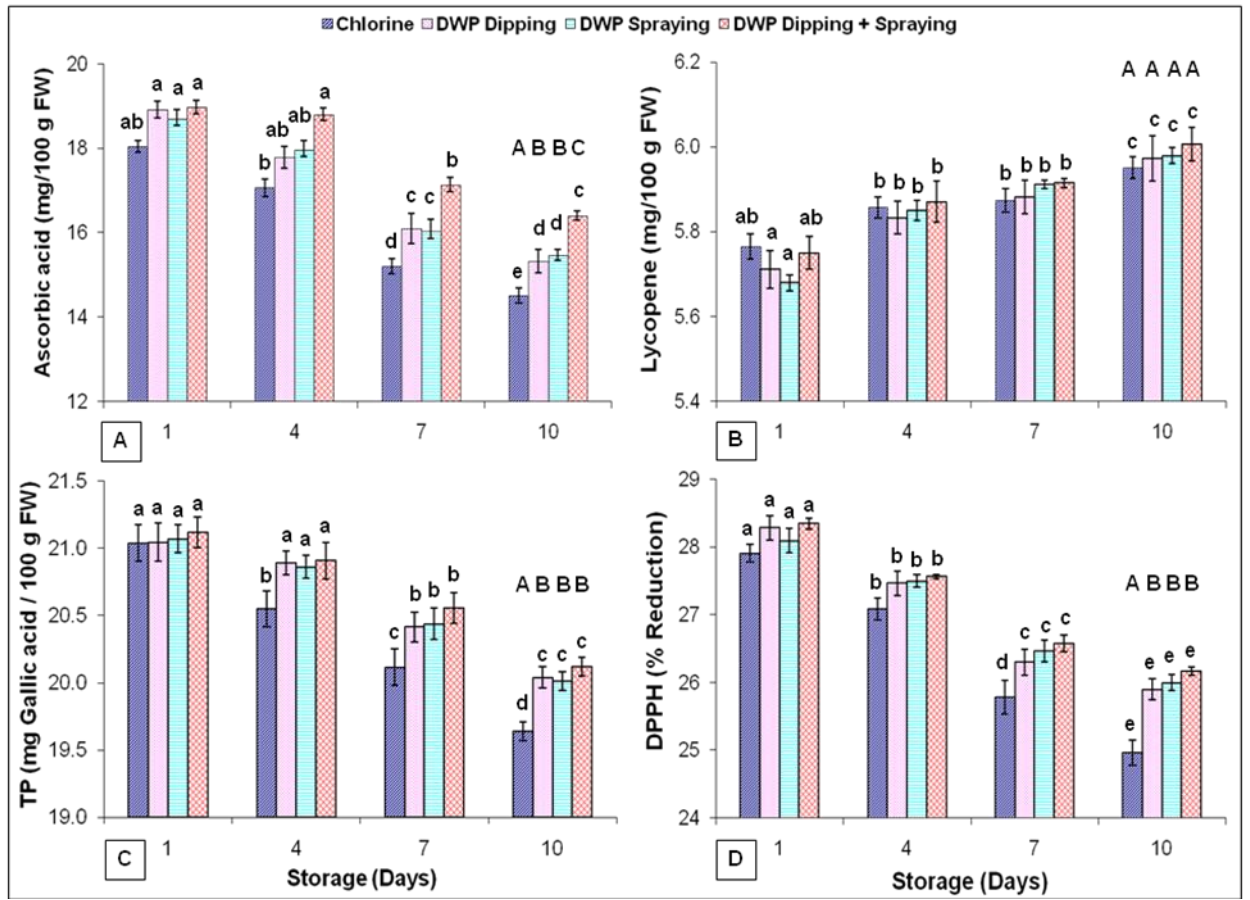
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Fig. 3



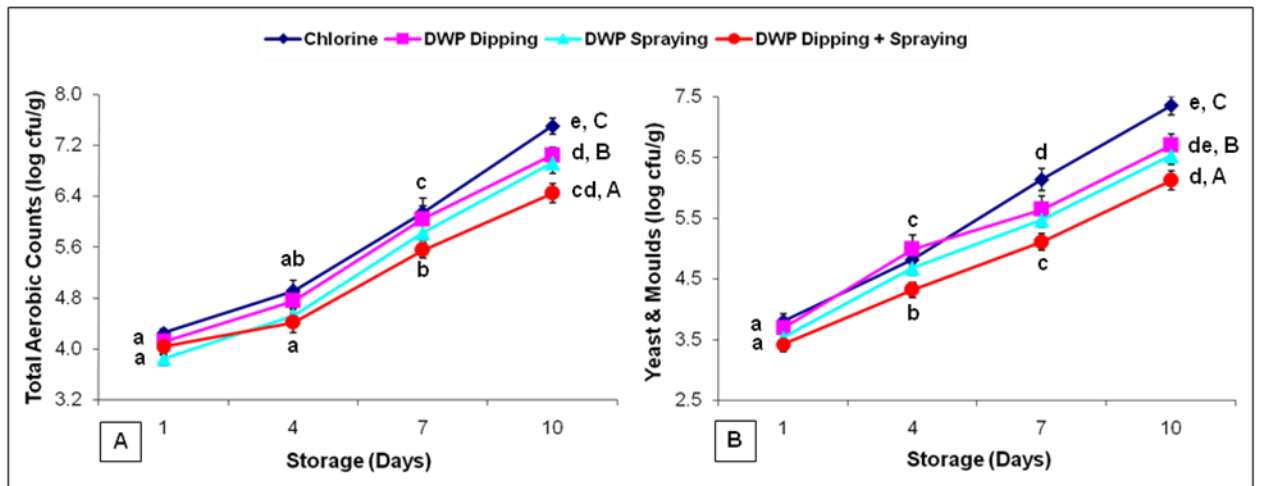
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Fig. 4



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Fig. 5



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