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## Co-colonisation with *Aspergillus Fumigatus* and *Pseudomonas Aeruginosa* Is Associated With Poorer Health In Cystic Fibrosis Patients: an Irish registry analysis

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RESEARCH ARTICLE

Open Access



# Co-colonisation with *Aspergillus fumigatus* and *Pseudomonas aeruginosa* is associated with poorer health in cystic fibrosis patients: an Irish registry analysis

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## Abstract

**Background:** Pulmonary infection is the main cause of death in cystic fibrosis (CF). *Aspergillus fumigatus* (AF) and *Pseudomonas aeruginosa* (PA) are the most prevalent fungal and bacterial pathogens isolated from the CF airway, respectively. Our aim was to determine the effect of different colonisation profiles of AF and PA on the clinical status of patients with CF.

**Methods:** A retrospective analysis of data from the Cystic Fibrosis Registry of Ireland from 2013 was performed to determine the effect of intermittent and persistent colonisation with AF or PA or co-colonisation with both microorganisms on clinical outcome measures in patients with CF. Key outcomes measured included forced expiratory volume in one second (FEV<sub>1</sub>), number of hospitalisations, respiratory exacerbations and antimicrobials prescribed, and complications of CF, including CF related diabetes (CFRD) and allergic bronchopulmonary aspergillosis (ABPA).

**Results:** The prevalence of AF and PA colonisation were 11% (5% persistent, 6% intermittent) and 31% (19% persistent, 12% intermittent) in the Irish CF population, respectively. Co-colonisation with both pathogens was associated with a 13.8% reduction in FEV<sub>1</sub> ( $p = 0.016$ ), higher levels of exacerbations ( $p = 0.042$ ), hospitalisations ( $p = 0.023$ ) and antimicrobial usage ( $p = 0.014$ ) compared to non-colonised patients and these clinical outcomes were comparable to those persistently colonised with PA. Intermittent and persistent AF colonisation were not associated with poorer clinical outcomes or ABPA. Patients with persistent PA had a higher prevalence of CFRD diagnosis ( $p = 0.012$ ).

**Conclusions:** CF patients co-colonised with AF and PA had poor clinical outcomes comparable to patients persistently colonised with PA, emphasising the clinical significance of co-colonisation with these microorganisms.

**Keywords:** Co-colonisation, Cystic Fibrosis, *Aspergillus fumigatus*, *Pseudomonas aeruginosa*, Pulmonary function

## Background

Cystic fibrosis (CF) is the most common inherited life shortening condition affecting Caucasians. It is estimated that there are over 70,000 people living with CF worldwide [1]. It is a multi-organ disease however up to 95% of morbidity and mortality is caused by airway infections

and the associated inflammation [2]. Mutations in the CF transmembrane conductance regulator (CFTR) gene result in dysfunctional or absent CFTR protein which ultimately results in thick airway mucus and impaired mucocilliary clearance. This provides the perfect environment for microorganisms to colonise and persist. The path of disease progression is established early in life with deterioration in lung function beginning as early as age 6 [3]. The recurrent onslaught of airway infection and inflammation results in reduced lung function and eventually respiratory failure.

Airway infection management is the cornerstone of CF care and it is now widely accepted that the CF airways

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are colonised with a community of microorganisms from an early age [2–4]. Microorganisms previously not considered to be significant in CF microbiology are emerging as having roles in disease progression [4, 5]. The discovery of complex and diverse microbial communities in the airways is forcing a paradigm shift in the treatment of airway infections. Considering this, research into coinfections and polymicrobial infections is imminent. Here we aimed to determine the clinical implications of coinfection with the most common fungal and bacterial pathogen isolated from the CF airways.

*Aspergillus fumigatus* (AF) is the most common fungal pathogen isolated from the CF airways with a reported prevalence between 16 and 58% [6–10]. AF rarely causes invasive infections in CF however it can cause allergic bronchopulmonary aspergillosis (ABPA), a hypersensitivity reaction to AF with detrimental consequences on airway inflammation. Generally intermittent or persistent AF colonisation is not treated unless ABPA is confirmed. Criteria for diagnosis of ABPA are based on the CF Foundation consensus [10]. At present there is no consensus for the diagnosis or treatment of asymptomatic AF colonisation in CF. Recent studies have utilized a combination of AF qPCR, galactomannan, AF specific IgE and IgG to decipher the complexity of AF colonisation status in CF patients. Classifying patients into proposed subgroups and delineating those with ABPA from those who are AF sensitised, AF non-sensitised and those with aspergillus bronchitis [11]. Some studies have provided valuable insights into how AF may influence the progression of CF lung disease [9, 12, 13] however the true extent of the impact on disease progression is unknown.

*Pseudomonas aeruginosa* (PA) is the most common bacterial pathogen in CF [14] and in 2013, the Cystic Fibrosis Foundation in the US reported 48.7% of patients were colonised with PA [15]. Early PA colonisation of the CF airway is frequently intermittent and eradication is possible using inhaled antibiotics. However re-colonisation can occur and in some cases despite intense efforts to re-eradicate the organism, the colonisation becomes persistent and a decline in lung function ensues [16, 17]. The transition from intermittent to chronic or persistent PA colonisation is a seminal clinical event in a CF patient's life and persistent PA colonisation is associated with increased mortality [18, 19].

Despite substantial improvement in median survival, CF remains a life-shortening disease with the mean age of death in Ireland being only 22.8 years [20] and pulmonary insufficiency is the main cause of death. There is a paucity of published data on the prevalence of persistent AF colonisation or AF and PA co-colonisation in CF patients. We aimed to determine the prevalence of persistent and intermittent colonisation with PA or AF and co-colonisation with both pathogens in the Irish CF

population. Furthermore we aimed to determine the association of these different colonisation patterns with the clinical status of patients with CF.

## Methods

### Study population

Data from patients with CF registered in the Cystic Fibrosis Registry of Ireland (CFRI) database in 2013 was used in this study. The number of live patients registered with the CFRI was 1158 on the last day of 2013 (92.5% of the Irish CF population) and data from 749 patients was included in this study (409 patients with incomplete data sets were excluded). A retrospective cohort study was carried out to establish whether colonisation with AF and/or PA impacts on patient health. The primary clinical outcome measure was forced expiratory volume in one second percent predicted (FEV<sub>1</sub>%) and secondary clinical outcome measures were the number of hospitalisations, respiratory exacerbations (treated with intravenous antibiotics), prescribed antimicrobials and complications of CF, including pancreatic insufficiency resulting in CFRD and ABPA. CFRD diagnosis was defined as being treated with insulin for CF-related diabetes, ABPA diagnosis was reported by the physician.

Sputum ( $n = 1859$ ), throat swabs ( $n = 31$ ), cough swabs ( $n = 52$ ), nasal swab ( $n = 1$ ) and bronchoalveolar lavage ( $n = 10$ ) samples were recorded as positive for PA and/or AF by the individual hospitals. We employed stringent criteria to classify colonisation status: greater than 2 but less than 4 respiratory samples positive within the year was considered intermittent colonisation and greater than 4 samples positive within the year was considered persistent colonisation, in line with the Leeds criteria [21]. Patients were separated into 6 cohorts depending on their colonisation status: 1) intermittent PA colonisation (PA), 2) persistent PA colonisation (PAp), 3) intermittent AF colonisation (AF), 4) persistent AF colonisation (AFp), 5) colonised with both pathogens (AF + PA) and 6) negative for both pathogens (clear). Clinical outcome measures were compared between our six cohorts.

### Statistical analysis

D'Agostino and Pearson omnibus normality test was performed on FEV<sub>1</sub>% predicted data. FEV<sub>1</sub> data was not normally distributed and outcomes comparisons between groups were made using the Kruskal-Wallis test with Dunn's multiple comparisons. In order to adjust for the potential confounders: age, gender and CFTR mutation, linear regression was used for FEV<sub>1</sub>% predicted and logistic regression models were used for steroid use, ABPA and CFRD positivity yielding Odds Ratios (ORs) for the "poorer" outcomes.

The number of hospitalisations, respiratory exacerbations and the number of antimicrobials were modelled with a Poisson or Negative Binomial regression. The degree of

zero-inflation and over dispersion was assessed by comparing the model Pearson statistic to the sample size for each outcome. In all cases a zero-inflated Negative Binomial model (ZINB) provided the best-fit model yielding Incident Rate Ratios (IRRs). Following this analysis, post hoc tests of all pair-wise comparisons were conducted using a Bonferroni correction for 6 tests (Bonferroni adjusted  $p$ -value = 6 ×  $p$ -value), and Bonferroni-adjusted 95% confidence intervals (99.583% confidence required). Significance level was assumed at below 0.05. Analyses were conducted in IBM SPSS Statistics version 20 and SAS for Windows version 9.3. Detailed description of data analysis protocols is provided in Additional file 1. All  $p$  values shown are after adjustment for age, sex and CFTR mutation unless otherwise stated.

## Results and Discussion

### Demographics of the patient population

The male cohort made up the bigger subset of the CF population at 58.6%, reflecting higher mortality rates in females in adolescence and early adulthood [22]. The age distribution in our dataset was 4–69 years and the median age was 18.1 years. The demographic and clinical data of the cohorts are depicted in Table 1. As expected the FEV<sub>1</sub> percent predicted was lower in older patients reflecting a gradual decline with age (Fig. 1).

### Prevalence of AF and PA colonisation in the Irish CF population

The prevalence of AF and PA co-colonisation was 3.1% in the Irish CF population. The AF + PA group included AFp + PA ( $n = 4$ ), AF + PA ( $n = 5$ ), AF + PAp ( $n = 7$ ) and AFp + PAp ( $n = 8$ ) subgroups. No significant differences in FEV<sub>1</sub>, number of hospitalisations, number of respiratory exacerbations and antimicrobial prescribing were noted between these subgroups (Additional file 2: Table S1). AF colonisation was most prevalent in pre-adolescents and adolescents (Fig. 1), which contradicts current opinion that AF is more commonly an organism encountered in adulthood. To our knowledge this has not been previously reported. The prevalence of AF colonisation in the Irish CF population was 11%, of which 5% had persistent and 6%

intermittent colonisation (Table 1). Previously AF prevalence rates in CF have been reported to be up to 58% using sputum culture [8]. PA becomes more prevalent with age and FEV<sub>1</sub> decline, corroborating previous findings (Fig. 1) [23]. The prevalence of PA in the Irish CF population was 31%, of which 19% had persistent and 12% had intermittent colonisation (Table 1). The prevalence of PA colonisation in Ireland is lower than the 48.7% prevalence reported in the US [17]. Employing similar criteria for the classification of persistent and intermittent PA colonisation as our study, Lee et al., (2003) [21] determined the prevalence of persistent PA to be 18% consistent with our data and intermittent colonisation to be 34%, which is higher than our findings. Proesmans et al., (2006) [24] determined the prevalence of persistent and intermittent PA colonisation to be 31 and 18% respectively which are higher than our prevalence rates. Many of these studies differ in the criteria employed for categorising colonisation status making comparisons between them difficult. We employed stringent criteria to classify colonisation status in line with the Leed's criteria [21] as outlined in our methods. The Leed's criteria have been independently evaluated [24] and are the classification criteria recommended in the European CF Society patient registry guidelines [25]. We did not consider positivity of AF and PA in one sputum sample in a year as intermittent, as other studies have done [6–8, 26], to rule out inclusion of chance events, false positives or sputum contamination. For these reasons our prevalence for AF and PA colonisation may be lower than reported elsewhere [6–9, 15, 21, 24, 26]. Consensus criteria for the definition of intermittent and persistent AF colonisation, comparable to the Leeds criteria for PA, need to be formulated. We acknowledge that a limitation of this study is its cross-sectional design and multi-year analysis of registry data would provide further detail on transition from intermittent to persistent colonisation status and the impact on disease outcome measures.

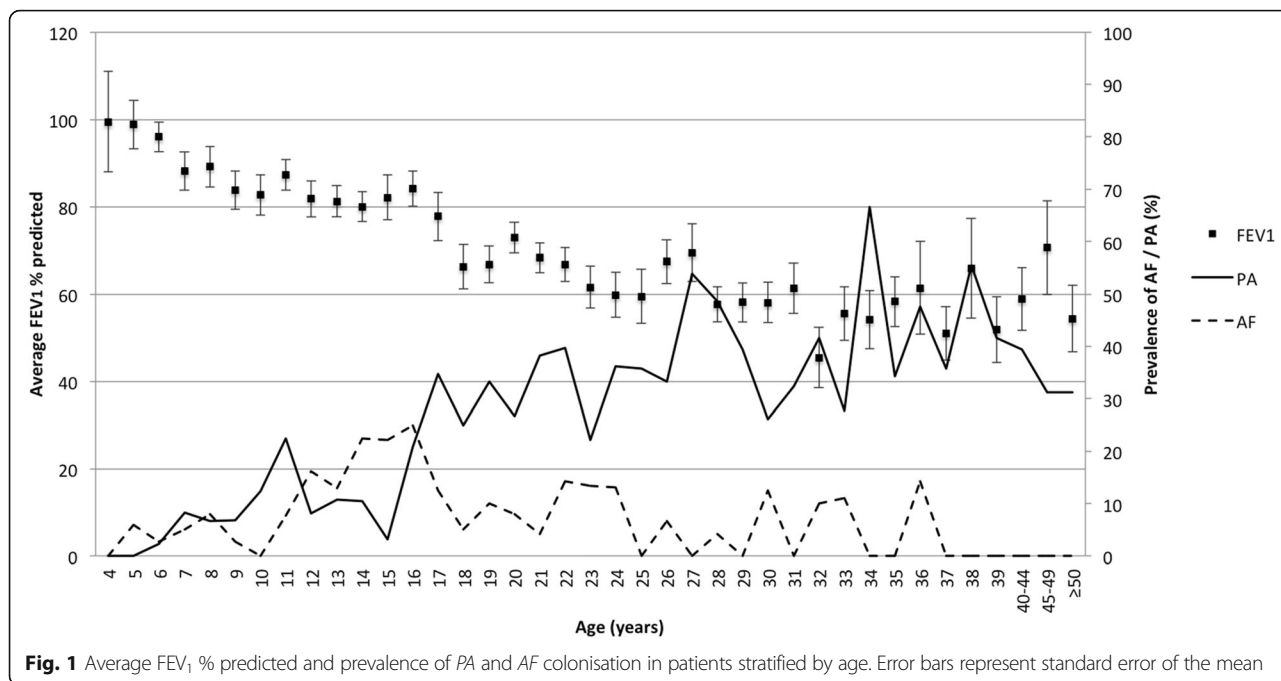
### Persistent colonisation with PA or co-colonisation with AF was associated with reduced lung function

Patients co-colonised with both AF and PA had an FEV<sub>1</sub> 13.8% lower than non-colonised patients ( $p = 0.016$ )

**Table 1** Demographic and clinical data of the study cohorts

	PA	PAp	AF	AFp	AF + PA	Clear
Number (%)	82 (10.9%)	130 (17.4%)	36 (4.8%)	26 (3.5%)	23 (3.1%)	452 (60.3%)
Median Age (range)	26 (7–69)	25 (6–48)	15 (6–36)	15 (5–29)	20 (9–45)	14 (4–56)
CFTR Genotype						
phe508del/phe508del	6.8%	10.3%	2.8%	2.9%	1.7%	30.8%
phe508del/other	3.8%	6%	1.6%	0.4%	1.1%	24.3%
other	0.5%	0.8%	0.27%	0.13%	0.27%	4%
unknown	0.13%	0.27%	0.13%	0%	0%	1.2%
Median BMI (range)	21.2 <sup>a</sup> (13.1–34.9)	20 (9–30.2)	19 (14.3–26.8)	18.7 (14.3–26.1)	19 (12.3–25.7)	18.9 (10.4–33.7)

<sup>a</sup>indicates missing data

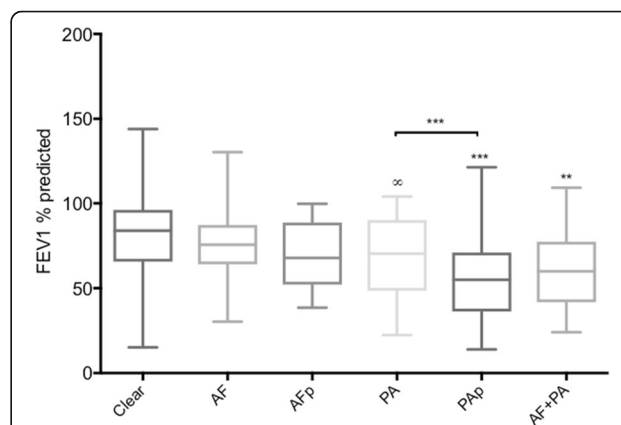


**Fig. 1** Average FEV<sub>1</sub> % predicted and prevalence of PA and AF colonisation in patients stratified by age. Error bars represent standard error of the mean

(Fig. 2) and co-colonised patients had comparable reductions in lung function to patients persistently colonised with PA. Consistent with our data, Amin et al., (2010) [9] reported that persistent infection with both AF and PA was associated with lower lung function. The transition of PA colonisation from intermittent to persistent is a disease milestone that cannot be reversed and is associated with worse prognosis [18, 27]. We have shown that patients co-colonised with PA and AF had similar reductions in lung function as patients with persistent PA. This is the first time this has been reported. In a recent longitudinal study the sputum microbiology and clinical outcomes of 770 adolescents with CF were recorded and AF was the only species that was associated with an increased risk for infection with PA [28]. This highlights the importance of monitoring the co-colonisation status of CF patients. Perhaps AF colonisation establishes a milieu that enables PA to become persistent and attention to AF during this phase may prevent progression to persistent PA. Further data is required to explore this hypothesis.

Patients with persistent PA colonisation had an FEV<sub>1</sub> 11.9% lower than patients who were intermittently colonised with PA (unadjusted  $p = 0.0074$ ), which remained significant following adjustment for confounding factors ( $p < 0.001$ ) (Fig. 2). Previously intermittent PA colonisation has been associated with lower FEV<sub>1</sub> [21] (when adjusted for age and gender only). Following data adjustment for confounding factors (age, gender and CFTR mutation) the FEV<sub>1</sub> of patients intermittently colonised with PA were not significantly different from the FEV<sub>1</sub> of non-colonised patients. Persistent PA colonisation negatively impacted

on lung function in agreement with others [21, 23]. Early onset PA colonisation can be eradicated in over 90% of colonised patients by antibiotic therapy [29], therefore efforts should be targeted towards early detection of PA colonisation and preventing establishment of persistent colonisation. Interestingly 2 studies have now shown that polymerase chain reaction methods can detect PA 4.5 months [30] to 8 months [31] in advance of current culture-based techniques used commonly in diagnostic



**Fig. 2** Average FEV<sub>1</sub> percent predicted in the six colonisation cohorts. Average FEV<sub>1</sub> % predicted for patients with no PA or AF colonisation (Clear), intermittent AF colonisation (AF), persistent AF colonisation (AFP), intermittent PA colonisation (PA), persistent PA colonisation (PAP) and both AF and PA co-colonisation (AF + PA). Error bars represent min and max values. Asterisks refer to comparison with Clear, unless over a horizontal bar, \*\*  $p \leq 0.01$ , \*\*\*  $p \leq 0.001$  (adjusted data). ∞ indicates significance only prior to data adjustment

laboratories. This provides evidence to support the use of molecular-based methods for superior detection of PA.

Patients with intermittent and persistent *AF* colonisation had 4 and 11% lower FEV<sub>1</sub>, respectively, than non-colonised patients however these were not statistically significant. A previous study showed patients with *AF* sensitisation or persistent carriage had 16.5% lower FEV<sub>1</sub> than the control group [32]. Amin et al., (2010) [9] showed that patients persistently colonised with *AF* had a 3.6% decrease in FEV<sub>1</sub> compared to patients who were clear of *AF* however when the data was adjusted for baseline pulmonary function, this decrease was not significant. In agreement with our data, other studies suggest no correlation between *AF* colonisation and a decline in pulmonary function [33, 34].

#### Persistent colonisation with PA or co-colonisation with both PA and AF was associated with more frequent hospitalisations, more respiratory exacerbations and higher use of antimicrobials

For the first time we have shown that being culture positive for both *AF* and *PA* is associated with similar levels of hospital admissions, exacerbations and antimicrobial prescribing as being persistently colonised with *PA* (Table 2). Patients co-colonised with both *AF* and *PA* had a 165% increase in hospital admissions per person ( $p = 0.023$ ), a 112% increase in the number of respiratory exacerbations per person ( $p = 0.042$ ) and 48% increase in antimicrobial prescribing ( $p = 0.009$ ) when compared to patients who were clear of both pathogens (Table 2). Co-colonised patients had a 64% increase in respiratory exacerbations compared to patients intermittently colonised with *PA* ( $p = 0.015$ ) (Table 2).

Patients persistently colonised with *PA* had an 89% increase in the number of hospital admissions per person

within the year ( $p = 0.004$ ), a 91% increase in respiratory exacerbations ( $p < 0.001$ ) and a 55% increase in antimicrobial prescribing ( $p < 0.001$ ) compared to non-colonised patients (Table 2). In comparison to intermittently colonised patients, those persistently colonised with *PA* had a 60% increase in the number of exacerbations ( $p = 0.001$ ) and 23% increase in antibiotic prescribing ( $p = 0.014$ ) (Table 2). There was no increase in the number of hospitalisations, respiratory exacerbations or antimicrobial prescribing in patients who were intermittently colonised with *PA* compared to non-colonised patients.

Patients intermittently or persistently colonised with *AF* showed no increase in the number of hospitalisations or respiratory exacerbations per person compared to non-colonised patients. However intermittently and persistently colonised patients had a 63% and 90% increase in antimicrobial prescribing compared to non-colonised patients ( $p = <0.001$  and  $p = 0.001$  respectively) (Table 2). The prevalence of *Aspergillus* sp was previously shown to be higher in CF patients receiving prophylactic antibiotic therapy [33] and we also observed an association between *AF* colonisation and antimicrobial prescribing. We speculate that this finding may reflect antimicrobial suppression of bacteria facilitating fungal growth. It is also possible that *AF* colonisation establishes a milieu suitable for bacterial colonisation, which would impact on antimicrobial prescribing. No differences were observed between the cohorts regarding inhaled steroid use, steroids taken daily and steroids taken alternative days (Additional file 3: Figure S1).

#### Patients colonised with PA had a higher prevalence of CFRD

A total of 14.7% of patients with CF had diabetes requiring insulin in 2013. Patients persistently colonised with

**Table 2** Incident Rate Ratio (IRR) of the incidence of hospitalisations, respiratory exacerbations and prescribed antimicrobials between cohorts

Comparison	Hospitalisations		Respiratory Exacerbations		Antimicrobials	
	IRR (95% CI)	<i>p</i> value	IRR (95% CI)	<i>p</i> value	IRR (95% CI)	<i>p</i> -value
PA vs Clear	1.00 (0.44, 2.30)	1	0.76 (0.37, 1.55)	1	1.19 (0.93, 1.51)	0.259
PAp vs Clear	1.89 (1.11, 3.22)	<b>0.004</b>	1.91 (1.19, 3.05)	<b>&lt;0.001</b>	1.55 (1.29, 1.86)	<b>&lt;0.001</b>
AF vs Clear	1.16 (0.54, 2.48)	1	0.95 (0.45, 1.98)	1	1.63 (1.19, 2.21)	<b>&lt;0.001</b>
AFp vs Clear	1.97 (0.75, 5.20)	0.271	1.59 (0.74, 3.44)	0.494	1.9 (1.38, 2.62)	<b>&lt;0.001</b>
AF + PA vs Clear	2.65 (1.01, 6.97)	<b>0.023</b>	2.12 (0.95, 4.72)	<b>0.042</b>	1.48 (1.04, 2.11)	<b>0.009</b>
PA vs PAp	0.53 (0.23, 1.23)	0.189	0.4 (0.19, 0.81)	<b>0.001</b>	0.77 (0.60, 0.98)	<b>0.014</b>
AF vs Afp	0.59 (0.21, 1.67)	0.872	0.59 (0.24, 1.47)	0.775	0.86 (0.56, 1.30)	1
AF + PA vs PA	0.38 (0.12, 1.21)	0.099	0.36 (0.13, 0.95)	<b>0.015</b>	0.8 (0.54, 1.19)	0.668
AF + PA vs PAp	1.40 (0.53, 3.73)	1	1.11 (0.51, 2.43)	1	0.96 (0.66, 1.38)	1
AF + PA vs AF	0.45 (0.15, 1.33)	0.209	0.47 (0.17, 1.31)	0.205	1.09 (0.69, 1.74)	1
AF + PA vs AFp	1.30 (0.38, 4.44)	1	0.79 (0.28, 2.21)	1	0.78 (0.49, 1.25)	0.8

Italicized bold text highlights significance of  $<0.05$ .

*PA* had a higher prevalence of CFRD diagnosis than non-colonised patients ( $p = 0.012$ ) (Additional file 4: Table S2). A direct link between insulin deficiency and clinical decline has been shown and patients with CFRD were more likely to be persistently colonised with *Pseudomonas sp.* [35].

#### AF colonisation was not associated with prevalence of ABPA

The overall prevalence of ABPA was 5.9%. Patients co-colonised with *AF* and *PA* had the highest prevalence of ABPA (17.4%) but this was not significantly higher than the other groups (Additional file 4: Table S2). Interestingly, patients intermittently or persistently colonised with *AF* showed no increased prevalence of ABPA. It is not currently understood why some *AF* colonised patients are vulnerable to developing ABPA and others are not. A number of single nucleotide polymorphisms (SNPs) have been linked to susceptibility to ABPA. SNPs in the IL-4 receptor alpha chain (IL-4R $\alpha$ ) gene [36], the mannose-binding lectin (MBL) 2 gene [37, 38], the toll-like receptor 9 (TLR9) gene [39] and the pulmonary surfactant protein (SPA-2) gene [40] have been identified as genetic risk factors for the development of ABPA. In particular, SNPs in the promoter region of the IL-10 gene were found to be associated with *Aspergillus* colonisation and ABPA in patients with CF. Further studies in the CF setting could yield interesting insights into the susceptibility of CF patients to *Aspergillus* colonisation and development of allergic disease. Culture of *AF* from respiratory secretions of CF is only a secondary criterion for the diagnosis of ABPA, due to the large number of asymptotically *AF* colonised CF patients. A retrospective study on 236 patients with CF found that 25% of patients had positive *AF* culture results but of these only 15 patients (6.5% of the total population) were ABPA positive [41]. In parallel with our findings, positive culture results for *Aspergillus* did not seem to represent a specific risk factor for ABPA [40, 42].

The CF airways are colonised with a diverse community of microorganisms from an early age [2–4]. Interactions between the varieties of dominant and low abundance microorganisms in these communities are likely to impact patient health. The most commonly isolated bacteria and fungi from CF samples are *PA* and *AF*, respectively and we have shown that co-colonisation with these two microorganisms is associated with poor clinical outcomes. Previous studies have shown that *PA* and *AF* are capable of interacting and competing, which may have implications on CF airway disease. Pyocyanin and 1-hydroxy-phenazine produced by *PA* both have anti-*AF* activity [43] and *PA* has been shown to inhibit *AF* biofilm formation by direct cell contact and by excreted molecules [44, 45]. More recently the Pf4 bacteriophage produced by *PA* was shown

to reduce both biofilm formation and pre-formed biofilms of *AF* and this effect was linked to inhibition of *AF* metabolism via Pf4 iron sequestration [46]. *AF* has been shown to enhance the production of *PA* elastase when in co-culture and these co-culture supernatants were more damaging to lung epithelial cell lines than *PA* supernatants alone [47]. We hypothesize that these *PA*-*AF* interactions may play a role in the damaging pathology associated with CF airway disease and may explain why patients who are co-colonised with *PA* and *AF* have a poorer prognosis than those who are clear of both pathogens. Further studies on the interaction of these two microorganisms are warranted to expand our understanding of how co-colonisation may impact the CF airways.

#### Conclusions

Persistent *PA* colonisation was associated with more respiratory exacerbations, more hospitalisations and lower lung function than non-colonised patients. For the first time we have shown that co-colonisation with *AF* and *PA* resulted in comparable levels of hospitalisations, respiratory exacerbations and lung function reduction as persistent *PA* colonisation. Patients with persistent *PA* had a higher prevalence of CFRD diagnosis. Intermittent and persistent *AF* colonisation was not associated with poorer clinical outcomes or ABPA. This is a cross-sectional registry analysis therefore a prospective longitudinal study is warranted to confirm these findings.

#### Additional files

**Additional file 1:** Supplementary Methods. Further details on the statistics performed on this data (DOCX 140 kb)

**Additional file 2: Table S1.** Comparison of disease outcome measures between the co-colonised sub-groups (PNG 74 kb)

**Additional file 3: Figure S1.** Percentage of patients prescribed steroids in each cohort. Percentage of patients prescribed inhaled steroids, oral steroids every day and oral steroids every alternative day for patients within the 6 cohorts (PNG 109 kb)

**Additional file 4: Table S2.** Odds ratios are presented to determine the odds of a particular colonisation status being linked to development of CFRD and/ or ABPA. (PNG 75 kb)

#### Abbreviations

ABPA: Allergic Bronchopulmonary Aspergillosis; *AF*: *Aspergillus fumigatus*; *AF + PA*: *AF* and *PA* co-colonised; *AFp*: Persistent *AF*; *CF*: Cystic Fibrosis; CFRD: CF related diabetes; CFRI: CF registry of Ireland; CFTR: CF transmembrane conductance regulator; FEV<sub>1</sub>: Forced Expiratory Volume in 1 second; IRR: Incident Rate Ratios; OR: Odds Ratio; *PA*: *Pseudomonas aeruginosa*; *PAp*: Persistent *PA*; qPCR: quantitative Polymerase Chain Reaction; SNP: Single nucleotide polymorphism; ZINB: Zero-inflated Negative Binomial

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### Availability of data and materials

All the relevant data is presented within the main manuscript and supplementary files. The CFRI dataset is not a publicly available dataset. Requests for anonymous data/information from the CFRI are considered, and are subjected to a data application process. Data/information requests are considered by CFRI's Research Committee whose decision whether to provide data or information is final.

### Authors' contributions

ER was involved in study design, data analysis, data interpretation and writing and editing the manuscript. RS performed the data analysis and edited the manuscript. AJ extracted data from the CFRI, helped with editing the manuscript and performed sub-analysis in response to reviewer's comments. SMcC was involved in data interpretation and manuscript editing. JR was involved in study design, data analysis, data interpretation and writing and editing the manuscript. PG was involved in study design, data interpretation and manuscript editing. All authors read and approved the final manuscript.

### Competing interests

PG has served on advisory boards and received honoraria from Vertex, Novartis, Pharmaxis and PTC pharmaceuticals in the past however none of these organisations have contributed financially or in any other way to this work. For all other co-authors there are no conflicts of interest to report.

### Consent for publication

Not applicable.

### Ethics approval and consent to participate

Cystic fibrosis registry of Ireland (CFRI) data were used in this study, and the registry was given ethical approval by each Irish cystic fibrosis centre and clinic to collect data on cystic fibrosis patients. Informed consent is provided by registry participants and/or their parent/guardian at registration. The individual ethics committees are as follows; University hospital Galway, University hospital Limerick, Cork University hospital, Beaumont hospital, St Vincents hospital, Tallaght hospital, Our Lady's Children's hospital, Crumlin, Temple Street hospital, Our Lady of Lourdes hospital, Drogheda, Cavan & Monaghan hospital, Sligo University hospital, Mayo General hospital and Waterford Regional hospital.

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### References

- Renwick J, et al. The microbial community of the cystic fibrosis airway is disrupted in early life. *PLoS ONE*. 2014;9(12), e109798.
- Zhao J, et al. Decade-long bacterial community dynamics in cystic fibrosis airways. *Proc Natl Acad Sci U S A*. 2012;109(15):5809–14.
- Willger SD, et al. Characterization and quantification of the fungal microbiome in serial samples from individuals with cystic fibrosis. *Microbiome*. 2014;2:40.
- Spicuzza L, et al. Emerging pathogens in cystic fibrosis: ten years of follow-up in a cohort of patients. *Eur J Clin Microbiol Infect Dis*. 2009;28(2):191–5.
- Steinkamp G, et al. Prospective evaluation of emerging bacteria in cystic fibrosis. *J Cyst Fibros*. 2005;4(1):41–8.
- Becker JW, et al. Prevalence of allergic bronchopulmonary aspergillosis and atopy in adult patients with cystic fibrosis. *Chest*. 1996;109(6):1536–40.
- Skov M, et al. Prevalence of allergic bronchopulmonary aspergillosis in cystic fibrosis in an area with a high frequency of atopy. *Respir Med*. 2005;99(7):887–93.
- Valenza G, et al. Prevalence and antimicrobial susceptibility of microorganisms isolated from sputa of patients with cystic fibrosis. *J Cyst Fibros*. 2008;7(2):123–7.
- Amin R, et al. The effect of chronic infection with *Aspergillus fumigatus* on lung function and hospitalization in patients with cystic fibrosis. *Chest*. 2010;137(1):171–6.
- Stevens DA, et al. Allergic bronchopulmonary aspergillosis in cystic fibrosis—state of the art: Cystic Fibrosis Foundation Consensus Conference. *Clin Infect Dis*. 2003;37 Suppl 3:S225–64.
- Baxter CG, et al. Novel immunologic classification of aspergillosis in adult cystic fibrosis. *J Allergy Clin Immunol*. 2013;132(3):560–6. e10.
- Coughlan CA, et al. The effect of *Aspergillus fumigatus* infection on vitamin D receptor expression in cystic fibrosis. *Am J Respir Crit Care Med*. 2012;186(10):999–1007.
- Mirkovic B, et al. The basophil surface marker CD203c identifies *Aspergillus* species sensitization in patients with cystic fibrosis. *J Allergy Clin Immunol*. 2016;137(2):436–43. e9.
- Hoiby N. Recent advances in the treatment of *Pseudomonas aeruginosa* infections in cystic fibrosis. *BMC Med*. 2011;9:32.
- Marshal B, et al. Cystic Fibrosis Foundation Patient Registry 2013 Annual Data Report to the Center Directors Bethesda, Maryland. *Cyst. Fibros. Found.*, 2014. p. 1–92.
- Hogardt M, Heesemann J. Adaptation of *Pseudomonas aeruginosa* during persistence in the cystic fibrosis lung. *Int J Med Microbiol*. 2010;300(8):557–62.
- Schelstraete P, et al. Eradication therapy for *Pseudomonas aeruginosa* colonization episodes in cystic fibrosis patients not chronically colonized by *P. aeruginosa*. *J Cyst Fibros*. 2013;12(1):1–8.
- Emerson J, et al. *Pseudomonas aeruginosa* and other predictors of mortality and morbidity in young children with cystic fibrosis. *Pediatr Pulmonol*. 2002;34(2):91–100.
- Nixon GM, et al. Clinical outcome after early *Pseudomonas aeruginosa* infection in cystic fibrosis. *J Pediatr*. 2001;138(5):699–704.
- CFRI. The Cystic Fibrosis Registry of Ireland 2013 Annual Report. Ireland: Cyst. Fibros. Regist. Ire; 2013. p. 1–158.
- Lee TW, et al. Evaluation of a new definition for chronic *Pseudomonas aeruginosa* infection in cystic fibrosis patients. *J Cyst Fibros*. 2003;2(1):29–34.
- Rosenfeld M, et al. Gender gap in cystic fibrosis mortality. *Am J Epidemiol*. 1997;145(9):794–803.
- Kerem E, et al. Factors associated with FEV1 decline in cystic fibrosis: analysis of the ECFS patient registry. *Eur Respir J*. 2014;43(1):125–33.
- Proesmans M, et al. Evaluating the "Leeds criteria" for *Pseudomonas aeruginosa* infection in a cystic fibrosis centre. *Eur Respir J*. 2006;27(5):937–43.
- ECFS. European Cystic Fibrosis Society Patient Registry. 2014.
- Bakare N, et al. Prevalence of *Aspergillus fumigatus* and other fungal species in the sputum of adult patients with cystic fibrosis. *Mycoses*. 2003;46(1–2):19–23.
- Kosorok MR, et al. Acceleration of lung disease in children with cystic fibrosis after *Pseudomonas aeruginosa* acquisition. *Pediatr Pulmonol*. 2001;32(4):277–87.
- Hector A, et al. Microbial colonization and lung function in adolescents with cystic fibrosis. *J Cyst Fibros*. 2016;15(3):340–9.
- Ratjen F, et al. Treatment of early *Pseudomonas aeruginosa* infection in patients with cystic fibrosis: the ELITE trial. *Thorax*. 2010;65(4):286–91.
- Xu J, et al. Early detection of *Pseudomonas aeruginosa*—comparison of conventional versus molecular (PCR) detection directly from adult patients with cystic fibrosis (CF). *Ann Clin Microbiol Antimicrob*. 2004;3:21.
- Hery-Arnaud G, et al. Evaluation of quantitative PCR for early diagnosis of *Pseudomonas aeruginosa* infection in cystic fibrosis: a prospective cohort study. *Clin Microbiol Infect*. 2017;23(3):203–7.
- Fillaux J, et al. Assessment of *Aspergillus* sensitization or persistent carriage as a factor in lung function impairment in cystic fibrosis patients. *Scand J Infect Dis*. 2012;44(11):842–7.

33. Bargon J, et al. Prophylactic antibiotic therapy is associated with an increased prevalence of *Aspergillus* colonization in adult cystic fibrosis patients. *Respir Med*. 1999;93(11):835–8.
34. De Vrankrijker AM, et al. *Aspergillus fumigatus* colonization in cystic fibrosis: implications for lung function? *Clin Microbiol Infect*. 2011;17(9):1381–6.
35. Milla CE, Warwick WJ, Moran A. Trends in pulmonary function in patients with cystic fibrosis correlate with the degree of glucose intolerance at baseline. *Am J Respir Crit Care Med*. 2000;162(3 Pt 1):891–5.
36. Knutsen AP, et al. IL-4 alpha chain receptor (IL-4Ralpha) polymorphisms in allergic bronchopulmonary aspergillosis. *Clin Mol Allergy*. 2006;4:3.
37. Vaid M, et al. Distinct alleles of mannose-binding lectin (MBL) and surfactant proteins A (SP-A) in patients with chronic cavitary pulmonary aspergillosis and allergic bronchopulmonary aspergillosis. *Clin Chem Lab Med*. 2007;45(2):183–6.
38. Kaur S, et al. Elevated levels of mannan-binding lectin [corrected] (MBL) and eosinophilia in patients of bronchial asthma with allergic rhinitis and allergic bronchopulmonary aspergillosis associate with a novel intronic polymorphism in MBL. *Clin Exp Immunol*. 2006;143(3):414–9.
39. Carvalho A, et al. Polymorphisms in toll-like receptor genes and susceptibility to pulmonary aspergillosis. *J Infect Dis*. 2008;197(4):618–21.
40. Saxena S, et al. Association of polymorphisms in the collagen region of SP-A2 with increased levels of total IgE antibodies and eosinophilia in patients with allergic bronchopulmonary aspergillosis. *J Allergy Clin Immunol*. 2003;111(5):1001–7.
41. Mroueh S, Spock A. Allergic bronchopulmonary aspergillosis in patients with cystic fibrosis. *Chest*. 1994;105(1):32–6.
42. Milla CE, Wielinski CL, Regelmann WE. Clinical significance of the recovery of *Aspergillus* species from the respiratory secretions of cystic fibrosis patients. *Pediatr Pulmonol*. 1996;21(1):6–10.
43. Kerr JR, et al. *Pseudomonas aeruginosa* pyocyanin and 1-hydroxyphenazine inhibit fungal growth. *J Clin Pathol*. 1999;52(5):385–7.
44. Mowat E, et al. *Pseudomonas aeruginosa* and their small diffusible extracellular molecules inhibit *Aspergillus fumigatus* biofilm formation. *FEMS Microbiol Lett*. 2010;313(2):96–102.
45. Shirazi F, et al. Biofilm Filtrates of *Pseudomonas aeruginosa* Strains Isolated from Cystic Fibrosis Patients Inhibit Preformed *Aspergillus fumigatus* Biofilms via Apoptosis. *PLoS ONE*. 2016;11(3), e0150155.
46. Penner JC, et al. Pf4 bacteriophage produced by *Pseudomonas aeruginosa* inhibits *Aspergillus fumigatus* metabolism via iron sequestration. *Microbiology*. 2016;162(9):1583–94.
47. Smith K, et al. *Aspergillus fumigatus* enhances elastase production in *Pseudomonas aeruginosa* co-cultures. *Med Mycol*. 2015;53(7):645–55.

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