$\frac{1}{2}$ Biorheology 00 (2010) 1–16 DOI 10.3233/BIR-2010-0568

$\frac{3}{2}$ and $\frac{3}{2}$ and $\frac{3}{2}$ and $\frac{3}{2}$ and $\frac{3}{2}$ and $\frac{3}{2}$ $\frac{1}{4}$ A full-range, multi-variable, CFD-based 5 6 methodology to identity abnormal near-wall 6 $\frac{1}{2}$ methodology to identify abnormal near-wall $\frac{1}{2}$ 8 hours a drug assign is a stantage concrete $\frac{1}{2}$ $\frac{1}{9}$ hemodynamics in a stented coronary artery $\frac{1}{9}$

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11 **11 11 11 11** 12 Jonathan B. Murphy [∗] and Fergal J. Boyle 12

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 14 $$ $$ Received 30 December 2009

15 15 Accepted in revised form 11 June 2010

 16 16

17 **Abstract.** The benefit of coronary stent implantation is reduced by excessive intimal hyperplasia which re-narrows the artery ¹⁸ and the prevention of which is still a primary concern for clinicians. Abnormal hemodynamics create non-physiological vis-¹⁸ 19 cous stress on the artery wall, one of the root causes of intimal hyperplasia following stent implantation. A methodology to 19 ₂₀ comprehensively evaluate the viscous stress on the artery wall following stent implantation would be useful to evaluate a stent's ₂₀ hemodynamic performance.

²¹ The proposed methodology employs 3D computational fluid dynamics, the variables wall shear stress (WSS), WSS gra-22 dient (WSSG), WSS angle gradient (WSSAG) and a statistical analysis to evaluate the viscous stress. The methodology is 22 ₂₃ demonstrated and compared to a commonly used "threshold technique" for evaluating a stent's hemodynamic performance. ₂₃

24
even be misleading. Furthermore, all three of the aforementioned variables should be considered as each provides a different ²⁵ perspective on the abnormalities that can arise in the arterial viscous stress. ²⁵ It is demonstrated that the threshold technique is not adequate to fully analyse the viscous stress on the artery wall and can

26 26 The hemodynamic performance of a stent can be assessed more comprehensively than with previously used methods by examining the arterial viscous stresses using the proposed methodology. 27

- 28 28 Keywords: Stent, intimal hyperplasia, viscous stress, statistical analysis
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- 30
- 31

32 32 **1. Introduction**

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 CORRECT AND THE CONDUCTS AND A SUPREME EXAMPLE TO THE CONDUCTS AND THE CONDUCTS CONDUCTS AND THE CONDUCTS CONDUCTS CONDUCTS CONDUCTS CONDUCTS CONDU 33 33 34 Coronary artery disease (CAD) is one of the leading causes of mortality in the developed world. CAD 34 35 occurs due to the build up of plaque in the coronary arteries which supply blood to the heart muscle. 35 36 This build up of plaque, a condition known as atherosclerosis, deprives the heart muscle of crucial 36 $_{37}$ oxygen and nutrients provided by the coronary blood supply. In the early 1990s bare metal stents (BMSs) $_{37}$ 38 were introduced to restore patency to the diseased coronary artery. Basically, a BMS is a metal scaffold 38 39 inserted into the artery and then, most commonly, expanded by a balloon to relieve the narrowing caused ₃₉ 40 by atherosclerosis. Unfortunately, the stented artery is susceptible to restenosis, defined as a greater than 40 $_{41}$ 50% re-blockage of the artery [15]. The prevalence of restenosis with BMSs varies between 10% and $_{41}$ 42 50% depending on the type of stent implanted [18,9,19,28,35]. Restenosis is caused by the excessive 42 43 growth of new tissue in the stented segment of the artery, a process termed intimal hyperplasia (IH). 43

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Giantures Revoluti II (GR-II) sterit (Cook Inc., Bloomington, IN, USA) and the Cordis Be-Velopsiting

is y stert (Johnson and Johnson Interventional System, Warren, NJ, USA). These contrasting signs are chosen to investig ¹ components of the WSS vector are then post-processed to obtain the WSS, WSSG and WSSAG on ¹ ² these surfaces. Histograms of the amount of area contained between specific intervals of the variable ² 3 values are produced. A statistical analysis is then conducted on the full range of these variables to get ³ 4 4 the maximum information possible from the predicted results. ⁵ This methodology is applied to evaluate the abnormal near-wall haemodynamics resulting from im-6 6 plantation of three different stents in the left anterior descending (LAD) coronary artery. The stents re-7 7 semble the Palmaz–Schatz (PS) stent (Johnson and Johnson Interventional System, Warren, NJ, USA), 8 the Gianturco–Roubin II (GR-II) stent (Cook Inc., Bloomington, IN, USA) and the Cordis Bx-Velocity 8 9 (Bx) stent (Johnson and Johnson Interventional System, Warren, NJ, USA). These contrasting stent 9 10 designs are chosen to investigate if the notable differences in their geometries produce significantly dif-
¹⁰ 11 ferent haemodynamic disturbances in the artery. Also, the prior knowledge of the *in vivo* performance 11 ¹² of these stents is useful when analysing the results as abnormal near-wall haemodynamics may have ¹² 13 contributed to their restenosis rates. The PS stent consists of two 7 mm long slotted-tube sections joined 13 ¹⁴ by a central articulation and is commonly used in computational modelling due to its generic design. In ¹⁴ 15 15 one clinical trial, the PS stent had a restenosis rate of 20.6% [28]. The GR-II stent on the other hand ¹⁶ performed poorly in the same trial with a restenosis rate of 47.3% [28]. The GR-II stent has a coil design ¹⁶ 17 consisting of widely-spaced interdigitating loops. The Bx stent is of a closed cell design with thick struts 17 18 connected by S-shaped connectors. Implantation of the Bx stent resulted in a restenosis rate of 31.4% in 18 19 19 the ISAR-STERO II trial [35]. 20 20

21 21 *2.2. Computational fluid dynamics*

23 23 *2.2.1. Computational domain*

24 24 The 3D solid model is created beginning with a solid cylinder measuring the length and external 25 diameter of the stent; this is the region of interest with details provided in Table 1. The geometry of the 25 26 stent is then removed from the cylinder. The solid model is extended proximal and distal to the stented 26 ₂₇ section by adding cylinders 21.3 mm long, which equals to the entrance length for fully-developed ₂₇ 28 laminar flow. These cylinders have diameters of 3.2 mm which creates a stent-to-artery deployment ratio 28 29 29 of 1.09:1 similar to a normal *in vivo* value [28]. These extra lengths ensure the region of interest is not 30 affected by the inlet and outlet boundary flow conditions. A tapered section of 2 mm in length connects 30 31 31 the stented and unstented sections similar to a previous numerical study [24].

22 22

32 A sensitivity analysis was performed to investigate the effect of taper length on the WSS in the region 32 33 of interest. The mean WSS in a 14.3 mm long section of a 3.5 mm diameter artery was 0.797, 0.804 and 33 34 0.809 N/m when the taper lengths were 1.5, 2.0 and 2.5 mm, respectively, and the smaller diameter was 34

 $T₁$ 1 1 \pm

35 35

 $\frac{1}{1}$ 2 Primary Scaffolding 2 3 Direction 3 $4 \qquad \qquad 4$ 5 5 5 5 $\sqrt{}$ 5 5 5 5 6 7 7 $\sqrt{}$ 5 6 6 7 7 $\sqrt{2}$ 7 $\sqrt{2$ 8 and λ and 9 and \mathbb{P} and \mathbb{P} 10 \sim 10 \sim 10 11 $\sqrt{2}$ $\sqrt{2}$ 11 12 Scaffolding Scaffolding 12 13 and the contract of the con

14
Fig. 1. Illustration of the primary and secondary scaffolding direction of part of the PS stent. The arterial tissue protrudes a 15 depth δ into the stented artery and is supported in the primary scaffolding direction and partially supported (shaded section) in 15 16 the secondary scaffolding direction. 16

18 3.2 mm. Therefore, either increasing or decreasing the taper length by 0.5 mm, resulted in less than a 18 19 19 1% change in mean WSS in the region of interest.

₂₀ In the geometric model, the assumption is made that the stenotic plaque has been completely com-₂₁ pressed against the wall. Then, a novel methodology to numerically predict tissue prolapse between stent ₂₁ 22 struts [32] is employed. Briefly, the prolapsing tissue creates a variable artery radius r given by

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$$
\frac{r}{R_o} = 1 - \frac{x\delta}{2R_o} \left\{ 1 + \cos\frac{2\pi}{L} \left(z - \frac{L}{2} \right) \right\},\tag{1}
$$

²⁶ where R_0 is the external diameter of the stent, L is the distance between the stent struts in the primary $\frac{26}{27}$ 27

scaffolding direction and δ is the prolapse depth equal to the product of L and a constant C obtained 28 from finite element analysis data [36]. The variable x is a prolapse reduction factor initiated at a distance ²⁹ of 0.5L from struts offering secondary scaffolding support as shown in Fig. 1.

 $\frac{30}{21}$ This prolapsing tissue is then removed from the solid model. The solid models of the stented region $\frac{30}{21}$ 31 31 of interest and the tapered section are shown in Fig. 2. The geometric characteristics given in Table 1 $\frac{32}{32}$ $\frac{33}{33}$ detail some of the major scaffolding features of each stent model.

34 34 *2.2.2. Computational mesh*

Prolapse

Frochaye

Frochaye

Secondary

Seco 35 The computational domains are discretised using an unstructured mesh topology. This is achieved in 36 ANSYS Workbench 12.0 using the advancing front method. A mesh convergence study is conducted for 37 all stents to ensure the results are independent of the computational mesh density. Results are considered 38 mesh converged when the difference in the RMS values of the WSS and WSSG between successive mesh 39 densities is less than 4% along a sample line similar to a strategy employed in a previous numerical study 40 [23]. Mesh convergence is achieved with 4,551,484 elements for the PS stent, 3,038,536 elements for 41 the GR-II stent and 5,840,890 elements for the Bx stent. Results from the mesh convergence study are 42 shown in Fig. 3 for the GR-II stent.

43 43 *2.2.3. Governing flow equations*

⁴⁴ The flow in the stented artery is assumed steady, laminar and incompressible. In this work, a transient ⁴⁴ ⁴⁵ analysis is not deemed necessary to demonstrate the advantage of the current methodology. To predict ⁴⁵ 46 46

17 17

³¹ Fig. 3. Magnitude of WSS and WSSG along a sample axial line which crosses two struts of the GR-II stent for three mesh ³¹ densities.

³³ the flow the CFD code solves the general form of the conservation of mass and momentum equations³³ which are given in vector form in Eqs (2) and (3), respectively 34 35

$$
\frac{\partial \rho}{\partial t} + \vec{\nabla} \cdot (\rho \vec{V}) = 0, \tag{2}
$$

$$
\frac{\partial \rho \vec{V}}{\partial t} + \vec{\nabla} \cdot (\rho \vec{V} \otimes \vec{V}) = -\vec{\nabla} p + \vec{\nabla} \cdot (\vec{\vec{\tau}}_{ij}),\tag{3}
$$

41

44

42 where ρ is the fluid density, p is the static pressure, \vec{V} is the velocity vector, μ is the fluid dynamic 42 43 viscosity and $\vec{\tau}_{ij}$ is the shear stress tensor written 43

$$
\vec{\tau}_{ij} = \mu (\vec{\nabla} \vec{V} + \vec{\nabla} \vec{V}^{\mathrm{T}}). \tag{4}
$$

(6)

6 *J.B. Murphy and F.J. Boyle / CFD-based methodology to assess coronary stents*

1 At low shear rates $(0-100 \text{ s}^{-1})$ blood exhibits the non-Newtonian behaviour of variable fluid dynamic ¹ 2 2 viscosity which is dependant on the shear rate. This non-Newtonian nature of the flow is accommodated 3 3 using the Carreau model [17] written

4 4

$$
\mu = \mu_{\infty} + (\mu_0 - \mu_{\infty})[1 + (\gamma \lambda)^2]^{((q-1)/2)},\tag{5}
$$

 6 where γ is the shear rate calculated as the second invariant of the strain-rate tensor and the constants are $\frac{1}{7}$

8 a set of the contract of the $\mu_0 = 0.056 \text{ N} \cdot \text{s} \cdot \text{m}^{-2}, \qquad \lambda = 3.31 \text{ s},$ 10 10 $\lambda = 3.31$ s,

$$
\mu_{\infty} = 0.00345 \text{ N} \cdot \text{s} \cdot \text{m}^{-2}, \qquad q = 0.375.
$$

¹² The commercial software code ANSYS CFX Version 12 (Canonsburg, PA, USA) is used to solve the ¹² ¹³ governing equations in the computational domain using a vertex-centred finite volume scheme which is ¹³ ¹⁴ second order accurate in space.¹⁴

15 $16 \t 2.2 \t 1.$ Domain y continuous *2.2.4. Boundary conditions*

17 17 Boundary conditions must be applied at all exterior boundaries of the computational domain to define ₁₈ the flow. A fully-developed, laminar, axial-velocity-profile is applied at the inlet given by

$$
\vec{V} = \vec{V}_{\text{Max}} \left(1 - \frac{r^2}{R_0^2} \right),\tag{7}
$$

where the variable *r* is the radial coordinate measured from the centreline and
$$
R_o
$$
 is the wall radius of 1.6 mm. \vec{V}_{Max} is the maximum centreline velocity given a value which corresponds to a blood volume flow rate of 55 ml/min. This corresponds to the mean flow rate in the human LAD coronary artery under resting conditions [33]. The decision to use these mean flow properties is based on the desire to predict constant typical values of the WSS acting in the stented artery. A fixed static pressure of zero N/m² is applied at the outlet of the domain. The no-slip boundary condition is applied on all surfaces representative of the artery wall and the stent struts.

30 30 *2.2.5. Solution strategy*

 $\mu_{9} = 0.056 \text{ N} \cdot \text{s} \cdot \text{m}^{-2}$, $\lambda = 3.31 \text{ s}$,
 $\mu_{\infty} = 0.00345 \text{ N} \cdot \text{s} \cdot \text{m}^{-2}$, $q = 0.375$.

e commercial software code ANSYS CFX Version 12 (Canonsburg, PA, USA) is used to solve

verviring equations in t 31 The CFD code uses a coupled solver, solving the governing flow equations as a single system at 31 ₃₂ each timestep. This solution approach uses a fully implicit discretisation of the equations at any given ₃₂ ₃₃ timestep. The timestep behaves like an 'acceleration parameter', to guide the approximate solutions in a ₃₃ ₃₄ physically based manner to a steady-state solution. The convergence criterion for the maximum velocity ₃₄ as and density residuals is set at 10^{-5} . The discretised equations are solved using parallel processing with $_{35}$ $_{36}$ a MeTiS multilevel weighted k-way partitioning algorithm. The computations are conducted on a HP $_{36}$ 37 xw6400 64-bit workstation with a quad Intel (Xeon) 2 GHz processor with 6 GB of RAM and 80 GB 37 38 hard disk space. 38

29 29

39 39 $\frac{40}{40}$ 2.0. Fost processing 40 *2.3. Post-processing*

41 41 *2.3.1. Introduction*

⁴² The post-processing of the predicted WSS vectors, conducted in Tecplot 360 2008 (Bellevue, WA, ⁴² ⁴³ USA), produces the magnitudes of the WSS, WSSG and WSSAG on the arterial surfaces. The methodol-⁴⁴ ogy employed to calculate the distribution of these variables is described below along with a description ⁴⁴ 45 45 of the statistical analysis.46 46

1 1 *2.3.2. Wall shear stress*

² The dot product of the unit normal vector of a surface denoted \vec{l} and the viscous stress tensor denoted ² ³ $\vec{\vec{\tau}}_{ij}$ yields the WSS vector and in Cartesian coordinates is written ³

$$
\vec{i} \cdot \vec{\vec{\tau}}_{ij} = \vec{\tau}_{w_{xyz}} = \tau_{w,x}\vec{i} + \tau_{w,y}\vec{j} + \tau_{w,z}\vec{k}.\tag{8}
$$

7 and 2008 ⁸ The magnitude of the WSS vector is equal to the viscous shear stress acting on the surface and calculated ⁸ $9 - 9$ as

 12 and 12 and 12 and 12 and 12 and 12

4 4

$$
WSS = (\tau_{w,x}^2 + \tau_{w,y}^2 + \tau_{w,z}^2)^{1/2}.
$$
\n(9) 10

13 Physiologic arterial $WSS > 1.5$ N/m² has been shown to promote endothelial quiescence in the artery 13 14 [31] and to reduce IH [4]. Numerous studies have shown positive correlation between IH and sites with 14 15 15 lower than normal physiologic WSS values [11,21,27,40,41].

16 and 16 16 *2.3.3. Wall shear stress gradient*

Enginement of the Vi 3.5 vector is equal of the Viscota since as samely on an stanche and canceral

WSS = $(\tau_{w,2}^2 + \tau_{w,3}^2)^{1/2}$.

ysiologic arterial WSS > 1.5 N/m² has been shown to promote endothelial quiescence 17 2.0.0. We show shows given by 17 18 18 The WSSG is a measure of the spatial rate of change of the magnitude of the WSS vector. The WSSG ¹⁹ is obtained by first calculating the gradient of the WSS vector in Cartesian coordinates, which results in a ₂₀ nine component tensor. In order to calculate the magnitude of the WSSG acting on the artery surface, the ₂₀ 21 WSSG tensor must be transformed from the Cartesian coordinate system to a local coordinate system by $_{21}$ ₂₂ a standard component-wise tensor transformation explained in detail elsewhere [20]. The three mutually-23 orthogonal axes of a local coordinate system are taken as m , the WSS direction, n , tangential to the 23 24 surface and normal to m, and l, the surface normal direction. The components of the local WSSG tensor $_{24}$ $_{25}$ that act tangentially to the surface are then $_{25}$

$$
\vec{\nabla}\vec{\tau}_{w_{mnl}} = \begin{pmatrix}\n\frac{\partial \tau_{w,m}}{\partial m} & \frac{\partial \tau_{w,m}}{\partial n} \\
\frac{\partial \tau_{w,n}}{\partial m} & \frac{\partial \tau_{w,n}}{\partial n}\n\end{pmatrix}.
$$
\n(10)
$$
27
$$
\n28

29 29

30 The diagonal components $\partial \tau_{w,m}/\partial m$ and $\partial \tau_{w,n}/\partial n$ create tension between adjacent endothelial 30 31 31 cells which line the artery. This causes widening and shrinking of the cellular gaps. The components 32 $\partial \tau_{w,m}/\partial n$ and $\partial \tau_{w,n}/\partial m$ cause relative movement of adjacent cells. Lei et al. [29] suggest that the 32 33 components causing the intracellular tension are the most important with respect to IH and, with this in 33 34 34 mind, the magnitude of the WSSG is calculated as

35 35

$$
WSSG = \left[\left(\frac{\partial \tau_{w,m}}{\partial m} \right)^2 + \left(\frac{\partial \tau_{w,n}}{\partial n} \right)^2 \right]^{1/2}.
$$
\n³⁶
\n38\n
$$
(11) \quad \frac{37}{37}
$$

39 39 *2.3.4. Wall shear stress angle gradient*

⁴⁰ The directional changes in the WSS vector, not accounted for in the WSSG calculation, may also lead ⁴⁰ ⁴¹ to IH [30]. A mesh independent WSS directional parameter is formulated by using the gradient operator. ⁴¹ ⁴² Using the WSS vector at the node of interest as a reference, angular differences given by 43 43

44
\n45
$$
\phi = \pm \cos^{-1}\left(\frac{\vec{\tau}_o \times \vec{\tau}_r}{|\vec{\tau}_o| \times |\vec{\tau}_r|}\right),
$$
 -180 $\phi \le 180,$ (12) 45
\n46

1 are assigned to the neighbouring nodes where $\vec{\tau}_o$ is the WSS vector at the node of interest and $\vec{\tau}_r$ is the ² WSS vector at the neighbour node. Taking the spatial gradient of these angular differences at the node² 3 3 of interest yields the wall shear stress angle gradient (WSSAG) vector written

4 4

7 and 2008 and 2008

$$
\overline{WSSAG} = \frac{\partial \phi}{\partial x}\vec{i} + \frac{\partial \phi}{\partial y}\vec{j} + \frac{\partial \phi}{\partial z}\vec{k}.
$$
\n(13)

 $8₈$ The magnitude of the WSSAG is calculated as $8₈$

$$
WSSAG = \left(\left(\frac{\partial \phi}{\partial x} \right)^2 + \left(\frac{\partial \phi}{\partial y} \right)^2 + \left(\frac{\partial \phi}{\partial z} \right)^2 \right)^{1/2}
$$

 12 and 12 and 12 and 12 and 12 and 12 13 and is the maximum rate of change of WSS angle with respect to unit space. Endothelial cells have been 13 ¹⁴ shown to align themselves in the WSS direction [31], and the WSSAG can therefore be used to identify ¹⁴ 15 15 sites where gaps may be produced between endothelial cells which may lead to IH.

16 16 *2.3.5. Statistical analysis*

¹⁷ As the CFD solver employs a vertex-centred finite volume scheme, the WSS, WSSG and WSSAG are ¹⁷ ¹⁸ computed at the vertices of the surface faces. Face-averaged values of these variables are then calculated ¹⁸ ¹⁹ for each face as ¹⁹ 20 20

$$
\phi_j = \frac{\sum_{i=1}^n \phi_i}{n} \tag{15}
$$

24 where ϕ is the variable, j is the face number and the summation is over the n vertices attached to the 24 $_{25}$ face j. The area distribution of each face-averaged variable is visualised using histograms by displaying $_{25}$ ₂₆ the amount of area contained between specific intervals of the variable value. In addition to this qual-₂₇ itative technique, the area-weighted mean, standard deviation and kurtosis of the distribution of each ₂₇ 28 28 variable are also calculated for quantitative analysis.

e magnitude of the WSSAG is calculated as

WSSAG = $\left(\frac{\partial \phi}{\partial x}\right)^2 + \left(\frac{\partial \phi}{\partial y}\right)^2 + \left(\frac{\partial \phi}{\partial z}\right)^2\right)^{1/2}$

is the maximum rate of change of WSS angle with respect to unit space. Endothelial cells have been to identi ₂₉ The area-weighted mean represents the average value of the variable in the stented region of the ₂₉ ₃₀ artery. The standard deviation provides a measure of the typical difference between variable values and ₃₀ 31 the mean value. A high standard deviation signifies the existence of areas where the variable value may 31 32 be much higher or lower than the mean. The kurtosis provides an extra measure of the variable value 32 ₃₃ deviations from the mean in the distribution and is equal to the average of the tesseracts of the deviations ₃₃ $_{34}$ normalised by the tesseract of the standard deviation. A high kurtosis can therefore signify that the value $_{34}$ ₃₅ of the standard deviation is being driven upwards by a small number of data points where the deviation ₃₅ $_{36}$ is very high. The lowest kurtosis value possible is 1. This would happen if all variable values were at one $_{36}$ ₃₇ standard deviation from the mean. In general, the kurtosis quantifies the influence of data points with ₃₇ ₃₈ very high deviations on the standard deviation. ³⁸

39 39

40 40 $\frac{41}{41}$ at the district of $\frac{41}{41}$ **3. Results**

$42 \t 21 \t 1 \t 1$ 43 43 *3.1. Introduction*

⁴⁴ CFD results are presented for the models of the LAD artery implanted separately with the three dif-⁴⁵ ferent stents. The next three subsections contain the results for the calculated WSS, WSSG and WSSAG ⁴⁵ 46 46

(14)

¹ variables, respectively. The results are displayed in histogram form for each variable. The histograms ¹ ² display the amount of arterial tissue area contained between specific intervals of each face-averaged ² 3 3 variable. The area in the histograms is normalised by the total area analysed, which is the tissue area ⁴ confined within the axial limits of the stent. Greyscale contour plots are also provided to help visualise ⁴ ⁵ the predicted variables on the artery wall. The distribution of the three variables is quantified by the ⁵ 6 6 statistical analysis given with the histograms.

7 and 2008 and 2008

8 8 *3.2. Wall shear stress*

9 9 ₁₀ The WSS results are presented in Figs 4 and 5. The WSS for the fully-developed flow near the inlet of ₁₀ the unstented section is constant with a value of 1.21 N/m² for all stents. The statistical analysis of the $_{11}$ ¹² distribution of WSS shown in Fig. 4 reveals that the mean WSS is similar for the PS and GR-II stents ¹² ¹³ with values of 0.753 and 0.757 N/m², respectively. The Bx stent has a significantly lower mean value $\frac{13}{13}$ 14 of 0.516 N/m². Figure 5 shows large amounts of area around the Bx stent struts where the WSS is low $_{14}$ $15 \, (\leq 0.5 \, \text{N/m}^2)$ which explains this comparatively lower mean value.

16 16 The standard deviation is approximately 32% higher for the GR-II stent compared to the PS and Bx 17 stents. This highlights the wider distribution of arterial WSS values visible in Fig. 4. This is due to the 17 ₁₈ high WSS values at the peaks of the prolapse and lower values in the troughs around the GR-II stent ₁₈ ¹⁹ struts shown in Fig. 5. Data in Table 1 shows that the GR-II stent allows more tissue to prolapse into $\frac{19}{19}$ ₂₀ the artery than the other stents. The higher standard deviation quantifies a haemodynamic effect of the ₂₀ $_{21}$ anger volume of protapsing ussue with the OK-II stent. larger volume of prolapsing tissue with the GR-II stent.

22 The kurtosis is 60% and 66% higher for the PS stent compared to the GR-II and Bx stents, respectively, $\frac{22}{2}$ ₂₃ indicating the standard deviation value is influenced more so by small arterial tissue areas where the ₂₃ value of the WSS deviates greatly from the mean. Figure 5 shows these areas of tissue at the peaks of the $_{24}$ ₂₅ prolapsing tissue at the articulation site. The thinner struts of the PS stent have lead to a more uniform ₂₅ distribution of WSS in each of the closed cells in comparison to the wider spread of values in the closed $_{26}$

²⁷ creating large areas of low WSS around its struts that only briefly recover to values above 1 N/m² in 27 ²⁸ the cell centres. The mean WSS values are similar, but the analysis uncovers the distinction between $\frac{28}{3}$ ²⁹ the PS and GR-II stents. The PS stent has a reasonably uniform WSS distribution over the majority of $\frac{29}{29}$ ³⁰ the stented region with small high-deviation regions at the articulation and very close to the struts. In $\frac{30}{21}$ 31 contrast the GR-II stent has larger areas of low WSS near the struts which spatially change quickly to $\frac{32}{32}$ $\frac{33}{33}$ $\frac{115}{33}$ and $\frac{33}{33}$ high values.

 The threshold method of comparison shows 22.8%, 32.2% and 50.0% of the stented region exposed to 34 WSS in the range of $0.0-0.5$ N/m² for the PS, GR-II and Bx stents, respectively. The threshold method $_{35}$ therefore favours the PS stent followed by the GR-II and then Bx stent, which is in agreement with 36 the analysis above. However, the threshold method does not identify the complex stent-design-related 37 38 distribution of WSS on the artery wall.

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40 *3.3. Wall shear stress gradient*

⁴² Figures 6 and 7 contain the results for the WSSG. The mean value for the GR-II stent is 74% higher ⁴² ⁴³ than that for the PS stent and 55% higher than that for the Bx stent. The GR-II stent struts which traverse 43 ⁴⁴ the flow create proximal and distal areas of low WSS. The WSS value then quickly increases in the axial ⁴⁴ ⁴⁵ direction as discussed in Section 3.2. This creates the large WSSGs between the struts which are visible ⁴⁵ 46

Wall Shear Stress Gradient [N/m³]: 200 500 1000 2000 3000 40
40 41 Fig. 7. Contour plots of the WSSG on the arterial tissue in the stented arteries.

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is larger areas with high WSSSG values of 1000-2000 M/m² as shown in the histogram in Fig. 6.
Increasing the state and distinguish of 1000-2000 M/m² as shown in the histogram in Fig. 7. This demonstrates how the histog ¹ The standard deviation of the WSSG for the GR-II stent is 858.8 N/m³, 12% and 38% higher than the ¹ 2 2 PS and Bx stents, respectively. This signifies large spatial fluctuations in the WSSG acting on the arterial ³ tissue. Figure 7 illustrates this effect, with large regions of high WSSG ($>$ 3000 N/m³) on the proximal ³ 4 side of the smallest inter-strut regions and low WSSG ($\langle 200 \text{ N/m}^3$) in the middle of the largest inter-5 5 strut regions of the GR-II stent. The standard deviation is 24% higher for the PS stent when compared to 6 6 the Bx stent. However, the higher kurtosis value for the PS stent indicates that the standard deviation is ⁷ influenced by small areas of tissue with very high WSSG values ($>$ 3000 N/m³). In contrast the Bx stent ⁷ 8 has larger areas with high WSSG values of 1000–2000 N/m³ as shown in the histogram in Fig. 6 and 8 9 the contour plot in Fig. 7. This demonstrates how the histograms, statistical measures and contour plots 9 10 compliment each other to reveal the detail of the variable distributions produced by the different stent 10 11 designs. 11 and 11 and 12 and 1 12 The large WSSGs produced in the inter-strut regions of the GR-II stent have implicated it as the worst 12 13 performing stent with regard to this variable. Comparing Bx and PS stents, the higher mean and lower 13 14 kurtosis values place the Bx stent second, with the PS again performing the best. Using thresholds, the 14 15 15 PS stent has 0.6% of its stented region with *WSSG* < 200 N/m³ compared to 1.2% for the GR-II and Bx 16 16 stents. This negligible amount of tissue below the threshold value is insufficient to distinguish between designs.

17 the stents. This is a clear demonstration of the weakness of the threshold method to fully analyse the 17 18 18 predicted variable on the artery wall. 19 19

20 20 *3.4. Wall shear stress angle gradient*

 21 To the authors' knowledge no upper threshold has ever been defined in the literature for the WSSAG, 21 ²² such that an artery with WSSAG values above this threshold can be considered to be at an elevated risk $\frac{22}{100}$ ²³ for IH. Indeed, the authors' consider this to be the first time this variable has been predicted in a model $\frac{^{23}}{^{24}}$ ²⁴ of a stented artery. The performance of the stents with regard to this variable can easily be evaluated $\frac{24}{\epsilon}$ 25 using the analysis technique applied for the other variables.

²⁶ 26 26 26 26 The results for the WSSAG are presented in Figs 8 and 9. A semi-logarithmic scale is used on the ²⁷ histogram in Fig. 8 to accommodate the large range of this variable. The mean WSSAG value for the $\frac{27}{20}$ ²⁸ GR-II stent is 33.4 rad/mm, 163% higher than that for the PS stent and 42% higher than that for the Bx ²⁹ stent. The flow traversing struts of the GR-II stent have created large areas of separation and reattachment ³⁰ proximal and distal to the struts creating very high WSSAG values (>100 rad/mm). These regions of $\frac{30}{\epsilon}$ $\frac{31}{20}$ high WSSAG are also visible around the S-connectors of the Bx stent as shown in Fig. 9. These effects $\frac{31}{20}$ $\frac{32}{2}$ are quantifiable from the comparison of the mean values.

³³ The standard deviation is the highest for the GR-II stent, 79% higher than the PS stent and 88% higher ³³ ³⁴ than the Bx stent. Figure 9 shows large areas of low WSSAG (<5 rad/mm) between the large inter-³⁵ strut regions. This effect coupled with the very high WSSAG near the GR-II struts discussed above is $\frac{35}{20}$ ³⁶ highlighted by the high standard deviation value.³⁶

³⁷ The standard deviations are similar for the PS and Bx stents. However, the kurtosis provides an extra³⁷ $\frac{38}{10}$ measure of contrast of the WSSAG distribution between the two stents. The higher kurtosis for the PS $\frac{38}{10}$ ³⁹ stent signifies small areas of tissue under very high WSSAG values (>100 rad/mm) influencing the ³⁹ ⁴⁰ standard deviation, whereas the lower kurtosis for the Bx stent signifies larger amounts of tissue in the ⁴¹ high range of WSSAG (20–100 rad/mm). This distribution is displayed in the histogram in Fig. 8.

⁴² The analysis shows that the GR-II stent is the worst stent with more area under the high WSSAG ⁴² ⁴³ values. This is followed by the Bx stent and finally the PS stent which performs the best. The high ⁴³ ⁴⁴ kurtosis values with this variable compared to other variables reflects that the WSSAG is generally low ⁴⁴ ⁴⁵ in the stented region but has high magnitudes in regions of highly disturbed flow. 46 46

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⁴⁵ stented artery. A CFD analysis is conducted to predict the WSS vectors acting on the arterial tissue in the

essement. The threshold medd only slightly dissinguishes between the stars are the systement of the SNS clearly demonstrates that the GR-II is the poorest performing stent with regard to rable. This is a prime example of t ¹ stented artery. The components of the WSS vectors are then post-processed to calculate the magnitude of ¹ ² the predicted WSS, WSSG and WSSAG. A statistical analysis is then conducted to obtain the maximum ² 3 3 information from the variables. ⁴ Both the proposed analysis technique and the threshold method identify the Bx as being the less ⁴ ⁵ attractive of the stents with regard to WSS. However, using the histograms as a visual aid and quantifying ⁵ 6 6 the results using the statistical analysis much more information is revealed about the distribution of ⁷ the variable. Examination of the WSSG variable reveals a weakness in the threshold method of stent ⁷ 8 assessment. The threshold method only slightly distinguishes between the stents. The statistical analysis 8 9 of the WSSG clearly demonstrates that the GR-II is the poorest performing stent with regard to this 9 ¹⁰ variable. This is a prime example of the benefit of the proposed methodology. There is no threshold ¹⁰ 11 method comparison to be made for the WSSAG, as no threshold has been defined in the literature. The 11 12 statistical analysis has once again proved its robustness in easily identifying how the features of the 12 ¹³ stent influence the distribution of this variable in the stented artery. The GR-II stent is again the poorest ¹³ 14 performing stent. ¹⁵ Each of the variables employed in this methodology highlights the different adverse haemodynamic¹⁵ ¹⁶ features which could lead to IH development. If stent performance were based solely on WSS as is ¹⁶ 17 sometimes the case [1], the Bx would be implicated as the worst stent. However, the multi-variable 17 ¹⁸ approach employed in this work seems to favour the stents in the order of PS, Bx and GR-II. This also ¹⁸ ¹⁹ correlates with the order of stent restenosis rates indicating that adverse haemodynamics may be partly ¹⁹ 20 20 responsible for the *in vivo* performance of these stents. This methodology is applicable to BMSs, DESs 21 and any future stents that alter the haemodynamics of the artery after implantation. The results identify 21 22 22 one of the key stimuli for IH in the implanted artery and as such are important for all types of stent. 23 23 Limitations include the assumptions of fully-developed, steady, laminar flow, a rigid stent and arterial 24 24 wall and an idealised model of prolapsing tissue. Since actual tissue prolapse *in vivo* is likely be patient 25 25 specific, an option to omit the idealised prolapse model in order to investigate the effect of the stent 26 26 geometry alone could be considered. However, the shape of the prolapsing tissue is determined by the ²⁷ stent geometry and as such, is in itself a feature of the stent. Since tissue prolapse is known to occur ²⁷ 28 28 *in vivo* [16], the inclusion of the idealised prolapse model should produce more realistic results. In the 29 29 analysis of the results, the kurtosis is used to indicate the existence of small areas exposed to highly 30 30 abnormal variable values. With regard to stent performance, the assumption is made that large areas 31 of moderately abnormal variable values are worse than small areas of highly abnormal variable values. ³¹ 32 This assumption is based on the fact that tissue growth generally occurs where necessary to restore 32 33 the variables back to normal values [27,12]. It is likely that the large areas with moderately abnormal ³³ 34 34 variable values would require a greater volume of tissue growth to be restored back to normal values, 35 35 and are therefore more likely to result in restenosis. 36 36 The main objective of this work is to introduce a more complete method of post-processing and 37 analysing results obtained from a simulation performed on physiologically realistic models of the stented 37 38 artery. The methodology presented here provides a novel, accurate, and efficient way to assess and com-
38 39 pare different stents based on the haemodynamic WSS-based variables and this should assist in stent 39 40 40 design in the future. 41 41 42 42 43 References 43 **References**

44 44 ⁴⁵ [1] R. Balossino, F. Gervaso, F. Migliavacca et al., Effects of different stent designs on local hemodynamics in stented arteries, 46 46

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